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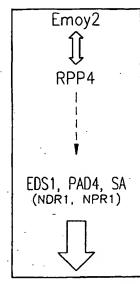
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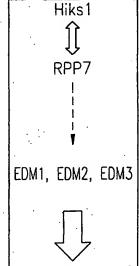
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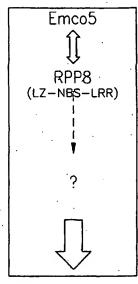
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(54) Title: PLANT GENES, THE EXPRESSION OF WHICH ARE ALTERED BY PATHOGEN INFECTION

#### RPP-DEPENDENT DEFENSE PATHWAYS







RESISTANCE

(57) Abstract: Methods to identify genes, the expression of which is altered in response to pathogen infection, are provided, as well as the genes identified thereby.

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# PLANT GENES, THE EXPRESSION OF WHICH ARE ALTERED BY PATHOGEN INFECTION

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#### **Cross-Reference to Related Applications**

This application is a continuation-in-part application of U.S. application Serial No. 60/232,778, filed on September 15, 2000, and of U.S. application Serial No. 60/300,183, filed on June 22, 2001, the disclosures of which are incorporated by reference herein.

#### Field of the Invention

The present invention generally relates to the field of plant molecular biology, and more specifically to the regulation of gene expression in plants in response to stress, e.g., to pathogen exposure.

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#### **Background of the Invention**

Plants are capable of activating a large array of defense mechanisms in response to pathogen attack. A crucial factor determining the success of these mechanisms is the speed of their activation. Consequently, there is considerable interest in understanding how plants recognize pathogen attack and control expression of defense mechanisms.

Some potential pathogens trigger a very rapid resistance response called gene-for-gene resistance. This occurs when the pathogen carries an avirulence (avr) gene that triggers specific recognition by a corresponding host resistance (R) gene. R gene specificity is generally quite narrow, in most cases only pathogens carrying a particular avr gene are recognized. Recognition is thought to be mediated by ligand-receptor binding. R genes have been studied extensively in recent years. For a review of R genes, see Ellis et al. (1998); Jones et al. (1997); and Ronald (1998).

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One of the defense mechanisms triggered by gene-for-gene resistance is programmed cell death at the infection site. This is called the hypersensitive response, or HR. Pathogens that induce the HR, or cause cell death by other means, activate a systemic resistance response called systemic acquired resistance (SAR). During SAR, levels of salicylic acid (SA) rise throughout the plant, defense genes such as pathogenesis related (PR) genes are expressed, and the plant becomes more resistant to pathogen attack. SA is a crucial component

of this response. Plants that cannot accumulate SA due to the presence of a transgene that encodes an SA-degrading enzyme (nahG), develop a HR in response to challenge by avirulent pathogens, but do not exhibit systemic expression of defense genes and do not develop resistance to subsequent pathogen attack (Ryals et al., 1996). The nature of the systemic signal that triggers SAR is a subject of debate (Shulaev et al., 1995; Vernooji et al., 1994). SA clearly moves from the site of the HR to other parts of the plant, but if this is the signal, it must be effective at extremely low concentration (Willits et al., 1998).

SAR is quite similar to some reactions that occur locally in response to attack by virulent (those that cause disease) or avirulent (those that trigger genefor-gene resistance) pathogens. In general, activation of defense gene expression occurs more slowly in response to virulent pathogens than in response to avirulent pathogens. Some pathogens trigger expression of defense genes through a different signaling pathway that requires components of the jasmonic acid (JA) and ethylene signaling pathways (Creelman et al., 1997).

One approach to understanding the signal transduction networks that control defense mechanisms is to use genetic methods to identify signaling components and determine their roles within the network. Considerable progress has been made using this approach in *Arabidopsis*-pathogen model systems.

#### R gene signal transduction

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Genes such as NDR1 and EDS1, as well as DND1 and the lesion-mimic genes, likely act in signal transduction pathways downstream from R-avr recognition. NDR1 and EDS1 are required for gene-for-gene mediated resistance to avirulent strains of the bacterial pathogen Pseudomonas syringae and the oomycete pathogen Peronospora parasitica. Curiously, ndr1 mutants are susceptible to one set of avirulent pathogens, whereas eds1 mutants are susceptible to a non-overlapping set (Aarts et el., 1998). The five cloned R genes that require EDS1 all belong to the subset of the nucleotide binding site-leucine rich repeat (NBS-LRR) class of R genes that contain sequences similar to the cytoplasmic domains of Drosophila Toll and mammalian interleukin 1 transmembrane receptors (TIR-NBS-LRR). The two genes that require NDR1 belong to the leucine-zipper (LZ-NBS-LRR) subclass of NBS-LRR genes. There is another LZ-NBS-LRR gene, RPP8, that does not require EDS1 or NDR1, so

the correlation between R gene structure and requirement for EDSI or NDRI is not perfect. Nevertheless, these results show that R genes differ in their requirements for downstream factors and that these differences are correlated with R gene structural type.

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NDR1 encodes a protein with two predicted transmembrane domains (Century et al. 1997). RPM1, which requires NDR1 to mediate resistance, is membrane-associated, despite the fact that its primary sequence does not include any likely membrane-integral stretches (Boyes et al., 1998). It is possible that part of the function of NDR1 is to hold R proteins close to the membrane. EDS1 encodes a protein with blocks of homology to triacyl glycerol lipases (Falk et al., 1999). The significance of this homology is not known, but it is tempting to speculate that EDS1 is involved in synthesis or degradation of a signal molecule. EDS1 expression is inducible by SA and pathogen infection, suggesting that EDS1 may be involved in signal amplification (Falk et al., 1999).

It has been extremely difficult to isolate mutations in genes other than the R genes that are required for gene-for-gene resistance. A selection procedure was devised (McNellis et al., 1998) on the basis of precisely controlled inducible expression of the avr gene avrRpt2 in plants carrying the corresponding resistance gene RPS2. Expression of avrRpt2 in this background is lethal, as it triggers a systemic HR. It is now possible to select for mutants with subtle defects in gene-for-gene signaling by requiring growth on a concentration of inducer slightly higher than the lethal dose.

Putative plant receptor proteins encoded by RPP genes (recognition of P. parasitica) mediate specific recognition of Peronospora isolates and trigger defense reactions. Recently, McDowell et al. (2000) reported that two members of this class, RPP7 and RPP8 (the latter of which encodes a LZ-NBS-LRR type R protein) were not significantly suppressed by mutations in either EDS1 or NDR1, and that RPP7 resistance was also not compromised by mutations in EIN2, JAR1 or COI1, which affect ethylene or jasmonic acid signaling, or in coi1/npr1 or coi1/NahG backgrounds. The authors suggested that RPP7 initiates resistance through a novel signaling pathway that is independent of salicylic acid accumulation or jasmonic acid response components.

### SA-dependent signaling

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SA levels increase locally in response to pathogen attack, and systemically in response to the SAR-inducing signal. SA is necessary and sufficient for activation of PR gene expression and enhanced disease resistance. Physiological analyses and characterization of certain lesion-mimic mutants strongly suggest that there is a positive autoregulatory loop affecting SA concentrations (Shirasu et al., 1997; Hunt et al., 1997; Weymann et al., 1995). Several mutants with defects in SA signaling have been characterized. These include npr1, in which expression of PR genes in response to SA is blocked; cpr1, cpr5, and cpr6, which constitutively express PR genes; the npr1 suppressor ssi1; pad4, which has a defect in SA accumulation; and eds5, which has a defect in PR1 expression.

Expression of the defense genes *PR1*, *BG2*, and *PR5* in response to SA treatment requires a gene called *NPR1* or *NIM1*. Mutations in *npr1* abolish SAR, and cause enhanced susceptibility to infection by various pathogens (Cao et al., 1994; Delaney et al., 1995; Glazebrook et al., 1996; Shah et al., 1997). NPR1 appears to be a positive regulator of *PR* gene expression that acts downstream from SA. *NPR1* encodes a novel protein that contains ankyrin repeats (which are often involved in protein-protein interactions (Cao et al., 1997; Ryals et al., 1997), and that is localized to the nucleus in the presence of SA (Dong et al., 1998). Consequently, it is unlikely that NPR1 acts as a transcription factor to directly control *PR* gene expression, but its nuclear localization suggests that it may interact with such transcription factors.

PAD4 appears to act upstream from SA. In pad4 plants infected with a virulent P. syringae strain, SA levels, synthesis of the antimicrobial compound camalexin, and PR1 expression are all reduced (Zhou et al., 1998). SA is necessary, but not sufficient, for activation of camalexin synthesis (Zhou et al., 1998; Zhao et al., 1996). The camalexin defect in pad4 plants is reversible by exogenous SA (Zhou et al., 1998). Mutations in pad4 do not affect SA levels, camalexin synthesis, or PR1 when plants are infected with an avirulent P. syringae strain (Zhou et al., 1998). Taken together, these results suggest that PAD4 is required for signal amplification to activate the SA pathway in response to pathogens that do not elicit a strong defense response (Zhou et al., 1998).

#### JA-dependent signaling

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JA signaling affects diverse processes including fruit ripening, pollen development, root growth, and response to wounding (Creelman et al., 1997). The jar1 and coil mutants fail to respond to JA (Feys et al., 1994; Staswick et al., 1992). COII has been cloned, and found to encode protein containing leucine-rich repeats and a degenerate F-box motif (Xie et al., 1998). These features are characteristic of proteins that function in complexes that ubiquitinate protein targeted for degradation.

In the past few years it has become apparent that JA plays an important role in regulation of pathogen defenses. For example, the induction of the defensin gene *PDF1.2* after inoculation of *Arabidopsis* with the avirulent pathogen *Alternaria brassicicola* does not require SA or NPR1, but does require ethylene and JA signaling (Penninck et al., 1996).

SA signaling and JA signaling pathways are interconnected in complicated ways. Studies in other systems have shown that SA signaling and JA signaling are mutually inhibitory (Creelman et al., 1997; Harms et al., 1998). However, synthesis of camalexin in response to *P. syringae* infection is blocked in *nahG* (Zhou et al., 1998; Zhao et al., 1996) and *coil* (Glazebrook, 1999) plants, strongly suggesting that camalexin synthesis requires both SA and JA signaling.

#### Induced systemic resistance (ISR)

Some rhizosphere-associated bacteria promote disease resistance (van Loon et al., 1998). This phenomenon, called ISR, has been studied using *Pseudomonas fluorescens* strain WCS417r to colonize *Arabidopsis* roots (Pieterse et al., 1996). Colonized plants are more resistant to infection by the fungal pathogen *Fusarium oxysporum* f sp *raphani* and *P. syringae* (Pieterse et al., 1996). ISR occurs in *nahG* plants, indicating that it is not a SA-dependent phenomenon (Pieterse et al., 1996). Rather, ISR appears to be JA- and ethylene-dependent. The observation that ethylene can induce ISR in *jarl* mutants led to the hypothesis that ISR requires a JA signal followed by an ethylene signal (Pieterse et al., 1998). No changes in gene expression associated with ISR have been detected (Pieterse et al., 1998), suggesting that it is different from activation of *PDF1.2* expression by *A. brassicicola*.

Curiously, ISR requires NPR1 (Pieterse et al., 1996). This was unexpected in light of the fact that NPR1 was previously known to be involved only in SA-dependent processes and ISR is SA-independent. If the SA-dependent signal is received, NPR1 mediates a resistance response characterized by PR1 expression, whereas if the ISR signal is received, NPR1 mediates a different resistance response. It is difficult to imagine how this could occur, unless NPR1 is interacting with different 'adapter' molecules to mediate the different signals. The ankyrin repeats found in NPR1 could function in protein-protein interactions between NPR1 and adapter proteins. Identification of proteins that interact with NPR1, and characterization of plants with loss-of-function mutations affecting those proteins, would be very helpful for understanding how NPR1 acts in each pathway. It would also be worthwhile to determine if the ssil or cpr6 mutations suppress the ISR defect of npr1 mutants. Relevance to disease resistance

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Characterization of the effects of various mutations on resistance to different pathogens has revealed that there is considerable variation in the extent to which pathogens are affected by defense mechanisms. SAR is known to confer resistance to a wide array of pathogens, including bacteria, fungi, oomycetes, and viruses. JA signaling is important for limiting the growth of certain fungal pathogens. In *Arabidopsis*, the SA pathway mutants *npr1* and *pad4* show enhanced susceptibility to *P. syringae* and *P. parasitica* (Cao et al., 1994; Delaney et al., 1995; Shah et al., 1997; Zhou et al., 1998; Glazebrook et al., 1997).

Overexpression of rate-limiting defense response regulators may cause the signaling network to respond faster or more strongly to pathogen attack, thereby improving resistance: For example, overexpression of NPRI caused increased resistance to P. syringae and P. parasitica in a dosage dependent manner (Cao et al., 1998). Moreover, NPRI-overexpression had no obvious deleterious effects on plant growth, in contrast to mutations that lead to constitutive overexpression of defense responses, which generally cause dwarfism.

Thus, what is needed is the systematic identification of genes useful to confer resistance to pathogens.

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#### Summary of the Invention

The invention provides an isolated nucleic acid molecule (polynucleotide), e.g., DNA, comprising a plant or fungal nucleotide sequence, the expression of which is altered in response to stress, e.g., pathogen infection. For example, the invention provides a nucleic acid molecule comprising a plant or fungal nucleotide sequence comprising at least a portion of a key effector gene(s) responsible for host resistance to particular pathogens. To provide resistance or tolerance to a pathogen in a plant, this sequence may be overexpressed individually, in the sense or antisense orientation, or in combination with other sequences, to confer improved disease resistance or tolerance to a plant relative to a plant that does not comprise and/or express the sequence. The overexpression may be constitutive, or it may be preferable to express the effector gene(s) from an inducible promoter including a promoter which is responsive to external stimuli, such as chemical application, or to pathogen infection, e.g., so as to avoid possible deleterious effects on plant growth. In one embodiment of the invention, the promoter employed to express the nucleotide sequence of the invention may be one that mediates rapid and transient transcription after pathogen infection. Thus, the invention also provides an isolated nucleic acid molecule comprising a plant nucleotide sequence which alters, e.g., increases or decreases, the transcription of plant genes, e.g., open reading frames, in response to stress, e.g., pathogen infection.

As described herein, Arabidopsis plants of differing genotypes were infected with different strains of an oomycete, P. parasitica. RNA was isolated from each plant/pathogen pair and employed to prepare probes which were hybridized to a gene chip having nucleic acid sequences (probe sets) corresponding to approximately 8,200 Arabidopsis genes. Genes were then identified that were upregulated or downregulated in response to infection, including genes that were dependent on RPP7 or RPP8, which act via unconventional signaling cascades and are not dependent on defense regulators such as EDS1, NDR1, PAD4, NPR1, RAR1, PBS3 or salicyclic acid (SA). Among the genes showing strong Peronospora-induced expression changes, clusters of genes were identified that were specifically upregulated by RPP7 or RPP8, or both. In particular in one analysis, the expression of 184 genes (SEQ ID NOs:1-211 and 792) related to 217 probe sets was upregulated by either RPP7 or RPP8, or both, while in another analysis the expression of genes related to 194 probe sets (SEQ ID NOs:212-399 and 793) was

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upregulated by either RPP7 or RPP8, or both. Further, as described below, promoters of genes that were rapidly and transiently transcribed after P. parasitica infection and were RPP7/8-dependent were significantly enriched with both novel sequence motifs and potential binding sites of known transcription factors. Specifically, the promoters were significantly enriched with two novel sequence motifs (referred to as "motif 1" and "motif 2"), one of which was similar to binding sites for Myb transcription factors, sequence motifs related to WRKY binding sites, and two other novel sequence motifs ("motif 3" and "motif 4"). Moreover, comparisons between expression signatures of wild type plants as well as the RPP7 loss of function mutants, and edm1, edm2 and edm3 mutants, allow for predictions regarding the RPP7 pathway hierarchy.

In addition, more than 200 genes (SEQ ID NOs:75, 214, 228, 301, 339, 400-684, 792-795) were identified that were specifically controlled by the RPP4-dependent pathway, which mediates resistance of the Arabidopsis ecotype Col-0 to the Peronospora isolate Emoy2. According to their response to SA, an important mediator of local and systemic defense responses, and the protein biosynthesis inhibitor cycloheximide (CHX), these genes were further subcategorized into immediate early and secondary response genes. A set of immediate early SA responsive genes (SEQ ID NOs:150, 159, 117, 126, 208, 428, 426, 436, 430, 434, 478, 641, 609, 615, and 526) was found to consist exclusively of regulatory genes. Such genes are likely controlled by transcription factors acting closely downstream of SA. One highly conserved motif in the promoters of these immediate early genes was similar to binding sites of WRKY transcription factors (SEQ ID NOs:757-765). Other conserved promoter motifs appeared to be novel and may facilitate the cloning of their cognate transcription factors. A cluster of SA/CHX super-induced genes was found to show elevated expression ground states in the npr1 mutant (SEQ ID NOs:214, 621, 71, 439, 78, 80, 264, 265, 613, 583, 594, 616, 355, 503, 168, 447 and 551) suggesting an NPR1-dependent derepression mechanism in the control of some SA responsive immediate early genes.

Thus, the invention provides an isolated nucleic acid molecule comprising a plant or fungal nucleotide sequence encoding a polypeptide which is substantially similar to an *Arabidopsis* polypeptide encoded by a gene comprising an open reading frame, the expression of which is altered in response to a pathogen that induces a response mediated by *R* genes, including pathogens such as bacteria, fungi, oomycetes, viruses, nematodes and insects, e.g., aphids (see Hammond-Kosack and Jones (1997), which is specifically incorporated by

reference herein). For example, the plant or fungal nucleotide sequence encodes a polypeptide that is substantially similar to an Arabidopsis polypeptide encoded by a gene comprising an open reading frame, the expression of which is altered in response to infection by an oomycete such as Peronospora, and is RPP4-, RPP7-, and/or RPP8-dependent. These sequences can be identified by employing an array of nucleic acid samples, e.g., a plurality of oligonucleotides, each plurality corresponding to a different plant gene, on a solid substrate, e.g., a DNA chip, and probes corresponding to nucleic acid from pathogen-infected (e.g., wild-type) plant cells and to nucleic acid from uninfected plant cells or 10 plant cells having a mutation which alters the response to pathogen infection. Thus, genes, the expression of which are altered by pathogen infection, can be systematically identified. Preferably, the nucleotide sequence is from plant DNA, either a dicot or a monocot, which encodes a polypeptide that is substantially similar to an Arabidopsis polypeptide encoded by an open reading 15 frame comprising any one of SEQ ID NOs:1-684 and 792-795, or the complement thereof. More preferably, the nucleotide sequence is from plant DNA that is substantially similar to an Arabidopsis nucleic acid segment having any one of SEQ ID NOs: 1-684 and 792-795, or the complement thereof. The term "substantially similar", when used herein with respect to a polypeptide 20 means a polypeptide corresponding to a reference polypeptide, wherein the polypeptide has substantially the same structure and function as the reference polypeptide, e.g., where only changes in amino acid sequence are those which do not affect the polypeptide function. When used for a polypeptide or an amino acid sequence, the percentage of identity between the substantially similar and the reference polypeptide or amino acid sequence is at least 65%, 66%, 67%, 68%, 69%, 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, up to at least 99%, where the reference polypeptide is an Arabidopsis polypeptide encoded by an open reading frame comprising any one of SEQ ID NOs:1-684 and 792-795, or the complement thereof. However, the percent of identity between the substantially similar and the reference polypeptide may be less than 65% as long as the two polypeptides have the same or similar function, e.g., catalyze the same or similar reaction. Another indication that two polypeptides are substantially similar to

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each other is that an agent, e.g., an antibody, which specifically binds to one of the polypeptides, specifically binds to the other.

In its broadest sense, the term "substantially similar", when used herein with respect to a nucleotide sequence, means a nucleotide sequence corresponding to a reference nucleotide sequence, wherein the corresponding sequence is from a gene that encodes a polypeptide having substantially the same structure and function as the polypeptide encoded by a gene comprising the reference nucleotide sequence. The term "substantially similar" is specifically intended to include nucleotide sequences wherein the sequence has been modified to optimize expression in particular cells. The percentage of identity between the substantially similar nucleotide sequence and the reference nucleotide sequence is at least 65%, 66%, 67%, 68%, 69%, 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, up to at least 99%, wherein the reference sequence is preferably any one of SEQ ID NOs: 1-684 and 792-795, or the complement thereof. Sequence comparisons may be carried out using a Smith-Waterman sequence alignment algorithm (see e.g., Waterman (1995) or http://www hto.usc.edu/software/seqaln/index.html). The localS program, version 1.16, is preferably used with following parameters: match: 1, mismatch penalty: 0.33, open-gap penalty: 2, extended-gap penalty: 2. Further, a nucleotide sequence that is "substantially similar" to a reference nucleotide sequence hybridizes to the reference nucleotide sequence in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 2X SSC, 0.1% SDS at 50°C, more desirably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 1X SSC, 0.1% SDS at 50°C, more desirably still in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.5X SSC, 0.1% SDS at 50°C, preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.1X SSC. 0.1% SDS at 50°C, more preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 65°C.

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Hence, the isolated nucleic acid molecules of the invention also include the orthologs of the *Arabidopsis* sequences disclosed herein, i.e., the corresponding nucleic acid molecules in organisms other than *Arabidopsis*,

including, but not limited to, plants other than Arabidopsis, preferably cereal plants, e.g., corn, wheat or rice, as well as rye, turfgrass, sorghum, millet, sugarcane, soybean, barley, alfalfa, sunflower, canola, cotton, peanut, tobacco or sugarbeet, and in fungi. An ortholog is a gene from a different species that encodes a product having the same function as the product encoded by a gene from a reference organism. The encoded ortholog products likely have at least 70% amino acid sequence identity to each other. Hence, the invention includes an isolated nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide having at least 70% identity to a polypeptide encoded by one or more of the Arabidopsis sequences, although it is also envisioned that orthologous genes to those disclosed herein may encode a polypeptide with less than 70%, e.g., less than 65% amino acid sequence identity, but which polypeptide has the same or similar function. Databases such GenBank or one found at http://bioserver.myongjiac.kr/rjce.html (for rice) may be employed to identify sequences related to the Arabidopsis sequences, e.g., orthologs in cereal crops such as rice. Alternatively, recombinant DNA techniques such as hybridization or PCR may be employed to identify sequences related to the Arabidopsis sequences.

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Thus, the invention preferably includes an isolated nucleic acid molecule comprising a plant or fungal nucleotide sequence that encodes a polypeptide that has at least 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, and at least 99%, amino acid sequence identity to an Arabidopsis polypeptide encoded by an open reading frame comprising any one of SEQ ID NOs: 1-684 and 792-795, or a fragment (portion) thereof which encodes a polypeptide having substantially the same activity as a polypeptide encoded by an open reading frame comprising a corresponding sequence listed in SEQ ID NOs: 1-684 and 792-795. In one embodiment, the isolated nucleic acid molecule is not SEQ ID NOs: 1-684 and 792-795. The invention also provides anti-sense nucleic acid molecules corresponding to the open reading frames or genes identified as responsive to pathogen infection. Also provided are expression cassettes, e.g., recombinant vectors, and host cells, comprising the nucleic acid molecule of the invention in either sense or antisense orientation.

Also provided is an isolated nucleic acid molecule comprising a plant or fungal nucleotide sequence comprising a nucleic acid sequence having at least 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, and at least 99%, nucleic acid sequence similarity to an *Arabidopsis* open reading frame comprising any one of SEQ ID NOs: 1-684 and 792-795, the complement thereof, or a fragment (portion) thereof which encodes a polypeptide having substantially the same activity as a polypeptide encoded by an open reading frame comprising a corresponding sequence listed in SEQ ID NOs: 1-684 and 792-795.

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The nucleic acid molecules of the invention, their encoded polypeptides and compositions thereof, are useful to provide resistance to pathogens and as a diagnostic for the presence or absence of the pathogen by correlating the expression level or pattern of expression of one or more of the nucleic acid molecules of the invention or one or more of the polypeptides encoded thereby. As one embodiment of the invention includes isolated nucleic acid molecules that have increased expression in response to pathogen infection, the invention further provides compositions and methods for enhancing resistance to pathogen infection. The compositions of the invention include plant or fungal nucleotide sequences and the amino acid sequences for the polypeptides or partial-length polypeptides encoded thereby which are useful to provide tolerance or resistance to a plant to a pathogen, preferably by preventing or inhibiting pathogen infection. The resistance or tolerance may be accomplished by decreasing or eliminating expression of a plant gene necessary for pathogen infection and/or replication or by overexpressing a gene product that inhibits pathogen infection and/or replication. Methods of the invention involve stably transforming a plant with one or more of at least a portion of these nucleotide sequences which confer tolerance or resistance operably linked to a promoter capable of driving expression of that nucleotide sequence in a plant cell. By "portion" or "fragment", as it relates to a nucleic acid molecule, sequence or segment of the invention, when it is linked to other sequences for expression, is meant a sequence having at least 80 nucleotides, more preferably at least 150 nucleotides, and still more preferably at least 400 nucleotides. If not employed for expressing, a "portion" or "fragment" means at least 9, preferably 12, more

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preferably 15, even more preferably at least 20, consecutive nucleotides, e.g., probes and primers (oligonucleotides), corresponding to the nucleotide sequence of the nucleic acid molecules of the invention. By "resistant" is meant a plant which exhibits substantially no phenotypic changes as a consequence of infection with the pathogen. By "tolerant" is meant a plant which, although it may exhibit some phenotypic changes as a consequence of infection, does not have a substantially decreased reproductive capacity or substantially altered metabolism. Thus, some of the isolated nucleic acid molecules of the invention are useful in a method of combating a pathogen in an agricultural crop. The method comprises introducing to a plant, plant cell, or plant tissue an expression cassette comprising a nucleic acid molecule of the invention so as to yield a transformed differentiated plant, transformed cell or transformed tissue. Transformed cells or tissue can be regenerated to provide a transformed differentiated plant. The transformed differentiated plant preferably expresses the nucleic acid molecule in an amount that confers resistance to the transformed plant to pathogen infection relative to a corresponding nontransformed plant. The present invention also provides a transformed plant prepared by the method, progeny and seed thereof.

A transformed (transgenic) plant of the invention includes plants, the genome of which is augmented by a nucleic acid molecule of the invention, or in which the corresponding gene has been disrupted, e.g., to result in a loss, a decrease or an alteration, in the function of the product encoded by the gene, which plant may also have increased yields under conditions of pathogen infection and/or produce a better-quality product than the corresponding wild-type plant. The nucleic acid molecules of the invention are thus useful for targeted gene disruption, as well as markers and probes.

The invention also includes recombinant nucleic acid molecules which have been modified so as to comprise codons other than those present in the unmodified sequence. The recombinant nucleic acid molecules of the invention include those in which the modified codons specify amino acids that are the same as those specified by the codons in the unmodified sequence, as well as those that specify different amino acids, i.e., they encode a variant polypeptide having one or more amino acid substitutions relative to the polypeptide encoded by the unmodified sequence.

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The invention further includes a nucleotide sequence which is complementary to one (hereinafter "test" sequence) which hybridizes under stringent conditions with the nucleic acid molecules of the invention as well as RNA which is encoded by the nucleic acid molecule. When the hybridization is performed under stringent conditions, either the test or nucleic acid molecule of invention is preferably supported, e.g., on a membrane or DNA chip. Thus, either a denatured test or nucleic acid molecule of the invention is preferably first bound to a support and hybridization is effected for a specified period of time at a temperature of, e.g., between 55 and 70°C, in double strength citrate buffered saline (SC) containing 0.1% SDS followed by rinsing of the support at the same temperature but with a buffer having a reduced SC concentration. Depending upon the degree of stringency required such reduced concentration buffers are typically single strength SC containing 0.1% SDS, half strength SC containing 0.1% SDS and one-tenth strength SC containing 0.1% SDS.

The present invention also provides a method to identify a gene, the expression of which is altered in response to an external stimulus, e.g., pathogen infection. The method comprises contacting a plurality of samples comprising portions or fragments of isolated nucleic acid molecules with a probe which corresponds to a population of a nucleic acid sequences, the expression of which is altered in response to an external stimulus, so as to form a binary complex. Each sample corresponds to a different gene. Then complex formation is detected or determined. The method may be employed with nucleic acid samples and probes from any organism, e.g., any prokaryotic or eukaryotic organism. Preferably, the nucleic acid sample and probes are from a plant, such as a dicot or monocot. For example, the method comprises contacting a solid substrate comprising a plurality of samples comprising portions or fragments of isolated plant nucleic acid with a probe comprising plant nucleic acid corresponding to at least a portion of RNA from a pathogen infected plant so as to form a complex. Preferred pathogens are those which induce an R-gene dependent resistance response. Each individual sample comprises one or more nucleic acid sequences corresponding to a plant gene, e.g., a pool of oligonucleotides corresponding to the same gene or a portion of that gene. The plurality of samples is provided on a DNA chip. A second plurality of samples on a solid substrate, i.e., a DNA chip, each comprising a plurality of samples comprising portions or fragments of

isolated plant nucleic acid is contacted with a probe comprising plant nucleic acid corresponding to at least a portion of RNA from an uninfected or mutant plant or plant cells so as to form a complex. Then complex formation with nucleic acid from infected cells and from uninfected or mutant cells is compared.

The invention also provides a method for identifying a plant cell infected with a pathogen, e.g., one that induces a RPP4-, RPP7- and/or RPP8-dependent resistance response. The method comprises contacting nucleic acid obtained from a plant cell suspected of being infected with the pathogen with at least one, preferably at least two, oligonucleotides under conditions effective to amplify at least a portion of a nucleotide sequence in the isolated plant nucleic acid which is substantially similar to at least one of SEQ ID NOs: 1-684 and 792-795, so as to yield an amplified product. Then the presence of the amplified product is detected or determined. The presence of the amplified product, e.g., in an amount that is different than the amount of the corresponding amplified product from an uninfected or mutant plant, corresponding to one or more of SEQ ID NOs: 1-684 and 792-795 or an ortholog thereof, is indicative of pathogen infection.

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The invention provides an additional method for identifying a plant cell infected with a pathogen. The method comprises hybridizing a probe selected from SEQ ID NOs: 1-684 and 792-795 to nucleic acid obtained from a plant cell suspected of being infected with a pathogen. The amount of the probe hybridized to nucleic acid obtained from a cell suspected of being infected with a pathogen is compared to hybridization of the probe to nucleic acid isolated from an uninfected or mutant cell. A change in the amount of the hybridized probe in nucleic acid isolated from a cell suspected of being infected by a pathogen relative to the amount of hybridized probe in nucleic acid isolated from an uninfected or mutant cell is indicative of infection.

Also provided is an isolated nucleic acid molecule comprising a nucleotide sequence that directs transcription, e.g., a promoter, of a linked nucleic acid segment in a host cell, such as a plant cell, wherein transcription is altered, e.g., increased, in response to a pathogen infection. Preferably, the pathogen is R-dependent, and more preferably, one that triggers a response that is dependent on RPP4, RPP7 and/or RPP8, such as an oomycete (for example, Peronospora). It is preferred that the nucleotide sequence is from plant genomic

DNA which has at least 65%, 66%, 67%, 68%, 69%, 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, and 99%, nucleotide sequence identity to a sequence of a promoter from an *Arabidopsis* gene comprising any one of SEQ ID NOs: 1-684 and 792-795, e.g., SEQ ID NOs:774-788, or comprising motifs such as one of SEQ ID NOs:685-773. Thus, the invention also includes orthologs of *Arabidopsis* promoters. Preferably, the nucleotide sequence includes the promoter region from a gene corresponding to SEQ ID NOs: 1-770 and 792-795, which region preferably includes at least one copy of at least one of the following, e.g., a nucleic acid sequence comprising one of SEQ ID NOs:685-697 ("motif 1"), SEQ ID NOs:698-709 ("motif 2"), GGT/CCCA ("motif 3"), GNCCAAA ("motif 4"), or SEQ ID NOs:710-713, 714-756, or 757-773. The promoter sequence is preferably about 25 to 2000, e.g., 50 to 500 or 100 to 1400, nucleotides in length.

In one embodiment of the invention, the isolated nucleic acid molecule comprises a plant nucleotide sequence which is the promoter region for any one of SEQ ID NOs: 1-684 and 792-795, or is structurally related to the promoter for SEQ ID NOs: 1-684 and 792-795, i.e., is an orthologous promoter, and is linked to a plant structural gene or open reading frame. Hence, the present invention further provides an expression cassette or a recombinant vector containing the nucleic acid molecule, and the vector may be a plasmid. Such cassettes or vectors, when present in a plant, plant cell or plant tissue result in transcription of the linked nucleic acid segment in the plant, plant tissue or plant cell.

Transcription of the linked segment is altered in response to pathogen infection, including *Peronospora* infection. For promoters with motif 1, transcription of linked segments may be altered in response to agents or other stimuli that induce Myb-like transcription factors.

Generally, the promoters of the invention may be employed to express a nucleic acid segment that is operably linked to the promoter, for example, an open reading frame or a portion thereof, an anti-sense construct or a transgene. The open reading frame may be obtained from an insect resistance gene, a bacterial disease resistance gene, a fungal disease resistance gene, a viral disease resistance gene, a nematode disease resistance gene, a herbicide resistance gene, a gene affecting grain composition or quality, a nutrient utilization gene, a

mycotoxin reduction gene, a male sterility gene, a selectable marker gene, a screenable marker gene, a negative selectable marker, a positive selectable marker, a gene affecting plant agronomic characteristics, i.e., yield, standability and the like, or an environment or stress resistance gene, i.e., one or more genes that confer herbicide resistance or tolerance, insect resistance or tolerance, disease resistance or tolerance (viral, bacterial, fungal, oomycete, or nematode), stress tolerance or resistance (as exemplified by resistance or tolerance to drought, heat, chilling, freezing, excessive moisture, salt stress, or oxidative stress), increased yields, food content and makeup, physical appearance, male sterility, drydown, standability, prolificacy, starch properties or quantity, oil quantity and quality, amino acid or protein composition, and the like.

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Preferably, the promoters of the invention include a consecutive stretch of about 25 to 2000, including 50 to 500 or 100 to 250, and up to 1000 or 1500, contiguous nucleotides, e.g., 40 to about 743, 60 to about 743, 125 to about 743, 250 to about 743, 400 to about 743, 600 to about 743, of the promoter sequences from genes comprising any one of SEQ ID NOs:685-788, 714-756, and 757-773, or the promoter orthologs thereof, which include the minimal promoter region.

In a particular embodiment of the invention said consecutive stretch of about 25 to 2000, including 50 to 500 or 100 to 250, and up to 1000 or 1500, contiguous nucleotides, e.g., 40 to about 743, 60 to about 743, 125 to about 743, 250 to about 743, 400 to about 743, 600 to about 743, has at least 75%, preferably 80%, more preferably 90% and most preferably 95% sequence identity with a corresponding consecutive stretch of about 25 to 2000, including 50 to 500 or 100 to 250, and up to 1000 or 1500, contiguous nucleotides, e.g., 40 to about 743, 60 to about 743, 125 to about 743, 250 to about 743, 400 to about 743, 600 to about 743, of any one of SEQ ID NOs:685-788, 714-756, and 757-773, or the promoter orthologs thereof, which include the minimal promoter region.

The expression cassettes or vectors of the invention may optionally include other regulatory sequences, e.g., transcription terminator sequences, introns and/or enhancers, and may be contained in a host cell. The expression cassette or vector may augment the genome of a transformed plant or may be maintained extrachromosomally. The expression cassette or vector may further have a Ti plasmid and be contained in an Agrobacterium tumefaciens cell; it may

be carried on a microparticle, wherein the microparticle is suitable for ballistic transformation of a plant cell; or it may be contained in a plant cell protoplast. Further, the expression cassette can be contained in a plant, plant cell or plant tissue from a dicot or a monocot. In particular, the plant may be a cereal plant.

5 The present invention further provides a method of augmenting a plant genome by contacting plant cells with an expression cassette or vector of the invention, i.e., one having a nucleotide sequence that directs transcription of a linked nucleic acid segment in a plant cell, wherein transcription of the linked segment is altered in response to a pathogen such as an oomycete, e.g., 10 Peronospora, infection, and wherein the nucleic sequence is from plant DNA that has at least 65%, and more preferably at least 70%, identity to the sequence of a promoter from an Arabidopsis gene comprising any one of SEQ ID NOs: 1-684 and 792-795, so as to yield transformed plant cells; and regenerating the transformed plant cells to provide a differentiated transformed plant, wherein the 15 differentiated transformed plant expresses the linked fragment in the cells of the plant in response to infection. The present invention also provides a plant prepared by the method, progeny and seed thereof.

A transformed (transgenic) plant of the invention includes plants, the genome of which is augmented by a nucleic acid molecule of the invention, or in which the corresponding gene has been disrupted, e.g., to result in a loss, a decrease or an alteration, in the function of the product encoded by the gene, which plant may also have increased yields and/or produce a better-quality product than the corresponding wild-type plant. The nucleic acid molecules of the invention are thus useful for targeted gene disruption, as well as markers and probes.

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The invention also provides a method of plant breeding, e.g., to prepare a crossed fertile transgenic plant. The method comprises crossing a fertile transgenic plant comprising a particular nucleic acid molecule of the invention with itself or with a second plant, e.g., one lacking the particular nucleic acid molecule, to prepare the seed of a crossed fertile transgenic plant comprising the particular nucleic acid molecule. The seed is then planted to obtain a crossed fertile transgenic plant. The plant may be a monocot or a dicot. In a particular embodiment, the plant is a cereal plant.

The crossed fertile transgenic plant may have the particular nucleic acid molecule inherited through a female parent or through a male parent. The second plant may be an inbred plant. The crossed fertile transgenic may be a hybrid. Also included within the present invention are seeds of any of these crossed fertile transgenic plants.

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The present invention also provides a method to identify a nucleotide sequence that directs transcription of nucleic acid in the genome of a plant cell in response to pathogen exposure, by contacting a probe comprising plant nucleic acid, e.g., cRNA, isolated from tissues of a plant contacted with the pathogen with a plurality of isolated nucleic acid samples on a plurality of solid substrates, wherein each sample is a plurality of oligonucleotides corresponding to at least a portion of a plant gene, so as to form a complex between at least a portion of the probe and a nucleic acid sample(s) having sequences that are structurally related to the sequences in the probe. Then complex formation is determined or detected to determine which samples represent genes comprising promoters that are responsive to infection with the pathogen. The probe and/or samples may be nucleic acid from a dicot or from a monocot.

A method to shuffle the nucleic acid molecules of the invention is provided. This method involves fragmentation of a (parent) nucleic acid molecule corresponding to a nucleic acid sequence listed in SEQ ID NOs: 1-684 and 792-795, the ortholog thereof, or the corresponding gene thereof, followed by religation. This method allows for the production of polypeptides having altered activity relative to the polypeptide encoded by the parent nucleic acid molecule. Accordingly, the invention provides cells and transgenic plants containing nucleotide sequences produced through shuffling that encode polypeptides having altered activity relative to the polypeptide encoded by the parent nucleic acid molecule.

A computer readable medium containing the nucleic acid sequences of the invention as well as methods of use for the computer readable medium are provided. This medium allows a nucleic acid molecule corresponding to a nucleic acid sequence listed in SEQ ID NOs:1-399, 685-756 and 774-793 or 71, 78, 80, 117, 126, 150, 159, 168, 208, 214, 264-265, 355, 400-657, 659-684, 792-795 to be used as a reference sequence to search against databases. This medium also allows for computer-based manipulation of a nucleic acid sequence

corresponding to a nucleic acid sequence listed in SEQ ID NOs: 1-399, 685-756 and 774-793 or 71, 78, 80, 117, 126, 150, 159, 168, 208, 214, 264-265, 355, 400-657, 659-684, 792-795, and the corresponding gene or polypeptide encoded by the nucleic acid sequence.

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#### **Brief Description of the Figures**

Figure 1 depicts RPP-dependent defense pathways.

Figure 2 depicts nucleotide sequences including the promoter region and motifs therein for genes, the expression of which is altered in response to pathogen infection (SEQ ID NOs: 774-788).

Figure 3 is a schematic of the overlap in genes that are induced early after *P. parasitica* infection and in a *RPP8*-specific manner, genes that are induced late after infection and in a *RPP8*-specific manner, and genes induced early and late after infection in a *RPP7*-specific manner.

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Figure 4 shows a schematic of RPP4-pathway, RPP7-pathway and RPP8upregulated transcription factor genes.

#### **Detailed Description of the Invention**

#### **Definitions**

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The term "nucleic acid" refers to deoxyribonucleotides or ribonucleotides and polymers thereof in either single- or double-stranded form, composed of monomers (nucleotides) containing a sugar, phosphate and a base which is either a purine or pyrimidine. Unless specifically limited, the term encompasses nucleic acids containing known analogs of natural nucleotides which have similar binding properties as the reference nucleic acid and are metabolized in a manner similar to naturally occurring nucleotides. Unless otherwise indicated, a particular nucleic acid sequence also implicitly encompasses conservatively modified variants thereof (e.g., degenerate codon substitutions) and complementary sequences as well as the sequence explicitly indicated. Specifically, degenerate codon substitutions may be achieved by generating sequences in which the third position of one or more selected (or all) codons is substituted with mixed-base and/or deoxyinosine residues (Batzer et al., 1991; Ohtsuka et al., 1985; Rossolini et al., 1994). A "nucleic acid fragment" is a fraction of a given nucleic acid molecule. In higher plants, deoxyribonucleic

acid (DNA) is the genetic material while ribonucleic acid (RNA) is involved in the transfer of information contained within DNA into proteins. The term "nucleotide sequence" refers to a polymer of DNA or RNA which can be single-or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases capable of incorporation into DNA or RNA polymers. The terms "nucleic acid", "nucleic acid molecule", "nucleic acid fragment" or "nucleic acid sequence or segment" may also be used interchangeably with gene, cDNA, DNA and RNA encoded by a gene.

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The invention encompasses isolated or substantially purified nucleic acid or protein compositions. In the context of the present invention, an "isolated" or "purified" DNA molecule or an "isolated" or "purified" polypeptide is a DNA molecule or polypeptide that, by the hand of man, exists apart-from its native environment and is therefore not a product of nature. An isolated DNA molecule or polypeptide may exist in a purified form or may exist in a non-native environment such as, for example, a transgenic host cell. For example, an "isolated" or "purified" nucleic acid molecule or protein, or biologically active portion thereof, is substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. In one embodiment, an "isolated" nucleic acid is free of sequences that naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb, or 0.1 kb of nucleotide sequences that naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. A protein that is substantially free of cellular material includes preparations of protein or polypeptide having less than about 30%, 20%, 10%, 5%, (by dry weight) of contaminating protein. When the protein of the invention, or biologically active portion thereof, is recombinantly produced, preferably culture medium represents less than about 30%, 20%, 10%, or 5% (by dry weight) of chemical precursors or non-protein-of- interest chemicals. Fragments and variants of the disclosed nucleotide sequences and proteins or partial-length proteins encoded thereby are also encompassed by the present invention. By "fragment" or "portion" is meant a full length or less than

full length of the nucleotide sequence encoding, or the amino acid sequence of, a polypeptide or protein. Alternatively, fragments or portions of a nucleotide sequence that are useful as hybridization probes generally do not encode fragment proteins retaining biological activity. Thus, fragments or portions of a nucleotide sequence may range from at least about 9 nucleotides, about 12 nucleotides, about 20 nucleotides, about 50 nucleotides, about 100 nucleotides or more.

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The term "gene" is used broadly to refer to any segment of nucleic acid associated with a biological function. Thus, genes include coding sequences and/or the regulatory sequences required for their expression. For example, gene refers to a nucleic acid fragment that expresses mRNA, functional RNA, or specific protein, including regulatory sequences. Genes also include nonexpressed DNA segments that, for example, form recognition sequences for other proteins. Genes can be obtained from a variety of sources, including cloning from a source of interest or synthesizing from known or predicted sequence information, and may include sequences designed to have desired parameters.

"Naturally occurring" is used to describe an object that can be found in nature as distinct from being artificially produced by man. For example, a protein or nucleotide sequence present in an organism (including a virus), which can be isolated from a source in nature and which has not been intentionally modified by man in the laboratory, is naturally occurring.

A "marker gene" encodes a selectable or screenable trait.

"Selectable marker" is a gene whose expression in a cell gives the cell a selective advantage. The selective advantage possessed by the cells transformed with the selectable marker gene may be due to their ability to grow in the presence of a negative selective agent, such as an antibiotic or a herbicide, compared to the growth of non-transformed cells. The selective advantage possessed by the transformed cells, compared to non-transformed cells, may also be due to their enhanced or novel capacity to utilize an added compound as a nutrient, growth factor or energy source. Selectable marker gene also refers to a gene or a combination of genes whose expression in a cell gives the cell both a negative and/or a positive selective advantage.

The term "chimeric" refers to any gene or DNA that contains 1) DNA

sequences, including regulatory and coding sequences, that are not found together in nature, or 2) sequences encoding parts of proteins not naturally adjoined, or 3) parts of promoters that are not naturally adjoined. Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or comprise regulatory sequences and coding sequences derived from the same source, but arranged in a manner different from that found in nature.

A "transgene" refers to a gene that has been introduced into the genome by transformation and is stably maintained. Transgenes may include, for example, DNA that is either heterologous or homologous to the DNA of a particular plant to be transformed. Additionally, transgenes may comprise native genes inserted into a non-native organism, or chimeric genes. The term "endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene refers to a gene not normally found in the host organism but that is introduced by gene transfer.

The terms "protein," "peptide" and "polypeptide" are used interchangeably herein.

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By "variants" is intended substantially similar sequences. For nucleotide sequences, variants include those sequences that, because of the degeneracy of the genetic code, encode the identical amino acid sequence of the native protein. Naturally occurring allelic variants such as these can be identified with the use of well-known molecular biology techniques, as, for example, with polymerase chain reaction (PCR) and hybridization techniques. Variant nucleotide sequences also include synthetically derived nucleotide sequences, such as those generated, for example, by using site-directed mutagenesis which encode the native protein, as well as those that encode a polypeptide having amino acid substitutions. Generally, nucleotide sequence variants of the invention will have at least 40, 50, 60, to 70%, e.g., preferably 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, to 79%, generally at least 80%, e.g., 81%-84%, at least 85%, e.g., 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, to 98%, sequence identity to the native (endogenous) nucleotide sequence.

"DNA shuffling" is a method to introduce mutations or rearrangements, preferably randomly, in a DNA molecule or to generate exchanges of DNA sequences between two or more DNA molecules, preferably randomly. The

DNA molecule resulting from DNA shuffling is a shuffled DNA molecule that is a non-naturally occurring DNA molecule derived from at least one template DNA molecule. The shuffled DNA preferably encodes a variant polypeptide modified with respect to the polypeptide encoded by the template DNA, and may have an altered biological activity with respect to the polypeptide encoded by the template DNA.

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The nucleic acid molecules of the invention can be optimized for enhanced expression in plants of interest. See, for example, EPA035472; WO91/16432; Perlak et al., 1991; and Murray et al., 1989. In this manner, the genes or gene fragments can be synthesized utilizing plant-preferred codons. See, for example, Campbell and Gowri, 1990 for a discussion of host-preferred codon usage. Thus, the nucleotide sequences can be optimized for expression in any plant. It is recognized that all or any part of the gene sequence may be optimized or synthetic. That is, synthetic or partially optimized sequences may also be used. Variant nucleotide sequences and proteins also encompass sequences and protein derived from a mutagenic and recombinogenic procedure such as DNA shuffling. With such a procedure, one or more different coding sequences can be manipulated to create a new polypeptide possessing the desired properties. In this manner, libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides comprising sequence regions that have substantial sequence identity and can be homologously recombined in vitro or in vivo. Strategies for such DNA shuffling are known in the art. See, for example, Stemmer, 1994; Stemmer, 1994; Crameri et al., 1997; Moore et al., 1997; Zhang et al., 1997; Crameri et al., 1998; and U.S. Patent Nos. 5,605,793 and 5,837,458.

"Conservatively modified variations" of a particular nucleic acid sequence refers to those nucleic acid sequences that encode identical or essentially identical amino acid sequences, or where the nucleic acid sequence does not encode an amino acid sequence, to essentially identical sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given polypeptide. For instance the codons CGT, CGC, CGA, CGG, AGA, and AGG all encode the amino acid arginine. Thus, at every position where an arginine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the

encoded protein. Such nucleic acid variations are "silent variations" which are one species of "conservatively modified variations." Every nucleic acid sequence described herein which encodes a polypeptide also describes every possible silent variation, except where otherwise noted. One of skill will recognize that each codon in a nucleic acid (except ATG, which is ordinarily the only codon for methionine) can be modified to yield a functionally identical molecule by standard techniques. Accordingly, each "silent variation" of a nucleic acid which encodes a polypeptide is implicit in each described sequence.

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"Recombinant DNA molecule" is a combination of DNA sequences that are joined together using recombinant DNA technology and procedures used to join together DNA sequences as described, for example, in Sambrook et al., Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press (1989).

The terms "heterologous DNA sequence," "exogenous DNA segment" or "heterologous nucleic acid," each refer to a sequence that originates from a source foreign to the particular host cell or, if from the same source, is modified from its original form. Thus, a heterologous gene in a host cell includes a gene that is endogenous to the particular host cell but has been modified through, for example, the use of DNA shuffling. The terms also include non-naturally occurring multiple copies of a naturally occurring DNA sequence. Thus, the terms refer to a DNA segment that is foreign or heterologous to the cell, or homologous to the cell but in a position within the host cell nucleic acid in which the element is not ordinarily found. Exogenous DNA segments are expressed to yield exogenous polypeptides.

A "homologous" DNA sequence is a DNA sequence that is naturally associated with a host cell into which it is introduced.

"Wild-type" refers to the normal gene, or organism found in nature without any known mutation.

"Genome" refers to the complete genetic material of an organism.

"Vector" is defined to include, inter alia, any plasmid, cosmid, phage or Agrobacterium binary vector in double or single stranded linear or circular form which may or may not be self transmissible or mobilizable, and which can transform prokaryotic or eukaryotic host either by integration into the cellular genome or exist extrachromosomally (e.g. autonomous replicating plasmid with an origin of replication).

Specifically included are shuttle vectors by which is meant a DNA vehicle capable, naturally or by design, of replication in two different host organisms, which may be selected from actinomycetes and related species, bacteria and eukaryotic (e.g. higher plant, mammalian, yeast or fungal cells).

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"Cloning vectors" typically contain one or a small number of restriction endonuclease recognition sites at which foreign DNA sequences can be inserted in a determinable fashion without loss of essential biological function of the vector, as well as a marker gene that is suitable for use in the identification and selection of cells transformed with the cloning vector. Marker genes typically include genes that provide tetracycline resistance, hygromycin resistance or ampicillin resistance.

-"Expression cassette" as used herein means a DNA sequence capable of directing expression of a particular nucleotide sequence in an appropriate host cell, comprising a promoter operably linked to the nucleotide sequence of interest which is operably linked to termination signals. It also typically comprises sequences required for proper translation of the nucleotide sequence. The coding region usually codes for a protein of interest but may also code for a functional RNA of interest, for example antisense RNA or a nontranslated RNA, in the sense or antisense direction. The expression cassette comprising the nucleotide sequence of interest may be chimeric, meaning that at least one of its components is heterologous with respect to at least one of its other components. The expression cassette may also be one which is naturally occurring but has been obtained in a recombinant form useful for heterologous expression. The expression of the nucleotide sequence in the expression cassette may be under the control of a constitutive promoter or of an inducible promoter which initiates transcription only when the host cell is exposed to some particular external stimulus. In the case of a multicellular organism, the promoter can also be specific to a particular tissue or organ or stage of development.

Such expression cassettes will comprise the transcriptional initiation

region of the invention linked to a nucleotide sequence of interest. Such an
expression cassette is provided with a plurality of restriction sites for insertion of
the gene of interest to be under the transcriptional regulation of the regulatory
regions. The expression cassette may additionally contain selectable marker
genes.

The transcriptional cassette will include in the 5'-3' direction of transcription, a transcriptional and translational initiation region, a DNA sequence of interest, and a transcriptional and translational termination region functional in plants. The termination region may be native with the transcriptional initiation region, may be native with the DNA sequence of interest, or may be derived from another source. Convenient termination regions are available from the Ti-plasmid of A. tumefaciens, such as the octopine synthase and nopaline synthase termination regions. See also, Guerineau et al., 1991; Proudfoot, 1991; Sanfacon et al., 1991; Mogen et al., 1990; Munroe et al., 1990; Ballas et al., 1989; Joshi et al., 1987.

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An oligonucleotide corresponding to a nucleic acid molecule of the invention may be about 30 or fewer nucleotides in length (e.g., 9, 12, 15, 18, 20, 21 or 24, or any number between 9 and 30). Generally specific primers are upwards of 14 nucleotides in length. For optimum specificity and cost effectiveness, primers of 16-24 nucleotides in length may be preferred. Those skilled in the art are well versed in the design of primers for use processes such as PCR. If required, probing can be done with entire restriction fragments of the gene disclosed herein which may be 100's or even 1000's of nucleotides in length.

"Coding sequence" refers to a DNA or RNA sequence that codes for a specific amino acid sequence and excludes the non-coding sequences. It may constitute an "uninterrupted coding sequence", i.e., lacking an intron, such as in a cDNA or it may include one or more introns bounded by appropriate splice junctions. An "intron" is a sequence of RNA which is contained in the primary transcript but which is removed through cleavage and re-ligation of the RNA within the cell to create the mature mRNA that can be translated into a protein.

The terms "open reading frame" and "ORF" refer to the amino acid sequence encoded between translation initiation and termination codons of a coding sequence. The terms "initiation codon" and "termination codon" refer to a unit of three adjacent nucleotides ('codon') in a coding sequence that specifies initiation and chain termination, respectively, of protein synthesis (mRNA translation).

A "functional RNA" refers to an antisense RNA, ribozyme, or other RNA that is not translated.

The term "RNA transcript" refers to the product resulting from RNA polymerase catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript and is referred to as the mature RNA. "Messenger RNA" (mRNA) refers to the RNA that is without introns and that can be translated into protein by the cell. "cDNA" refers to a single- or a double-stranded DNA that is complementary to and derived from mRNA.

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"Regulatory sequences" and "suitable regulatory sequences" each refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences include enhancers, promoters, translation leader sequences, introns, and polyadenylation signal sequences. They include natural and synthetic sequences as well as sequences which may be a combination of synthetic and natural sequences. As is noted above, the term "suitable regulatory sequences" is not limited to promoters. However, some suitable regulatory sequences useful in the present invention will include, but are not limited to constitutive plant promoters, plant tissue-specific promoters, plant development specific promoters, inducible plant promoters and viral promoters.

"5' non-coding sequence" refers to a nucleotide sequence located 5' (upstream) to the coding sequence. It is present in the fully processed mRNA upstream of the initiation codon and may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency (Turner et al., 1995).

"3' non-coding sequence" refers to nucleotide sequences located 3' (downstream) to a coding sequence and include polyadenylation signal sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The use of different 3' non-coding sequences is exemplified by Ingelbrecht et al., 1989.

The term "translation leader sequence" refers to that DNA sequence portion of a gene between the promoter and coding sequence that is transcribed into RNA and is present in the fully processed mRNA upstream (5') of the translation start codon. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency.

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The term "mature" protein refers to a post-translationally processed polypeptide without its signal peptide. "Precursor" protein refers to the primary product of translation of an mRNA. "Signal peptide" refers to the amino terminal extension of a polypeptide, which is translated in conjunction with the polypeptide forming a precursor peptide and which is required for its entrance into the secretory pathway. The term "signal sequence" refers to a nucleotide sequence that encodes the signal peptide.

The term "intracellular localization sequence" or "signal sequence" refers to a nucleotide sequence that encodes an intracellular targeting signal. An "intracellular targeting signal" is an amino acid sequence that is translated in conjunction with a protein and directs it to a particular sub-cellular compartment. "Endoplasmic reticulum (ER) stop transit signal" refers to a carboxy-terminal extension of a polypeptide, which is translated in conjunction with the polypeptide and causes a protein that enters the secretory pathway to be retained in the ER. "ER stop transit sequence" refers to a nucleotide sequence that encodes the ER targeting signal. Other intracellular targeting sequences encode targeting signals active in seeds and/or leaves and vacuolar targeting signals.

"Promoter" refers to a nucleotide sequence, usually upstream (5') to its coding sequence, which controls the expression of the coding sequence by providing the recognition for RNA polymerase and other factors required for proper transcription. "Promoter" includes a minimal promoter that is a short DNA sequence comprised of a TATA- box and other sequences that serve to specify the site of transcription initiation, to which regulatory elements are added for control of expression. "Promoter" also refers to a nucleotide sequence that includes a minimal promoter plus regulatory elements that is capable of controlling the expression of a coding sequence or functional RNA. This type of promoter sequence consists of proximal and more distal upstream elements, the latter elements often referred to as enhancers. Accordingly, an "enhancer" is a DNA sequence which can stimulate promoter activity and may be an innate

element of the promoter or a heterologous element inserted to enhance the level or tissue specificity of a promoter. It is capable of operating in both orientations (normal or flipped), and is capable of functioning even when moved either upstream or downstream from the promoter. Both enhancers and other upstream promoter elements bind sequence-specific DNA-binding proteins that mediate their effects. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even be comprised of synthetic DNA segments. A promoter may also contain DNA sequences that are involved in the binding of protein factors which control the effectiveness of transcription initiation in response to physiological or developmental conditions.

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The "initiation site" is the position surrounding the first nucleotide that is part of the transcribed sequence, which is also defined as position +1. With respect to this site all other sequences of the gene and its controlling regions are numbered. Downstream sequences (i.e. further protein encoding sequences in the 3' direction) are denominated positive, while upstream sequences (mostly of the controlling regions in the 5' direction) are denominated negative.

Promoter elements, particularly a TATA element, that are inactive or that have greatly reduced promoter activity in the absence of upstream activation are referred to as "minimal or core promoters." In the presence of a suitable transcription factor, the minimal promoter functions to permit transcription. A "minimal or core promoter" thus consists only of all basal elements needed for transcription initiation, e.g., a TATA box and/or an initiator.

"Constitutive expression" refers to expression using a constitutive or regulated promoter. "Conditional" and "regulated expression" refer to expression controlled by a regulated promoter.

"Constitutive promoter" refers to a promoter that is able to express the gene that it controls in all or nearly all of the plant tissues during all or nearly all developmental stages of the plant. Each of the transcription-activating elements do not exhibit an absolute tissue-specificity, but mediate transcriptional activation in most plant parts at a level of ≥1% of the level reached in the part of the plant in which transcription is most active.

"Regulated promoter" refers to promoters that direct gene expression not constitutively, but in a temporally- and/or spatially-regulated manner, and

include both tissue-specific and inducible promoters. It includes natural and synthetic sequences as well as sequences which may be a combination of synthetic and natural sequences. Different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental conditions. New promoters of various types useful in plant cells are constantly being discovered, numerous examples may be found in the compilation by Okamuro et al., 1989. Since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

Typical regulated promoters useful in plants include but are not limited to safener-inducible promoters, promoters derived from the tetracycline-inducible system, promoters derived from salicylate-inducible systems, promoters derived from alcohol-inducible systems, promoters derived from glucocorticoid-inducible system, promoters derived from pathogen-inducible systems, and promoters derived from ecdysome-inducible systems.

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"Tissue-specific promoter" refers to regulated promoters that are not expressed in all plant cells but only in one or more cell types in specific organs (such as leaves or seeds), specific tissues (such as embryo or cotyledon), or specific cell types (such as leaf parenchyma or seed storage cells). These also include promoters that are temporally regulated, such as in early or late embryogenesis, during fruit ripening in developing seeds or fruit, in fully differentiated leaf, or at the onset of senescence.

"Inducible promoter" refers to those regulated promoters that can be turned on in one or more cell types by an external stimulus, such as a chemical, light, hormone, stress, or a pathogen.

"Operably-linked" refers to the association of nucleic acid sequences on single nucleic acid fragment so that the function of one is affected by the other. For example, a regulatory DNA sequence is said to be "operably linked to" or "associated with" a DNA sequence that codes for an RNA or a polypeptide if the two sequences are situated such that the regulatory DNA sequence affects expression of the coding DNA sequence (i.e., that the coding sequence or functional RNA is under the transcriptional control of the promoter). Coding sequences can be operably-linked to regulatory sequences in sense or antisense orientation.

"Expression" refers to the transcription and/or translation of an endogenous gene or a transgene in plants. For example, in the case of antisense constructs, expression may refer to the transcription of the antisense DNA only. In addition, expression refers to the transcription and stable accumulation of sense (mRNA) or functional RNA. Expression may also refer to the production of protein.

"Altered levels" refers to the level of expression in transgenic cells or organisms that differs from that of normal or untransformed cells or organisms.

"Overexpression" refers to the level of expression in transgenic cells or organisms that exceeds levels of expression in normal or untransformed cells or organisms.

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"Antisense inhibition" refers to the production of antisense RNA transcripts capable of suppressing the expression of protein from an endogenous gene or a transgene.

"Co-suppression" and "transwitch" each refer to the production of sense RNA transcripts capable of suppressing the expression of identical or substantially similar transgene or endogenous genes (U.S. Patent No. 5,231,020).

"Gene silencing" refers to homology-dependent suppression of viral genes, transgenes, or endogenous nuclear genes. Gene silencing may be transcriptional, when the suppression is due to decreased transcription of the affected genes, or post-transcriptional, when the suppression is due to increased turnover (degradation) of RNA species homologous to the affected genes. (English et al., 1996). Gene silencing includes virus-induced gene silencing (Ruiz et al., 1998).

"Silencing suppressor" gene refers to a gene whose expression leads to counteracting gene silencing and enhanced expression of silenced genes.

Silencing suppressor genes may be of plant, non-plant, or viral origin. Examples include, but are not limited to HC-Pro, P1-HC-Pro, and 2b proteins. Other examples include one or more genes in TGMV-B genome.

"Transcription stop fragment" refers to nucleotide sequences that contain one or more regulatory signals, such as polyadenylation signal sequences, capable of terminating transcription. Examples include the 3' non-regulatory regions of genes encoding nopaline synthase and the small subunit of ribulose bisphosphate carboxylase.

"Translation stop fragment" refers to nucleotide sequences that contain one or more regulatory signals, such as one or more termination codons in all three frames, capable of terminating translation. Insertion of a translation stop fragment adjacent to or near the initiation codon at the 5' end of the coding sequence will result in no translation or improper translation. Excision of the translation stop fragment by site-specific recombination will leave a site-specific sequence in the coding sequence that does not interfere with proper translation using the initiation codon.

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The terms "cis-acting sequence" and "cis-acting element" refer to DNA or RNA sequences whose functions require them to be on the same molecule. An example of a cis-acting sequence on the replicant is the viral replication origin.

The terms "trans-acting sequence" and "trans-acting element" refer to DNA or RNA sequences whose function does not require them to be on the same molecule.

"Chromosomally-integrated" refers to the integration of a foreign gene or DNA construct into the host DNA by covalent bonds. Where genes are not "chromosomally integrated" they may be "transiently expressed." Transient expression of a gene refers to the expression of a gene that is not integrated into the host chromosome but functions independently, either as part of an autonomously replicating plasmid or expression cassette, for example, or as part of another biological system such as a virus.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

- (a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full length cDNA or gene sequence, or the complete cDNA or gene sequence.
- 30 (b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences.

Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

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Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent identity between any two sequences can be accomplished using a mathematical algorithm. Preferred, non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller, 1988; the local homology algorithm of Smith et al., 1981; the homology alignment algorithm of Needleman and Wunsch, 1970; the search-for-similarity-method of Pearson and Lipman, 1988; the algorithm of Karlin and Altschul, 1990, modified as in Karlin and Altschul, 1993.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, California); the ALIGN program (Version 2.0) and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Version 8 (available from Genetics Computer Group (GCG), 575 Science Drive, Madison, Wisconsin, USA). Alignments using these programs can be performed using the default parameters. The CLUSTAL program is well described by Higgins et al., 1988; Higgins et al., 1989; Corpet et al., 1988; Huang et al., 1992; and Pearson et al., 1994. The ALIGN program is based on the algorithm of Myers and Miller, *supra*. The BLAST programs of Altschul et al., 1990, are based on the algorithm of Karlin and Altschul *supra*.

Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al., 1990). These initial neighborhood word hits act as seeds for initiating searches to find

longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when the cumulative alignment score falls off by the quantity X from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached.

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul (1993). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a test nucleic acid sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid sequence to the reference nucleic acid sequence is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul et al., 1997.

Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships between molecules. See Altschul et al., supra. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (e.g. BLASTN for nucleotide sequences, BLASTX for proteins) can be used. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, 1989). See <a href="http://www.ncbi.nlm.nih.gov.">http://www.ncbi.nlm.nih.gov.</a> Alignment may also be performed manually by inspection.

For purposes of the present invention, comparison of nucleotide sequences for determination of percent sequence identity to the promoter sequences disclosed herein is preferably made using the BlastN program (version 1.4.7 or later) with its default parameters or any equivalent program. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by the preferred program.

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- (c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences makes reference to a specified percentage of residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window, as measured by sequence comparison algorithms or by visual inspection. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity." Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a nonconservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California).
- (d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal

alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

(e)(i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, or 79%, preferably at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, and most preferably at least 95%, 96%, 97%, 98%, or 99% sequence identity, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 70%, more preferably at least 80%, 90%, and most preferably at least 95%.

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Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions (see below). Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T<sub>m</sub>) for the specific sequence at a defined ionic strength and pH. However, stringent conditions encompass temperatures in the range of about 1°C to about 20°C, depending upon the desired degree of stringency as otherwise qualified herein. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is when the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e)(ii) The term "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with at least 70%, 71%, 72%, 73%, 74%,

75%, 76%, 77%, 78%, or 79%, preferably 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, or even more preferably, 95%, 96%, 97%, 98% or 99%, sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch, 1970. An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution.

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For sequence comparison, typically one sequence acts as a reference sequence to which test-sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer, subsequence coordinates are designated if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

As noted above, another indication that two nucleic acid sequences are substantially identical is that the two molecules hybridize to each other under stringent conditions. The phrase "hybridizing specifically to" refers to the binding, duplexing, or hybridizing of a molecule only to a particular nucleotide sequence under stringent conditions when that sequence is present in a complex mixture (e.g., total cellular) DNA or RNA. "Bind(s) substantially" refers to complementary hybridization between a probe nucleic acid and a target nucleic acid and embraces minor mismatches that can be accommodated by reducing the stringency of the hybridization media to achieve the desired detection of the target nucleic acid sequence.

"Stringent hybridization conditions" and "stringent hybridization wash conditions" in the context of nucleic acid hybridization experiments such as Southern and Northern hybridizations are sequence dependent, and are different under different environmental parameters. Longer sequences hybridize specifically at higher temperatures. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Specificity is typically the function of post-

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hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the  $T_{m}$  can be approximated from the equation of Meinkoth and Wahl, 1984; T<sub>m</sub> 81.5°C + 16.6 (log M) +0.41 (%GC) - 0.61 (% form) - 500/L; where M is the molarity of monovalent cations. %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs.  $T_m$  is reduced by about 1°C for each 1% of mismatching; thus, T<sub>m</sub>, hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with >90% identity are sought, the T<sub>m</sub> can be decreased 10°C. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T<sub>m</sub>) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4°C lower than the thermal melting point (T<sub>m</sub>); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10°C lower than the thermal melting point (T<sub>m</sub>); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20°C lower than the thermal melting point (T<sub>m</sub>). Using the equation, hybridization and wash compositions, and desired T, those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T of less than 45°C (aqueous solution) or 32°C (formamide solution), it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen, 1993. Generally, highly stringent hybridization and wash conditions are selected to be about 5°C lower than the thermal melting point (T<sub>m</sub>) for the specific sequence at a defined ionic strength and pH.

An example of highly stringent wash conditions is 0.15 M NaCl at 72°C for about 15 minutes. An example of stringent wash conditions is a 0.2X SSC wash at 65°C for 15 minutes (see, Sambrook, *infra*, for a description of SSC buffer). Often, a high stringency wash is preceded by a low stringency wash to remove background probe signal. An example medium stringency wash for a duplex of, e.g., more than 100 nucleotides, is 1X SSC at 45°C for 15 minutes.

An example low stringency wash for a duplex of, e.g., more than 100 nucleotides, is 4-6X SSC at 40°C for 15 minutes. For short probes (e.g., about 10 to 50 nucleotides), stringent conditions typically involve salt concentrations of less than about 1.5 M, more preferably about 0.01 to 1.0 M, Na ion concentration (or other salts) at pH 7.0 to 8.3, and the temperature is typically at least about 30°C and at least about 60°C for long robes (e.g., >50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. In general, a signal to noise ratio of 2X (or higher) than that observed for an unrelated probe in the particular hybridization assay indicates detection of a specific hybridization. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the proteins that they encode are substantially identical. This occurs, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code.

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Very stringent conditions are selected to be equal to the  $T_m$  for a particular probe. An example of stringent conditions for hybridization of complementary nucleic acids which have more than 100 complementary residues on a filter in a Southern or Northern blot is 50% formamide, e.g., hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0. 1X SSC at 60 to 65°C. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C.

The following are examples of sets of hybridization/wash conditions that may be used to clone orthologous nucleotide sequences that are substantially identical to reference nucleotide sequences of the present invention: a reference nucleotide sequence preferably hybridizes to the reference nucleotide sequence in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 2X SSC, 0.1% SDS at 50°C, more desirably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 1X SSC, 0.1% SDS at 50°C, more desirably still in 7% sodium dodecyl sulfate (SDS), 0.5

M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.5X SSC, 0.1% SDS at 50°C, preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 50°C, more preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 65°C.

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By "variant" polypeptide is intended a polypeptide derived from the native protein by deletion (so-called truncation) or addition of one or more amino acids to the N-terminal and/or C-terminal end of the native protein; deletion or addition of one or more amino acids at one or more sites in the native protein; or substitution of one or more amino acids at one or more sites in the native protein. Such variants may results form, for example, genetic polymorphism or from human manipulation. Methods for such manipulations are generally known in the art.

Thus, the polypeptides of the invention may be altered in various ways including amino acid substitutions, deletions, truncations, and insertions. Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of the polypeptides can be prepared by mutations in the DNA. Methods for mutagenesis and nucleotide sequence alterations are well known in the art. See, for example, Kunkel, 1985; Kunkel et al., 1987; U. S. Patent No. 4,873,192; Walker and Gaastra, 1983, and the references cited therein. Guidance as to appropriate amino acid substitutions that do not affect biological activity of the protein of interest may be found in the model of Dayhoff et al., 1978, herein incorporated by reference. Conservative substitutions, such as exchanging one amino acid with another having similar properties, are preferred.

Thus, the genes and nucleotide sequences of the invention include both the naturally occurring sequences as well as mutant forms. Likewise, the polypeptides of the invention encompass both naturally occurring proteins as well as variations and modified forms thereof. Such variants will continue to possess the desired activity. The deletions, insertions, and substitutions of the polypeptide sequence encompassed herein are not expected to produce radical changes in the characteristics of the polypeptide. However, when it is difficult to predict the exact effect of the substitution, deletion, or insertion in advance of

doing so, one skilled in the art will appreciate that the effect will be evaluated by routine screening assays.

Individual substitutions deletions or additions that alter, add or delete a single amino acid or a small percentage of amino acids (typically less than 5%, more typically less than 1%) in an encoded sequence are "conservatively modified variations," where the alterations result in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art. The following five groups each contain amino acids that are conservative substitutions for one another: Aliphatic: Glycine (G), Alanine (A), Valine (V), Leucine (L), Isoleucine (I); Aromatic: Phenylalanine (F), Tyrosine (Y), Tryptophan (W); Sulfur-containing: Methionine (M), Cysteine (C); Basic: Arginine (R), Lysine (K), Histidine (H); Acidic: Aspartic acid (D), Glutamic acid (E), Asparagine (N), Glutamine (Q). See also, Creighton, 1984. In addition, individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids in an encoded sequence are also "conservatively modified variations."

"Production tissue" refers to mature, harvestable tissue consisting of nondividing, terminally-differentiated cells. It excludes young, growing tissue consisting of germline, meristematic, and not-fully-differentiated cells.

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"Germline cells" refer to cells that are destined to be gametes and whose genetic material is heritable.

The word "plant" refers to any plant, particularly to seed plant, and "plant cell" is a structural and physiological unit of the plant, which comprises a cell wall but may also refer to a protoplast. The plant cell may be in form of an isolated single cell or a cultured cell, or as a part of higher organized unit such as, for example, a plant tissue, or a plant organ.

"Plant tissue" includes differentiated and undifferentiated tissues or plants, including but not limited to roots, stems, shoots, leaves, pollen, seeds, tumor tissue and various forms of cells and culture such as single cells, protoplast, embryos, and callus tissue. The plant tissue may be in plants or in organ, tissue or cell culture.

The term "altered plant trait" means any phenotypic or genotypic change in a transgenic plant relative to the wild-type or non-transgenic plant host.

The term "transformation" refers to the transfer of a nucleic acid fragment into the genome of a host cell, resulting in genetically stable inheritance. Host cells containing the transformed nucleic acid fragments are referred to as "transgenic" cells, and organisms comprising transgenic cells are referred to as "transgenic organisms". Examples of methods of transformation of plants and plant cells include *Agrobacterium*-mediated transformation (De Blaere et al., 1987) and particle bombardment technology (Klein et al., 1987; U.S. Patent No. 4,945,050). Whole plants may be regenerated from transgenic cells by methods well known to the skilled artisan (see, for example, Fromm et al., 1990).

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"Transformed," "transgenic," and "recombinant" refer to a host cell or organism such as a bacterium or a plant into which a heterologous nucleic acid molecule has been introduced. The nucleic acid molecule can be stably integrated into the genome generally known in the art and are disclosed in Sambrook et al., 1989. See also Innis et al., 1995; and Gelfand, 1995; and Innis and Gelfand, 1999. Known methods of PCR include, but are not limited to, methods using paired primers, nested primers, single specific primers, degenerate primers, gene-specific primers, vector-specific primers, partially mismatched primers, and the like. For example, "transformed," "transformant," and "transgenic" plants or calli have been through the transformation process and contain a foreign gene integrated into their chromosome. The term "untransformed" refers to normal plants that have not been through the transformation process.

A "transgenic plant" is a plant having one or more plant cells that contain an expression vector.

"Transiently transformed" refers to cells in which transgenes and foreign DNA have been introduced (for example, by such methods as *Agrobacterium*-mediated transformation or biolistic bombardment), but not selected for stable maintenance.

"Stably transformed" refers to cells that have been selected and regenerated on a selection media following transformation.

"Transient expression" refers to transgene expression in cells, e.g., after transformation with recombinant virus or by such methods as *Agrobacterium*-mediated transformation, electroporation, or biolistic bombardment, but not selected for its stable maintenance.

"Genetically stable" and "heritable" refer to chromosomally-integrated genetic elements that are stably maintained in the plant and stably inherited by progeny through successive generations.

"Primary transformant" and "T0 generation" refer to transgenic plants that are of the same genetic generation as the tissue which was initially transformed (i.e., not having gone through meiosis and fertilization since transformation).

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"Secondary transformants" and the "T1, T2, T3, etc. generations" refer to transgenic plants derived from primary transformants through one or more meiotic and fertilization cycles. They may be derived by self-fertilization of primary or secondary transformants or crosses of primary or secondary transformants with other transformed or untransformed plants.

"Significant increase" is an increase that is larger than the margin of error inherent in the measurement technique, preferably an increase by about 2-fold or greater.

"Significantly less" means that the decrease is larger than the margin of error inherent in the measurement technique, preferably a decrease by about 2-fold or greater.

# I. The Nucleic Acid Molecules of the Invention and Polypeptide Encoded Thereby

This invention relates to isolated plant, e.g., Arabidopsis and rice, nucleic acid molecules, sequences and segments (fragments), the expression of which is altered in response to pathogen infection, as well as the endogenous plant promoters for those expressed molecules, sequences or segments. However, the expression of these genes may also be altered in response to non-pathogens, e.g., in response to environmental stimuli. The nucleic acid molecules can be used in pathogen control strategies, e.g., by overexpressing nucleic acid molecules which can confer tolerance to a cell, or by altering the expression of host genes which are required for pathogen infection, e.g., by "knocking out" the expression of at least one genomic copy of the gene. Plants having genetic disruptions in host genes may be less susceptible to infection, e.g., due to a decrease or absence of a host protein needed for infection, or, alternatively, hypersusceptible to infection. Plants that are hypersusceptible to infection may be useful to prepare transgenic

plants as the expression of the gene(s) which was disrupted may be related to gene silencing.

Preferred sources for the nucleic acid molecules of the invention include, but are not limited to, corn (Zea mays), Brassica sp. (e.g., B. napus, B. rapa, B. juncea), particularly those Brassica species useful as sources of seed oil, alfalfa (Medicago sativa), rice (Oryza sativa), rye (Secale cereale), sorghum (Sorghum bicolor, Sorghum vulgare), millet (e.g., pearl millet (Pennisetum glaucum), proso millet (Panicum miliaceum), foxtail millet (Setaria italica), finger millet (Eleusine coracana)), sunflower (Helianthus annuus), safflower (Carthamus tinctorius), wheat (Triticum aestivum), soybean (Glycine max), tobacco (Nicotiana tabacum), potato (Solanum tuberosum), peanuts (Arachis hypogaea), cotton (Gossypium barbadense, Gossypium hirsutum), sweet potato (Ipomoea batatus), cassava (Manihot esculenta), coffee (Cofea spp.), coconut (Cocos nucifera), pineapple (Ananas comosus), citrus trees (Citrus spp.), cocoa 15 (Theobroma cacao), tea (Camellia sinensis), banana (Musa spp.), avocado (Persea americana), fig (Ficus casica), guava (Psidium guajava), mango (Mangifera indica), olive (Olea europaea), papaya (Carica papaya), cashew (Anacardium occidentale), macadamia (Macadamia integrifolia), almond (Prunus amygdalus), sugar beets (Beta vulgaris), sugarcane (Saccharum spp.), oats, barley, vegetables, ornamentals, and conifers; duckweed (Lemna, see WO 20 00/07210, which includes members of the family Lemnaceae. There are known four genera and 34 species of duckweed as follows: genus Lemna (L. aequinoctialis, L. disperma, L. ecuadoriensis, L. gibba, L. japonica, L. minor, L. miniscula, L. obscura, L. perpusilla, L. tenera, L. trisulca, L. turionifera, L. valdiviana); genus Spirodela (S. intermedia, S. polyrrhiza, S. punctata); genus 25 Wo~ffia (Wa. angusta, Wa. arrhiza, Wa. australina, Wa. borealis, Wa. brasiliensis, Wa. columbiana, Wa. elongata, Wa. globosa, Wa. microscopica, Wa. neglecta) and genus Wo-fiella (W1. caudata, W1. denticulata, W1. gladiata, W1. hyalina, W1. lingulata, W1. repunda, W1. rotunda, and W1. neotropica). Any other genera or species of Lemnaceae, if they exist, are also aspects of the 30 present invention. Lemna gibba, Lemna minor, and Lemna miniscula are preferred, with Lemna minor and Lemna miniscula being most preferred. Lemna species can be classified using the taxonomic scheme described by Landolt,

Biosystematic Investigation on the Family of Duckweeds: The family of

Lemnaceae - A Monograph Study. Geobatanischen Institut ETH, Stiftung Rubel, Zurich (1986)); vegetables including tomatoes (Lycopersicon esculentum). lettuce (e.g., Lactuca sativa), green beans (Phaseolus vulgaris), lima beans (Phaseolus limensis), peas (Lathyrus spp.), and members of the genus Cucumis such as cucumber (C. sativus), cantaloupe (C. cantalupensis), and musk melon-5 (C. melo). Ornamentals include azalea (Rhododendron spp.), hydrangea (Macrophylla hydrangea), hibiscus (Hibiscus rosasanensis), roses (Rosa spp.), tulips (Tulipa spp.), daffodils (Narcissus spp.), petunias (Petunia hybrida), carnation (Dianthus caryophyllus), poinsettia (Euphorbia pulcherrima), and 10 chrysanthemum. Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (Pinus taeda), slash pine (Pinus elliotii), ponderosa pine (Pinus ponderosa), lodgepole pine (Pinus contorta), and Monterey pine (Pinus radiata), Douglas-fir (Pseudotsuga menziesii); Western hemlock (Tsuga canadensis); Sitka spruce (Picea glauca); redwood (Sequoia sempervirens); true firs such as silver fir (Abies amabilis) and balsam fir (Abies balsamea); and cedars such as Western red cedar (Thuja plicata) and Alaska yellow-cedar (Chamaecyparis nootkatensis). Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, 20 chickpea, etc. Legumes include, but are not limited to, Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, Acacia, aneth, artichoke, arugula, 25 blackberry, canola, cilantro, clementines, escarole, eucalyptus, fennel, grapefruit, honey dew, jicama, kiwifruit, lemon, lime, mushroom, nut, okra, orange, parsley, persimmon, plantain, pomegranate, poplar, radiata pine, radicchio, Southern pine, sweetgum, tangerine, triticale, vine, yams, apple, pear, quince, cherry, apricot, melon, hemp, buckwheat, grape, raspberry, chenopodium, blueberry, 30 nectarine, peach, plum, strawberry, watermelon, eggplant, pepper, cauliflower, Brassica, e.g., broccoli, cabbage, brussels sprouts, onion, carrot, leek, beet, broad bean, celery, radish, pumpkin, endive, gourd, garlic, snapbean, spinach, squash, turnip, asparagus, and zucchini and ornamental plants include impatiens, Begonia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula,

Saint Paulia, Agertum, Amaranthus, Antihirrhinum, Aquilegia, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossos, and Zinnia.

Other vegetable sources (and databases to identify orthologs of the invention) for the nucleic acid sequences of the invention include those are shown in Table 1.

Table 1

				<u> </u>
FAMILY	LATIN NAME	COMMON NAME	MAP REFERENCES RESOURCES	LINKS
Cucurbitaceae	Cucumis sativus	Cucumber		http://www.cucurbit.c
	Cucumis melo	Melon		http://genome.cornell edu/cgc/
·	Citrullus lanatus	Watermelon		
	Cucurbita pepo	Squash – summer		
	Cucurbita maxima	Squash - winter	·	<u> </u>
	Cucurbita moschata	Pumpkin /butternut		
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Solanaceae	Lycopersicon esculentum	Tomato	Genome center (www.genome.c lemson.edu)  11.6x BAC of L. cheesmanii	genome.cornell.edu/s olgenes http://ars- genome.cornell.edu/c gi- bin/WebAce/webace? db=solgenes http://genome.cornell edu/tgc/ http://tgrc.ucdavis.edu/

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J	}	1	1013, 1994	
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			Reports 12:293-	
			297, 1993	
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		j	132:1141-1160,	
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	Solanum	Eggplant		
	melongena			•
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	(Zea mays)	(Field Corn)		http://www.agron.mis
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Liliaceae	Allium cepa	Onion		
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		(Asparagus)	·	
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			.gov/pgdic/Map pro	

Preferred forage and turf grass nucleic acid sources for use in the methods of the invention include alfalfa, orchard grass, tall fescue, perennial ryegrass, creeping bent grass, and redtop. Preferably, the nucleic acid sources are crop plants and in particular cereals (for example, corn, alfalfa, sunflower, rice, *Brassica*, canola, soybean, barley, soybean, sugarbeet, cotton, safflower, peanut, sorghum, wheat, millet, tobacco, etc.), and even more preferably corn and soybean.

According to one embodiment, the present invention is directed to a nucleic acid molecule comprising a nucleotide sequence obtained or obtainable

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from any plant gene which encodes a polypeptide having at least 70% amino acid sequence identity to a polypeptide encoded by SEQ ID NOs. 1-684 or 789-795. or a promoter for said gene. Thus, based on the Arabidopsis nucleic acid sequences of the present invention, orthologs of those sequences may be identified or isolated from the genome of any desired organism, preferably from another plant, according to well known techniques based on their sequence similarity to the Arabidopsis coding sequences, e.g., hybridization, PCR or computer generated sequence comparisons. For example, all or a portion of a particular Arabidopsis sequence is used as a probe that selectively hybridizes to other gene sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen source organism. Further, suitable genomic and cDNA libraries may be prepared from any cell or tissue of an organism. Such techniques include hybridization screening of plated DNA libraries (either plaques or colonies; see, e.g., Sambrook et al., 1989) and amplification by PCR using oligonucleotide primers preferably corresponding to sequence domains conserved among related polypeptide or subsequences of the nucleotide sequences provided herein (see, e.g., Innis et al., 1990). These methods are particularly well suited to the isolation of gene sequences from organisms closely related to the organism from which the probe sequence is derived. The application of these methods using the Arabidopsis coding sequences as probes is well suited for the isolation of gene sequences from any source organism, preferably other plant species. In a PCR approach, oligonucleotide primers can be designed for use in PCR reactions to amplify corresponding DNA sequences from cDNA or genomic DNA extracted from any plant of interest. Methods for designing PCR primers and PCR cloning are generally known in the art as discussed hereinabove.

In hybridization techniques, all or part of a known nucleotide sequence is used as a probe that selectively hybridizes to other corresponding nucleotide sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen organism. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labeled with a detectable group such as <sup>32</sup>P, or any other detectable marker. Thus, for example, probes for hybridization can be made by labeling synthetic oligonucleotides based on the

sequence of the invention. Methods for preparation of probes for hybridization and for construction of cDNA and genomic libraries are generally known in the art and are disclosed in Sambrook et al., 1989. In general, sequences that hybridize to the sequences disclosed herein will have at least 40% to 50%, about 60% to 70% and even about 80% 85%, 90%, 95% to 98% or more identity with the disclosed sequences. That is, the sequence similarity of sequences may range, sharing at least about 40% to 50%, about 60% to 70%, and even about 80%, 85%, 90%, 95% to 98% sequence similarity.

The nucleic acid molecules of the invention can also be identified by, for example, a search of known databases for genes encoding polypeptides having a specified amino acid sequence identity. Methods of alignment of sequences for comparison are well known in the art and are described hereinabove.

## II. Expression Cassettes of the Invention

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The present invention also encompasses expression cassettes, preferably in the form of a recombinant vectors comprising the nucleic acid sequences of the invention. In such vectors, the expression cassette comprises regulatory elements for expression of the nucleotide sequences in a host cell capable of expressing the nucleotide sequences. Such regulatory elements usually comprise promoter and termination signals and preferably also comprise elements allowing efficient translation of polypeptides encoded by the nucleic acid sequences of the present invention. For efficient initiation of translation, sequences adjacent to the initiating methionine may require modification. For example, they can be modified by the inclusion of sequences known to be effective in plants. Joshi (1987) has suggested an appropriate consensus for plants and Clontech suggests a further consensus translation initiator (1993/1994 catalog, page 210). These consensuses are suitable for use with the nucleotide sequences of this invention. The sequences are incorporated into constructions comprising the nucleotide sequences, up to and including the ATG (whilst leaving the second amino acid unmodified), or alternatively up to and including the GTC subsequent to the ATG (with the possibility of modifying the second amino acid of the transgene).

Vectors comprising the nucleic acid sequences are usually capable of replication in particular host cells, e.g., as extrachromosomal molecules, and are

therefore used to amplify the nucleic acid sequences of this invention in the host cells. In a preferred embodiment, host cells for such vectors are plant cells.

## A. Promoters and Enhancers

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Expression of the nucleotide sequences in transgenic plants is driven by promoters shown to be functional in plants. The choice of promoter will vary depending on the temporal and spatial requirements for expression, and also depending on the target species. In many cases, expression in multiple tissues is desirable. Although many promoters from dicotyledons have been shown to be operational in monocotyledons and *vice versa*, ideally dicotyledonous promoters are selected for expression in dicotyledons, and monocotyledonous promoters for expression in monocotyledons. However, there is no restriction to the provenance of selected promoters; it is sufficient that they are operational in driving the expression of the nucleotide sequences in the desired cell.

These promoters include, but are not limited to, constitutive, inducible, temporally regulated, developmentally regulated, chemically regulated, stress-responsive, tissue-preferred and tissue-specific promoters. Promoter sequences are known to be strong or weak. A strong promoter provides for a high level of gene expression, whereas a weak promoter provides for a very low level of gene expression. An inducible promoter is a promoter that provides for the turning on and off of gene expression in response to an exogenously added agent, or to an environmental or developmental stimulus. A bacterial promoter such as the P<sub>tac</sub> promoter can be induced to varying levels of gene expression depending on the level of isothiopropylgalactoside added to the transformed bacterial cells. An isolated promoter sequence that is a strong promoter for heterologous nucleic acid is advantageous because it provides for a sufficient level of gene expression to allow for easy detection and selection of transformed cells and provides for a high level of gene expression when desired.

Preferred promoters that are expressed constitutively include promoters from genes encoding actin or ubiquitin and the CaMV 35S and 19S promoters. The nucleotide sequences of this invention can also be expressed under the regulation of promoters that are chemically regulated. This enables the nucleic acid sequence or encoded polypeptide to be synthesized only when the crop plants are treated with the inducing chemicals. Preferred technology for chemical induction of gene expression is detailed in the published application EP

0 332 104 (to Ciba-Geigy) and U.S. Patent 5,614,395. A preferred promoter for chemical induction is the tobacco PR-1a promoter.

Tissue-specific or tissue-preferential promoters useful in the present invention. Also useful are promoters which confer seed-specific expression, such as those disclosed by Schernthaner et al., 1988; anther (tapetal) specific promoter B6 (Huffman et al.); and pistil-specific promoters such as a modified S13 promoter (Dzelkalns et al., 1993).

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Preferred tissue specific expression patterns include green tissue-specific, root-specific, stem-specific, and flower-specific. Promoters suitable for expression in green tissue include many which regulate genes involved in photosynthesis and many of these have been cloned from both monocotyledons and dicotyledons. A preferred promoter is the maize PEPC promoter from the phosphoenol carboxylase gene (Hudspeth & Grula, 1989). A preferred promoter for root-specific expression is that described by de Framond (1991; EP 0 452 269 to Ciba-Geigy). A preferred stem specific promoter is that described in U.S. Patent No. 5,625,136 (to Ciba-Geigy) and which drives expression of the maize trpA gene.

Other promoters which direct specific or enhanced expression in certain plant tissues will be known to those of skill in the art in light of the present disclosure. These include, for example, the *rbcS* promoter, specific for green tissue; the *ocs*, *nos*, and *mas* promoters which have higher activity in roots or wounded leaf tissue; a truncated (-90 to +8) 35S promoter which directs enhanced expression in roots, an tubulin gene that directs expression in roots and promoters derived from zein storage protein genes which direct expression in endosperm. It is particularly contemplated that one may advantageously use the 16 bp *ocs* enhancer element from the octopine synthase (*ocs*) gene (Bonchez et al., 1989), especially when present in multiple copies, to achieve enhanced expression in roots.

Preferred plant promoters include, but are not limited to, a promoter such as the CaMV 35S promoter, an enhanced 35S promoter or others such as CaMV 19S, nos, Adh1, sucrose synthase, α-tubulin, ubiquitin, actin, cab, PEPCase or those associated with the R gene complex. Further suitable promoters include the U2 and U5 snRNA promoters from maize, the promoter from alcohol dehydrogenase, the Z4 promoter from a gene encoding the Z4 22 kD zein

protein, the Z10 promoter from a gene encoding a 10 kD zein protein, a Z27 promoter from a gene encoding a 27 kD zein protein, the A20 promoter from the gene encoding a 19 kD -zein protein, inducible promoters, such as the light inducible promoter derived from the pea rbcS gene and the actin promoter from rice; seed specific promoters, such as the phaseolin promoter from beans, may also be used. Other promoters useful in the practice of the invention are known to those of skill in the art.

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Examples of tissue specific promoters which have been described include the lectin (Vodkin, 1983; Lindstrom et al., 1990,) corn alcohol dehydrogenase 1 (Vogel et al., 1992; Dennis et al., 1984), corn light harvesting complex (Simpson, 1985; Bansal et al., 1992), corn heat shock protein (Odell et al., 1985; Rochester et al., 1986), pea small subunit RuBP carboxylase (Poulsen et al., 1986; Cashmore et al., 1983), Ti plasmid mannopine synthase (Langridge et al., 1989), Ti plasmid nopaline synthase (Langridge et al., 1989), petunia chalcone isomerase (vanTunen et al., 1988), bean glycine rich protein 1 (Keller et al., 1989), truncated CaMV 35s (Odell et al., 1985), potato patatin (Wenzler et al., 1989), root cell (Yamamoto et al., 1990), maize zein (Reina et al., 1990; Kriz et al., 1987; Wandelt et al., 1989; Langridge et al., 1983; Reina et al., 1990), globulin-1 (Belanger et al., 1991), α-tubulin, cab (Sullivan et al., 1989), PEPCase (Hudspeth & Grula, 1989), R gene complex-associated promoters (Chandler et al., 1989), and chalcone synthase promoters (Franken et al., 1991).

Inducible promoters that have been described include the ABA- and turgor-inducible promoters, the promoter of the auxin-binding protein gene (Schwob et al., 1993), the UDP glucose flavonoid glycosyl-transferase gene promoter (Ralston et al., 1988), the MPI proteinase inhibitor promoter (Cordero et al., 1994), and the glyceraldehyde-3-phosphate dehydrogenase gene promoter (Kohler et al., 1995; Quigley et al., 1989; Martinez et al., 1989).

Several tissue-specific regulated genes and/or promoters have been reported in plants. These include genes encoding the seed storage proteins (such as napin, cruciferin, beta-conglycinin, and phaseolin) zein or oil body proteins (such as oleosin), or genes involved in fatty acid biosynthesis (including acyl carrier protein, stearoyl-ACP desaturase, and fatty acid desaturases (fad 2-1)), and other genes expressed during embryo development (such as Bce4, see, for example, EP 255378 and Kridl et al., 1991). Particularly useful for seed-specific

expression is the pea vicilin promoter (Czako et al., 1992. (See also U.S. Pat. No. 5,625,136, herein incorporated by reference.) Other useful promoters for expression in mature leaves are those that are switched on at the onset of senescence, such as the SAG promoter from Arabidopsis (Gan et al., 1995, 270 (5244), 1986-8).

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A class of fruit-specific promoters expressed at or during antithesis through fruit development, at least until the beginning of ripening, is discussed in U.S. 4,943,674, the disclosure of which is hereby incorporated by reference. cDNA clones that are preferentially expressed in cotton fiber have been isolated (John et al., 1992). cDNA clones from tomato displaying differential expression during fruit development have been isolated and characterized (Mansson et al., 1985, Slater et al., 1985). The promoter for polygalacturonase gene is active in fruit ripening. The polygalacturonase gene is described in U.S. Patent No. 4,535,060, U.S. Patent No. 4,769,061, U.S. Patent No. 4,801,590, and U.S. Patent No. 5,107,065, which disclosures are incorporated herein by reference.

Other examples of tissue-specific promoters include those that direct expression in leaf cells following damage to the leaf (for example, from chewing insects), in tubers (for example, patatin gene promoter), and in fiber cells (an example of a developmentally-regulated fiber cell protein is E6 (John et al., 1992). The E6 gene is most active in fiber, although low levels of transcripts are found in leaf, ovule and flower.

The tissue-specificity of some "tissue-specific" promoters may not be absolute and may be tested by one skilled in the art using the diphtheria toxin sequence. One can also achieve tissue-specific expression with "leaky" expression by a combination of different tissue-specific promoters (Beals et al., 1997). Other tissue-specific promoters can be isolated by one skilled in the art (see U.S. 5,589,379). Several inducible promoters ("gene switches") have been reported. Many are described in the review by Gatz (1996 and 1997). These include tetracycline repressor system, *Lac* repressor system, copper-inducible systems, salicylate-inducible systems (such as the PR1a system), glucocorticoid-(Aoyama, 1997) and ecdysome-inducible systems. Also included are the benzene sulphonamide- (U.S. Patent No. 5,364,780) and alcohol- (WO 97/06269 and WO 97/06268) inducible systems and glutathione S-transferase promoters. Other studies have focused on genes inducibly regulated in response to

environmental stress or stimuli such as increased salinity. drought, pathogen and wounding. (Graham et al., 1985; Graham et al., 1985, Smith et al., 1986). Accumulation of metallocarboxypeptidase-inhibitor protein has been reported in leaves of wounded potato plants (Graham et al., 1981). Other plant genes have been reported to be induced methyl jasmonate, elicitors, heat-shock, anaerobic stress, or herbicide safeners.

Frequently it is desirable to have continuous or inducible expression of a DNA sequence throughout the cells of an organism in a tissue-independent manner. For example, increased resistance of a plant to infection by soil- and air borne pathogens might be accomplished by genetic manipulation of the plant's genome to comprise a continuous promoter operably linked to a heterologous or homologous-pathogen-resistance gene such that pathogen-resistance proteins are continuously expressed throughout the plant's tissues.

Alternatively, it might be desirable to inhibit expression of a native DNA sequence within a plant's tissues to achieve a desired phenotype. In this case, such inhibition might be accomplished with transformation of the plant to comprise a constitutive, tissue-independent promoter operably linked to an antisense nucleotide sequence, such that constitutive expression of the antisense sequence produces an RNA transcript that interferes with translation of the mRNA of the native DNA sequence.

Other elements include those that can be regulated by endogenous or exogenous agents, e.g., by DNA binding proteins such as zinc finger proteins, including naturally occurring zinc finger proteins or chimeric zinc finger proteins (see, e.g., U.S. Patent No. 5,789,538, WO 99/48909; WO 99/45132; WO 98/53060; WO 98/53057; WO 98/53058; WO 00/23464; WO 95/19431; and WO 98/54311) or myb-like transcription factors. For example, a chimeric zinc finger protein may include amino acid sequences which bind to a specific DNA sequence (the zinc finger) and amino acid sequences that activate (e.g., GAL 4 sequences) or repress the transcription of the sequences linked to the specific DNA sequence.

## B. 5' and 3' Sequences

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In addition to promoters, a variety of 3 transcriptional terminators are also available for use in the present invention. Transcriptional terminators are responsible for the termination of transcription and correct mRNA

polyadenylation. The 3' nontranslated regulatory DNA sequence preferably includes from about 50 to about 1,000, more preferably about 100 to about 1,000, nucleotide base pairs and contains plant transcriptional and translational termination sequences. Appropriate transcriptional terminators and those which are known to function in plants include the CaMV 35S terminator, the tml terminator, the nopaline synthase terminator, the pea rbcS E9 terminator, the terminator for the T7 transcript from the octopine synthase gene of Agrobacterium tumefaciens, and the 3' end of the protease inhibitor I or II genes from potato or tomato, although other 3' elements known to those of skill in the art can also be employed.

The 5' regulatory region of the expression cassette may also include other enhancing sequences. Numerous sequences have been found to enhance gene expression in transgenic plants. These include sequences which have been shown to enhance expression such as intron sequences (e.g., from Adh1, bronze1 or the sucrose synthase intron) and viral leader sequences (e.g., from TMV, MCMV and AMV). For example, a number of non-translated leader sequences derived from viruses are known to enhance expression. Specifically, leader sequences from Tobacco Mosaic Virus (TMV), Maize Chlorotic Mottle Virus (MCMV), and Alfalfa Mosaic Virus (AMV) have been shown to be effective in enhancing expression (e.g., Gallie et al., 1987; Skuzeski et al., 1990). Other leaders known in the art include but are not limited to: Picornavirus leaders, for example, EMCV leader (Encephalomyocarditis 5 noncoding region) (Elroy-Stein et al., 1989); Potyvirus leaders, for example, TEV leader (Tobacco Etch Virus) (Allison et al., 1986); MDMV leader (Maize Dwarf Mosaic Virus); Human immunoglobulin heavy-chain binding protein (BiP) leader, (Macejak et al., 1991); Untranslated leader from the coat protein mRNA of alfalfa mosaic virus (AMV RNA 4), (Jobling et al., 1987; Tobacco mosaic virus leader (TMV), (Gallie et al., 1989; and Maize Chlorotic Mottle Virus leader (MCMV) (Lommel et al., 1991. See also, Della-Cioppa et al., 1987.

# 30 C. Targeting Sequences

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It may be preferable to target expression of the nucleotide sequences of the present invention to different cellular localizations in the plant. In some cases, localization in the cytosol may be desirable, whereas in other cases, localization in some subcellular organelle, e.g., the nucleus, may be preferred. Subcellular

localization of transgene encoded enzymes is undertaken using techniques well known in the art. Typically, the DNA encoding the target peptide from a known organelle-targeted gene product is manipulated and fused upstream of the nucleotide sequence. Many such target sequences are known for the chloroplast and their functioning in heterologous constructions has been shown. The expression of the nucleotide sequences of the present invention is also targeted to the endoplasmic reticulum or to the vacuoles of the host cells. Techniques to achieve this are well-known in the art.

### D. Marker Genes

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In order to improve the ability to identify transformants, one may desire to employ a selectable or screenable marker gene as, or in addition to, the preselected nucleic acid sequence or segment. "Marker genes" are genes that impart a distinct phenotype to cells expressing the marker gene and thus allow such transformed cells to be distinguished from cells that do not have the marker. Such genes may encode either a selectable or screenable marker, depending on whether the marker confers a trait which one can 'select' for by chemical means, i.e., through the use of a selective agent (e.g., a herbicide, antibiotic, or the like), or whether it is simply a trait that one can identify through observation or testing, i.e., by 'screening' (e.g., the R-locus trait). Of course, many examples of suitable marker genes are known to the art and can be employed in the practice of the invention.

Included within the terms selectable or screenable marker genes are also genes which encode a "secretable marker" whose secretion can be detected as a means of identifying or selecting for transformed cells. Examples include markers which encode a secretable antigen that can be identified by antibody interaction, or even secretable enzymes which can be detected by their catalytic activity. Secretable proteins fall into a number of classes, including small, diffusible proteins detectable, e.g., by ELISA; small active enzymes detectable in extracellular solution (e.g., α-amylase, β-lactamase, phosphinothricin acetyltransferase); and proteins that are inserted or trapped in the cell wall (e.g., proteins that include a leader sequence such as that found in the expression unit of extensin or tobacco PR-S).

With regard to selectable secretable markers, the use of a gene that encodes a polypeptide that becomes sequestered in the cell wall, and which polypeptide

includes a unique epitope is considered to be particularly advantageous. Such a secreted antigen marker would ideally employ an epitope sequence that would provide low background in plant tissue, a promoter-leader sequence that would impart efficient expression and targeting across the plasma membrane, and would produce protein that is bound in the cell wall and yet accessible to antibodies. A normally secreted wall protein modified to include a unique epitope would satisfy all such requirements.

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571, 1987).

Elements of the present disclosure are exemplified in detail through the use of particular marker genes. However in light of this disclosure, numerous other possible selectable and/or screenable marker genes will be apparent to those of skill in the art in addition to the one set forth herein below. Therefore, it will be understood that the following discussion is exemplary rather than exhaustive. In light of the techniques disclosed herein and the general recombinant techniques which are known in the art, the present invention renders possible the introduction of any gene, including marker genes, into a recipient cell to generate a transformed plant cell, e.g., a monocot cell.

Possible selectable markers for use in connection with the present invention include, but are not limited to, a neo gene, which codes for kanamycin resistance and can be selected for using kanamycin, G418, a gene encoding resistance to bleomycin, and the like; a bar gene which codes for bialaphos resistance; a gene which encodes an altered EPSP synthase protein thus conferring glyphosate resistance; a nitrilase gene such as bxn from Klebsiella ozaenae which confers resistance to bromoxynil; a mutant acetolactate synthase gene (ALS) which confers resistance to imidazolinone, sulfonylurea or other ALS-inhibiting chemicals (European Patent Application 154,204, 1985); a methotrexate-resistant DHFR gene; a dalapon dehalogenase gene that confers resistance to the herbicide dalapon; or a mutated anthranilate synthase gene that confers resistance to 5-methyl tryptophan. Where a mutant EPSP synthase gene is employed, additional benefit may be realized through the incorporation of a suitable chloroplast transit peptide, CTP (European Patent Application 0 218

An illustrative embodiment of a selectable marker gene capable of being used in systems to select transformants is the genes that encode the enzyme phosphinothricin acetyltransferase, such as the bar gene from Streptomyces

hygroscopicus or the pat gene from Streptomyces viridochromogenes (U.S. Patent No. 5,550,318). The enzyme phosphinothricin acetyltransferase (PAT) inactivates the active ingredient in the herbicide bialaphos, phosphinothricin (PPT). PPT inhibits glutamine synthetase, causing rapid accumulation of ammonia and cell death. The success in using this selective system in conjunction with monocots was particularly surprising because of the major difficulties which have been reported in transformation of cereals.

Screenable markers that may be employed include, but are not limited to, a β-glucuronidase or *uid*A gene (GUS) which encodes an enzyme for which various chromogenic substrates are known; an R-locus gene, which encodes a product that regulates the production of anthocyanin pigments (red color) in plant tissues; a -lactamase gene, which encodes an enzyme for which various chromogenic substrates are known (e.g., PADAC, a chromogenic cephalosporin); a *xyl*E gene which encodes a catechol dioxygenase that can convert chromogenic catechols; an -amylase gene; a tyrosinase gene which encodes an enzyme capable of oxidizing tyrosine to DOPA and dopaquinone which in turn condenses to form the easily detectable compound melanin; a -galactosidase gene, which encodes an enzyme for which there are chromogenic substrates; a luciferase (*lux*) gene, which allows for bioluminescence detection; or an aequorin gene, which may be employed in calcium-sensitive bioluminescence detection, or a green fluorescent protein.

Genes from the maize R gene complex are contemplated to be particularly useful as screenable markers. The R gene complex in maize encodes a protein that acts to regulate the production of anthocyanin pigments in most seed and plant tissue. Maize strains can have one, or as many as four, R alleles which combine to regulate pigmentation in a developmental and tissue specific manner. A gene from the R gene complex was applied to maize transformation, because the expression of this gene in transformed cells does not harm the cells. Thus, an R gene introduced into such cells will cause the expression of a red pigment and, if stably incorporated, can be visually scored as a red sector. If a maize line carries dominant alleles for genes encoding the enzymatic intermediates in the anthocyanin biosynthetic pathway (C2, A1, A2, Bz1 and Bz2), but carries a recessive allele at the R locus, transformation of any cell from that line with R will result in red pigment formation. Exemplary lines include Wisconsin 22

which contains the rg-Stadler allele and TR112, a K55 derivative which is r-g, b, Pl. Alternatively any genotype of maize can be utilized if the C1 and R alleles are introduced together.

A further screenable marker contemplated for use in the present invention is firefly luciferase, encoded by the *lux* gene. The presence of the *lux* gene in transformed cells may be detected using, for example, X-ray film, scintillation counting, fluorescent spectrophotometry, low-light video cameras, photon counting cameras or multiwell luminometry. It is also envisioned that this system may be developed for populational screening for bioluminescence, such as on tissue culture plates, or even for whole plant screening.

## E. Other Sequences

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Plasmid vectors include additional DNA sequences that provide for easy selection, amplification, and transformation of the expression cassette in prokaryotic and eukaryotic cells, e.g., pUC-derived vectors such as pUC8, pUC9, pUC18, pUC19, pUC23, pUC119, and pUC120, pSK-derived vectors, pGEM-derived vectors, pSP-derived vectors, or pBS-derived vectors. The additional DNA sequences include origins of replication to provide for autonomous replication of the vector, additional selectable marker genes, preferably encoding antibiotic or herbicide resistance, unique multiple cloning sites providing for multiple sites to insert DNA sequences or genes encoded in the expression cassette, and sequences that enhance transformation of prokaryotic and eukaryotic cells.

Another vector that is useful for expression in both plant and prokaryotic cells is the binary Ti plasmid (as disclosed in Schilperoort et al., U.S. Patent No. 4,940,838) as exemplified by vector pGA582. This binary Ti plasmid vector has been previously characterized by An, cited *supra*. This binary Ti vector can be replicated in prokaryotic bacteria such as *E. coli* and *Agrobacterium*. The *Agrobacterium* plasmid vectors can be used to transfer the expression cassette to dicot plant cells, and under certain conditions to monocot cells, such as rice cells. The binary Ti vectors preferably include the nopaline T DNA right and left borders to provide for efficient plant cell transformation, a selectable marker gene, unique multiple cloning sites in the T border regions, the *colE1* replication of origin and a wide host range replicon. The binary Ti vectors carrying an

expression cassette of the invention can be used to transform both prokaryotic and eukaryotic cells, but is preferably used to transform dicot plant cells.

Virtually any DNA may be used for delivery to recipient cells to ultimately produce fertile transgenic plants in accordance with the present invention. For example, DNA segments in the form of vectors and plasmids, or linear DNA fragments, in some instance containing only the DNA element to be expressed in the plant, and the like, may be employed.

Vectors, plasmids, cosmids, YACs (yeast artificial chromosomes) and DNA segments for use in transforming such cells will, of course, generally comprise the cDNA, gene or genes which one desires to introduce into the cells. These DNA constructs can further include structures such as promoters, enhancers, polylinkers, or even regulatory genes as desired. The DNA segment or gene chosen for cellular introduction will often encode a protein which will be expressed in the resultant recombinant cells, such as will result in a screenable or selectable trait and/or which will impart an improved phenotype to the regenerated plant. However, this may not always be the case, and the present invention also encompasses transgenic plants incorporating non-expressed transgenes.

#### 20 III. Transformation

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The expression cassettes of the present invention can be introduced into a host cell, e.g., a plant cell, in a number of art-recognized ways. Those skilled in the art will appreciate that the choice of method might depend on the type of cell, e.g., monocotyledonous or dicotyledonous, targeted for transformation. Vectors which may be used to transform plant tissue with the expression cassettes of the present invention include both *Agrobacterium* vectors and ballistic vectors, as well as vectors suitable for DNA-mediated transformation, e.g., direct uptake or via electroporation. However, cells other than plant cells may be transformed with the expression cassettes of the invention.

Suitable methods of transforming plant cells include, but are not limited to, microinjection (Crossway et al., 1986), direct DNA transfer to plant cells by PEG precipitation; liposomes; electroporation (Riggs et al., 1986, Agrobacterium-mediated transformation (Hinchee et al., 1988), direct gene transfer (Paszkowski et al., 1984), and ballistic particle acceleration using

devices available from Agracetus, Inc., Madison, Wis. and BioRad, Hercules, Calif. (see, for example, Sanford et al., U.S. Pat. No. 4,945,050; and McCabe et al., 1988). Also see, Weissinger et al., 1988; Sanford et al., 1987 (onion); Christou et al., 1988 (soybean); McCabe et al., 1988 (soybean); Datta et al., 1990 (rice); Klein et al., 1988 (maize); Klein et al., 1988 (maize); Klein et al., 1988 (maize); Fromm et al., 1990 (maize); and Gordon-Kamm et al., 1990 (maize); Svab et al., 1990 (tobacco chloroplast); Koziel et al., 1993 (maize); Shimamoto et al., 1989 (rice); Christou et al., 1991 (rice); European Patent Application EP 0 332 581 (orchardgrass and other Pooideae); Vasil et al., 1993 (wheat); Weeks et al., 1993 (wheat).

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In one embodiment, a nucleotide sequence of the present invention is directly transformed into the plastid genome. Plastid transformation technology is extensively described in U.S. Patent Nos. 5,451,513, 5,545,817, and 5,545,818, in PCT application no. WO 95/16783, and in McBride et al., 1994. The basic technique for chloroplast transformation involves introducing regions 15 of cloned plastid DNA flanking a selectable marker together with the gene of interest into a suitable target tissue, e.g., using biolistics or protoplast transformation (e.g., calcium chloride or PEG mediated transformation). The 1 to 1.5 kb flanking regions, termed targeting sequences, facilitate orthologous 20 recombination with the plastid genome and thus allow the replacement or modification of specific regions of the plastome. Initially, point mutations in the chloroplast 16S rRNA and rps12 genes conferring resistance to spectinomycin and/or streptomycin are utilized as selectable markers for transformation (Svab et al., 1990; Staub et al., 1992). This resulted in stable homoplasmic transformants 25 at a frequency of approximately one per 100 bombardments of target leaves. The presence of cloning sites between these markers allowed creation of a plastid targeting vector for introduction of foreign genes (Staub et al., 1993). Substantial increases in transformation frequency are obtained by replacement of the recessive rRNA or r-protein antibiotic resistance genes with a dominant 30 selectable marker, the bacterial aadA gene encoding the spectinomycindetoxifying enzyme aminoglycoside-3'-adenyltransferase (Svab et al., 1993). Other selectable markers useful for plastid transformation are known in the art and encompassed within the scope of the invention. Typically, approximately 15-20 cell division cycles following transformation are required to reach a

homoplastidic state. Plastid expression, in which genes are inserted by orthologous recombination into all of the several thousand copies of the circular plastid genome present in each plant cell, takes advantage of the enormous copy number advantage over nuclear-expressed genes to permit expression levels that can readily exceed 10% of the total soluble plant protein. In a preferred embodiment, a nucleotide sequence of the present invention is inserted into a plastid targeting vector and transformed into the plastid genome of a desired plant host. Plants homoplastic for plastid genomes containing a nucleotide sequence of the present invention are obtained, and are preferentially capable of high expression of the nucleotide sequence.

Agrobacterium tumefaciens cells containing a vector comprising an expression-cassette of the present invention, wherein the vector-comprises a Ti plasmid, are useful in methods of making transformed plants. Plant cells are infected with an Agrobacterium tumefaciens as described above to produce a transformed plant cell, and then a plant is regenerated from the transformed plant cell. Numerous Agrobacterium vector systems useful in carrying out the present invention are known. For example, U.S. Pat. No. 4,459,355 discloses a method for transforming susceptible plants, including dicots, with an Agrobacterium strain containing the Ti plasmid. The transformation of woody plants with an Agrobacterium vector is disclosed in U.S. Patent No. 4,795,855. Further, U.S. Patent No. 4,940,838 to Schilperoort et al. discloses a binary Agrobacterium vector (i.e., one in which the Agrobacterium contains one plasmid having the vir region of a Ti plasmid but no T region, and a second plasmid having a T region but no vir region) useful in carrying out the present invention.

It is particularly preferred to use the binary type vectors of Ti and Ri plasmids of Agrobacterium spp. Ti-derived vectors transform a wide variety of higher plants, including monocotyledonous and dicotyledonous plants, such as soybean, cotton, rape, tobacco, and rice (Pacciotti et al., 1985: Byrne et al., 1987; Sukhapinda et al., 1987; Lorz et al., 1985; Potrykus, 1985; Park et al., 1985: Hiei et al., 1994. The use of T-DNA to transform plant cells has received extensive study and is amply described (EP 120516; Hoekema, 1985; Knauf, et al., 1983; and An. et al., 1985. For introduction into plants, the nucleotide sequences of the invention can be inserted into binary vectors as described in the examples.

Transformation of plants can be undertaken with a single DNA molecule or multiple DNA molecules (i.e., co-transformation), and both these techniques are suitable for use with the expression cassettes of the present invention.

Numerous transformation vectors are available for plant transformation, and the expression cassettes of this invention can be used in conjunction with any such vectors. The selection of vector will depend upon the preferred transformation technique and the target species for transformation.

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Preferred plant cells for transformation include, but are not limited to, cells from plant such as corn (Zea mays), Brassica sp. (e.g., B. napus, B. rapa, B. juncea), particularly those Brassica species useful as sources of seed oil, alfalfa (Medicago sativa), rice (Oryza sativa), rye (Secale cereale), sorghum (Sorghum bicolor, Sorghum vulgare), millet (e.g., pearl millet (Pennisetum glaucum), proso millet (Panicum miliaceum), foxtail millet (Setaria italica), finger millet (Eleusine coracana)), sunflower (Helianthus annuus), safflower (Carthamus tinctorius), wheat (Triticum aestivum), soybean (Glycine max), tobacco 15 (Nicotiana tabacum), potato (Solanum tuberosum), peanuts (Arachis hypogaea), cotton (Gossypium barbadense, Gossypium hirsutum), sweet potato (Ipomoea batatus), cassava (Manihot esculenta), coffee (Cofea spp.), coconut (Cocos nucifera), pineapple (Ananas comosus), citrus trees (Citrus spp.), cocoa 20 (Theobroma cacao), tea (Camellia sinensis), banana (Musa spp.), avocado '(Persea americana), fig (Ficus casica), guava (Psidium guajava), mango (Mangifera indica), olive (Olea europaea), papaya (Carica papaya), cashew (Anacardium occidentale), macadamia (Macadamia integrifolia), almond (Prunus amygdalus), sugar beets (Beta vulgaris), sugarcane (Saccharum spp.), oats, barley, vegetables, ornamentals, and conifers; duckweed (Lemna, see WO 25 00/07210, which includes members of the family Lemnaceae. There are known four genera and 34 species of duckweed as follows: genus Lemna (L. aequinoctialis, L. disperma, L. ecuadoriensis, L. gibba, L. japonica, L. minor, L. miniscula, L. obscura, L. perpusilla, L. tenera, L. trisulca, L. turionifera, L. valdiviana); genus Spirodela (S. intermedia, S. polyrrhiza, S. punctata); genus . 30 Woffia (Wa. angusta, Wa. arrhiza, Wa. australina, Wa. borealis, Wa. brasiliensis, Wa. columbiana, Wa. elongata, Wa. globosa, Wa. microscopica, Wa. neglecta) and genus Wofiella (W1. caudata, W1. denticulata, W1. gladiata, W1. hyalina, W1. lingulata, W1. repunda, W1. rotunda, and W1. neotropica).

Any other genera or species of Lemnaceae, if they exist, are also aspects of the present invention. Lemna gibba, Lemna minor, and Lemna miniscula are preferred, with Lemna minor and Lemna miniscula being most preferred. Lemna species can be classified using the taxonomic scheme described by Landolt, Biosystematic Investigation on the Family of Duckweeds: The family of Lemnaceae - A Monograph Study. Geobatanischen Institut ETH, Stiftung Rubel, Zurich (1986)); vegetables including tomatoes (Lycopersicon esculentum), lettuce (e.g., Lactuca sativa), green beans (Phaseolus vulgaris), lima beans (Phaseolus limensis), peas (Lathyrus spp.), and members of the genus Cucumis such as cucumber (C. sativus), cantaloupe (C. cantalupensis), and musk melon (C. melo). Ornamentals include azalea (Rhododendron spp.), hydrangea (Macrophylla hydrangea), hibiscus (Hibiscus rosasanensis), roses (Rosa spp.), tulips (Tulipa spp.), daffodils (Narcissus spp.), petunias (Petunia hybrida), carnation (Dianthus caryophyllus), poinsettia (Euphorbia pulcherrima), and chrysanthemum. Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (Pinus taeda), slash pine (Pinus elliotii), ponderosa pine (Pinus ponderosa), lodgepole pine (Pinus contorta), and Monterey pine (Pinus radiata), Douglas-fir (Pseudotsuga menziesii); Western hemlock (Tsuga canadensis); Sitka spruce (Picea glauca); redwood (Sequoia sempervirens); true firs such as silver fir (Abies amabilis) and balsam fir (Abies balsamea); and cedars such as Western red cedar (Thuja plicata) and Alaska yellow-cedar (Chamaecyparis nootkatensis). Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, chickpea, etc. Legumes include, but are not limited to, Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, Acacia, aneth, artichoke, arugula, blackberry, canola, cilantro, clementines, escarole, eucalyptus, fennel, grapefruit, honey dew, jicama, kiwifruit, lemon, lime, mushroom, nut, okra, orange, parsley, persimmon, plantain, pomegranate, poplar, radiata pine, radicchio, Southern pine, sweetgum, tangerine, triticale, vine, yams, apple, pear, quince, cherry, apricot, melon, hemp, buckwheat, grape, raspberry, chenopodium, blueberry,

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nectarine, peach, plum, strawberry, watermelon, eggplant, pepper, caluliflower, Brassica, e.g., broccoli, cabbage, brussels sprouts, onion, carrot, leek, beet, broad bean, celery, radish, pumpkin, endive, gourd, garlic, snapbean, spinach, squash, turnip, asparagus, and zucchini and ornamental plants include impatiens, Begonia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula, Saint Paulia, Agertum, Amaranthus, Antihirrhinum, Aquilegia, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossos, and Zinnia. Other vegetables are in Table 1.

Preferred forage and turf grass for use in the methods of the invention include alfalfa, orchard grass, tall fescue, perennial ryegrass, creeping bent grass,—and-redtop.—

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Preferably, plants of the present invention are crop plants and in particular cereals (for example, corn, alfalfa, sunflower, rice, *Brassica*, canola, soybean, barley, soybean, sugarbeet, cotton, safflower, peanut, sorghum, wheat, millet, tobacco, and the like), and even more preferably rice, corn and soybean.

In a preferred embodiment, the host cells are monocot or dicot cells, including, but are not limited to, wheat, corn (maize), rice, oat, barley, millet, rye, rape and alfalfa, as well as asparagus, tomato, egg plant, apple, pear, quince, cherry, apricot, pepper, melon, lettuce, cauliflower, Brassica, e.g., broccoli, cabbage, brussels sprout, sugar beet, sugar cane, sweetcorn, onion, carrot, leek, cucumber, tobacco, aubergine, beet, broad bean, carrot, celery, chicory, cotton, radish, pumpkin, hemp, buckwheat, orchardgrass, creeping bent top, redtop, ryegrass, tobacco, turfgrass, tall fescue, cow pea, endive, gourd, grape, raspberry, chenopodium, blueberry, pineapple, avocado, mango, banana, groundnut, nectarine, papaya, garlic, pea, peach, peanut, pepper, pineapple, plum, potato, safflower, snap bean, spinach, squashes, strawberry, sunflower, sorghum, sweet potato, turnip, watermelon, legumes such as Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, and the like; and ornamental crops including Impatiens, Begonia, Petunia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula, Saint Paulia, Ageratum, Amaranthus, Anthirrhinum, Aquilegia,

Chrysanthemum, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossis, Zinnia, and the like. More preferably, the host cells are monocot cells such as maize, rice, wheat, barley, oats, and sorghum, which can be regenerated into a transgenic plant.

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Any plant tissue capable of subsequent clonal propagation, whether by organogenesis or embryogenesis, may be transformed with a vector of the present invention. The term "organogenesis," as used herein, means a process by which shoots and roots are developed sequentially from meristematic centers; the term "embryogenesis," as used herein, means a process by which shoots and roots develop together in a concerted fashion (not sequentially), whether from somatic cells or gametes. The particular tissue chosen will vary depending on the clonal propagation systems available for, and best suited to, the particular species being transformed. Exemplary tissue targets include leaf disks, pollen, embryos, cotyledons, hypocotyls, megagametophytes, callus tissue, existing meristematic tissue (e.g., apical meristems, axillary buds, and root meristems), and induced meristem tissue (e.g., cotyledon meristem and hypocotyl meristem).

The choice of plant tissue source for transformation will depend on the nature of the host plant and the transformation protocol. Useful tissue sources include callus, suspension culture cells, protoplasts, leaf segments, stem segments, tassels, pollen, embryos, hypocotyls, tuber segments, meristematic regions, and the like. The tissue source is selected and transformed so that it retains the ability to regenerate whole, fertile plants following transformation, i.e., contains totipotent cells. Type I or Type II embryonic maize callus and immature embryos are preferred *Zea mays* tissue sources. Selection of tissue sources for transformation of monocots is described in detail in U.S. Application Serial No. 08/112,245 and PCT publication WO 95/06128 (incorporated herein by reference).

For certain plant species, different antibiotic or herbicide selection markers may be preferred. Selection markers used routinely in transformation include the nptII gene which confers resistance to kanamycin and related antibiotics (Messing & Vierra, 1982); Bevan et al., 1983), the *bar* gene which confers resistance to the herbicide phosphinothricin (White et al., 1990, Spencer et al., 1990), the *hph* gene which confers resistance to the antibiotic hygromycin

(Blochinger & Diggelmann), and the *dhfr* gene, which confers resistance to methotrexate (Bourouis et al., 1983).

Thus, the present invention also provides a transformed (transgenic) plant cell, in planta or ex planta, including, but not limited to, a transformed plant cell from plants such as corn (Zea mays), Brassica sp. (e.g., B. napus, B. rapa, B. 5 juncea), particularly those Brassica species useful as sources of seed oil, alfalfa (Medicago sativa), rice (Oryza sativa), rye (Secale cereale), sorghum (Sorghum bicolor, Sorghum vulgare), millet (e.g., pearl millet (Pennisetum glaucum), proso millet (Panicum miliaceum), foxtail millet (Setaria italica), finger millet 10 (Eleusine coracana)), sunflower (Helianthus annuus), safflower (Carthamus tinctorius), wheat (Triticum aestivum), soybean (Glycine max), tobacco (Nicotiana tabacum), potato (Solanum tuberosum), peanuts (Arachis hypogaea), cotton (Gossypium barbadense, Gossypium hirsutum), sweet potato (Ipomoea batatus), cassava (Manihot esculenta), coffee (Cofea spp.), coconut (Cocos 15 nucifera), pineapple (Ananas comosus), citrus trees (Citrus spp.), cocoa (Theobroma cacao), tea (Camellia sinensis), banana (Musa spp.), avocado (Persea americana), fig (Ficus casica), guava (Psidium guajava), mango (Mangifera indica), olive (Olea europaea), papaya (Carica papaya), cashew (Anacardium occidentale), macadamia (Macadamia integrifolia), almond (Prunus amygdalus), sugar beets (Beta vulgaris), sugarcane (Saccharum spp.), 20 oats, barley, vegetables, ornamentals, and conifers; duckweed (Lemna, see WO 00/07210, which includes members of the family Lemnaceae. There are known four genera and 34 species of duckweed as follows: genus Lemna (L. aequinoctialis, L. disperma, L. ecuadoriensis, L. gibba, L. japonica, L. minor, L. 25 miniscula, L. obscura, L. perpusilla, L. tenera, L. trisulca, L. turionifera, L. valdiviana); genus Spirodela (S. intermedia, S. polyrrhiza, S. punctata); genus Woffia (Wa. angusta, Wa. arrhiza, Wa. australina, Wa. borealis, Wa. brasiliensis, Wa. columbiana, Wa. elongata, Wa. globosa, Wa. microscopica, Wa. neglecta) and genus Wofiella (W1. caudata, W1. denticulata, W1. gladiata, 30 W1. hyalina, W1. lingulata, W1. repunda, W1. rotunda, and W1. neotropica). Any other genera or species of Lemnaceae, if they exist, are also aspects of the present invention. Lemna gibba, Lemna minor, and Lemna miniscula are preferred, with Lemna minor and Lemna miniscula being most preferred. Lemna species can be classified using the taxonomic scheme described by Landolt,

Biosystematic Investigation on the Family of Duckweeds: The family of Lemnaceae - A Monograph Study. Geobatanischen Institut ETH, Stiftung Rubel, Zurich (1986)); vegetables including tomatoes (Lycopersicon esculentum), lettuce (e.g., Lactuca sativa), green beans (Phaseolus vulgaris), lima beans 5 (Phaseolus limensis), peas (Lathyrus spp.), and members of the genus Cucumis such as cucumber (C. sativus), cantaloupe (C. cantalupensis), and musk melon (C. melo). Ornamentals include azalea (Rhododendron spp.), hydrangea (Macrophylla hydrangea), hibiscus (Hibiscus rosasanensis), roses (Rosa spp.), tulips (Tulipa spp.), daffodils (Narcissus spp.), petunias (Petunia hybrida), 10 carnation (Dianthus caryophyllus), poinsettia (Euphorbia pulcherrima), and chrysanthemum. Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (Pinus taeda), slash pine (Pinus elliotii), ponderosa pine (Pinus ponderosa), lodgepole pine (Pinus contorta), and Monterey pine (Pinus radiata), Douglas-fir (Pseudotsuga 15 menziesii); Western hemlock (Tsuga canadensis); Sitka spruce (Picea glauca); redwood (Sequoia sempervirens); true firs such as silver fir (Abies amabilis) and balsam fir (Abies balsamea); and cedars such as Western red cedar (Thuja plicata) and Alaska yellow-cedar (Chamaecyparis nootkatensis). Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, chickpea, etc. Legumes include, but are not limited to, Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, Acacia, aneth, artichoke, arugula, blackberry, canola, cilantro, clementines, escarole, eucalyptus, fennel, grapefruit, honey dew, jicama, kiwifruit, lemon, lime, mushroom, nut, okra, orange, parsley, persimmon, plantain, pomegranate, poplar, radiata pine, radicchio, Southern pine, sweetgum, tangerine, triticale, vine, yams, apple, pear, quince, cherry, apricot, melon, hemp, buckwheat, grape, raspberry, chenopodium, blueberry, nectarine, peach, plum, strawberry, watermelon, eggplant, pepper, caluliflower, Brassica, e.g., broccoli, cabbage, brussels sprouts, onion, carrot, leek, beet, broad bean, celery, radish, pumpkin, endive, gourd, garlic, snapbean, spinach, squash, turnip, asparagus, and zucchini and ornamental plants include impatiens,

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Begonia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula, Saint Paulia, Agertum, Amaranthus, Antihirrhinum, Aquilegia, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossos, and Zinnia, as well as from vegetables including those described in Table 1.

In a preferred embodiment, the transformed cells, include, but are not limited to, transformed wheat, corn (maize), rice, oat, barley, millet, rye, rape and alfalfa, as well as asparagus, tomato, egg plant, apple, pear, quince, cherry, apricot, pepper, melon, lettuce, cauliflower, Brassica, e.g., broccoli, cabbage, brussels sprout, sugar beet, sugar cane, sweetcorn, onion, carrot, leek, cucumber, tobacco, aubergine, beet, broad bean, carrot, celery, chicory, cotton, radish, pumpkin, hemp, buckwheat, orchardgrass, creeping bent top, redtop, ryegrass, tobacco, turfgrass, tall fescue, cow pea, endive, gourd, grape, raspberry, chenopodium, blueberry, pineapple, avocado, mango, banana, groundnut, nectarine, papaya, garlic, pea, peach, peanut, pepper, pineapple, plum, potato, safflower, snap bean, spinach, squashes, strawberry, sunflower, sorghum, sweet potato, turnip, watermelon, legumes such as Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, and the like; and ornamental crops including Impatiens, Begonia, Petunia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula, Saint Paulia, Ageratum, Amaranthus, Anthirrhinum, Aquilegia, Chrysanthemum, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossis, Zinnia, and the like. Preferably, the host cells are monocot cells such as maize, rice, wheat, barley, oats, and sorghum, which can be regenerated into a transgenic plant.

# 30 IV. Identification of Transgenic Plants

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To confirm the presence of the preselected nucleic acid segment(s) or "transgene(s)" in the regenerating plants, a variety of assays may be performed. Such assays include, for example, "molecular biological" assays well known to those of skill in the art, such as Southern and Northern blotting, in situ

hybridization and nucleic acid-based amplification methods such as PCR or RT-PCR; "biochemical" assays, such as detecting the presence of a protein product, e.g., by immunological means (ELISAs and Western blots) or by enzymatic function; plant part assays, such as leaf or root assays; and also, by analyzing the phenotype of the whole regenerated plant, e.g., for disease or pest resistance.

DNA may be isolated from cell lines or any plant parts to determine the presence of the preselected nucleic acid segment through the use of techniques well known to those skilled in the art. Note that intact sequences will not always be present, presumably due to rearrangement or deletion of sequences in the cell.

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The presence of nucleic acid elements introduced through the methods of this invention may be determined by polymerase chain reaction (PCR). Using this technique discreet fragments of nucleic acid are amplified and detected by gel electrophoresis. This type of analysis permits one to determine whether a preselected nucleic acid segment is present in a stable transformant, but does not prove integration of the introduced preselected nucleic acid segment into the host cell genome. In addition, it is not possible using PCR techniques to determine whether transformants have exogenous genes introduced into different sites in the genome, i.e., whether transformants are of independent origin. It is contemplated that using PCR techniques it would be possible to clone fragments of the host genomic DNA adjacent to an introduced preselected DNA segment.

Positive proof of DNA integration into the host genome and the independent identities of transformants may be determined using the technique of Southern hybridization. Using this technique specific DNA sequences that were introduced into the host genome and flanking host DNA sequences can be identified. Hence the Southern hybridization pattern of a given transformant serves as an identifying characteristic of that transformant. In addition it is possible through Southern hybridization to demonstrate the presence of introduced preselected DNA segments in high molecular weight DNA, i.e., confirm that the introduced preselected DNA segment has been integrated into the host cell genome. The technique of Southern hybridization provides information that is obtained using PCR, e.g., the presence of a preselected DNA segment, but also demonstrates integration into the genome and characterizes each individual transformant.

It is contemplated that using the techniques of dot or slot blot hybridization which are modifications of Southern hybridization techniques one could obtain the same information that is derived from PCR, e.g., the presence of a preselected DNA segment.

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Both PCR and Southern hybridization techniques can be used to demonstrate transmission of a preselected DNA segment to progeny. In most instances the characteristic Southern hybridization pattern for a given transformant will segregate in progeny as one or more Mendelian genes (Spencer et al., 1992); Laursen et al., 1994) indicating stable inheritance of the gene. The nonchimeric nature of the callus and the parental transformants (R<sub>0</sub>) was suggested by germline transmission and the identical Southern blot hybridization patterns and intensities of the transforming DNA in callus, R<sub>0</sub> plants and R<sub>1</sub> progeny that segregated for the transformed gene.

Whereas DNA analysis techniques may be conducted using DNA isolated from any part of a plant, RNA may only be expressed in particular cells or tissue types and hence it will be necessary to prepare RNA for analysis from these tissues. PCR techniques may also be used for detection and quantitation of RNA produced from introduced preselected DNA segments. In this application of PCR it is first necessary to reverse transcribe RNA into DNA, using enzymes such as reverse transcriptase, and then through the use of conventional PCR techniques amplify the DNA. In most instances PCR techniques, while useful, will not demonstrate integrity of the RNA product. Further information about the nature of the RNA product may be obtained by Northern blotting. This technique will demonstrate the presence of an RNA species and give information about the integrity of that RNA. The presence or absence of an RNA species can also be determined using dot or slot blot Northern hybridizations. These techniques are modifications of Northern blotting and will only demonstrate the presence or absence of an RNA species.

While Southern blotting and PCR may be used to detect the preselected DNA segment in question, they do not provide information as to whether the preselected DNA segment is being expressed. Expression may be evaluated by specifically identifying the protein products of the introduced preselected DNA segments or evaluating the phenotypic changes brought about by their expression.

Assays for the production and identification of specific proteins may make use of physical-chemical, structural, functional, or other properties of the proteins. Unique physical-chemical or structural properties allow the proteins to be separated and identified by electrophoretic procedures, such as native or denaturing gel electrophoresis or isoelectric focussing, or by chromatographic techniques such as ion exchange or gel exclusion chromatography. The unique structures of individual proteins offer opportunities for use of specific antibodies to detect their presence in formats such as an ELISA assay. Combinations of approaches may be employed with even greater specificity such as Western blotting in which antibodies are used to locate individual gene products that have been separated by electrophoretic techniques. Additional techniques may be employed to absolutely confirm the identity of the product of interest such as evaluation by amino acid sequencing following purification. Although these are among the most commonly employed, other procedures may be additionally used.

Assay procedures may also be used to identify the expression of proteins by their functionality, especially the ability of enzymes to catalyze specific chemical reactions involving specific substrates and products. These reactions may be followed by providing and quantifying the loss of substrates or the generation of products of the reactions by physical or chemical procedures. Examples are as varied as the enzyme to be analyzed.

Very frequently the expression of a gene product is determined by evaluating the phenotypic results of its expression. These assays also may take many forms including but not limited to analyzing changes in the chemical composition, morphology, or physiological properties of the plant.

Morphological changes may include greater stature or thicker stalks. Most often changes in response of plants or plant parts to imposed treatments are evaluated under carefully controlled conditions termed bioassays.

#### 30 V. Utility

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Once an expression cassette of the invention has been transformed into a particular plant species, it may be propagated in that species or moved into other varieties of the same species, particularly including commercial varieties, using traditional breeding techniques. Particularly preferred plants of the invention

include the agronomically important crops listed above. The genetic properties engineered into the transgenic seeds and plants described above are passed on by sexual reproduction and can thus be maintained and propagated in progeny plants. The present invention also relates to a transgenic plant cell, tissue, organ, seed or plant part obtained from the transgenic plant. Also included within the invention are transgenic descendants of the plant as well as transgenic plant cells, tissues, organs, seeds and plant parts obtained from the descendants.

Preferably, the expression cassette in the transgenic plant is sexually transmitted. In one preferred embodiment, the coding sequence is sexually transmitted through a complete normal sexual cycle of the R0 plant to the R1 generation. Additionally preferred, the expression cassette is expressed in the cells, tissues, seeds or plant of a transgenic plant in an amount that is different than the amount in the cells, tissues, seeds or plant of a plant which only differs in that the expression cassette is absent.

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The transgenic plants produced herein are thus expected to be useful for a variety of commercial and research purposes. Transgenic plants can be created for use in traditional agriculture to possess traits beneficial to the grower (e.g., agronomic traits such as resistance to water deficit, pest resistance, herbicide resistance or increased yield), beneficial to the consumer of the grain harvested from the plant (e.g., improved nutritive content in human food or animal feed), or beneficial to the food processor (e.g., improved processing traits). In such uses, the plants are generally grown for the use of their grain in human or animal foods. However, other parts of the plants, including stalks, husks, vegetative parts, and the like, may also have utility, including use as part of animal silage or for ornamental purposes. Often, chemical constituents (e.g., oils or starches) of maize and other crops are extracted for foods or industrial use and transgenic plants may be created which have enhanced or modified levels of such components.

Transgenic plants may also find use in the commercial manufacture of proteins or other molecules, where the molecule of interest is extracted or purified from plant parts, seeds, and the like. Cells or tissue from the plants may also be cultured, grown in vitro, or fermented to manufacture such molecules.

The transgenic plants may also be used in commercial breeding programs, or may be crossed or bred to plants of related crop species. Improvements

encoded by the expression cassette may be transferred, e.g., from maize cells to cells of other species, e.g., by protoplast fusion.

The transgenic plants may have many uses in research or breeding, including creation of new mutant plants through insertional mutagenesis, in order to identify beneficial mutants that might later be created by traditional mutation and selection. An example would be the introduction of a recombinant DNA sequence encoding a transposable element that may be used for generating genetic variation. The methods of the invention may also be used to create plants having unique "signature sequences" or other marker sequences which can be used to identify proprietary lines or varieties.

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Thus, the transgenic plants and seeds according to the invention can be used in plant breeding which aims at the development of plants with improved properties conferred by the expression cassette, such as tolerance of viruses or other pests, or other stresses. The various breeding steps are characterized by well-defined human intervention such as selecting the lines to be crossed, directing pollination of the parental lines, or selecting appropriate descendant plants. Depending on the desired properties different breeding measures are taken. The relevant techniques are well known in the art and include but are not limited to hybridization, inbreeding, backcross breeding, multiline breeding, variety blend, interspecific hybridization, aneuploid techniques, etc. Hybridization techniques also include the sterilization of plants to yield male or female sterile plants by mechanical, chemical or biochemical means. Cross pollination of a male sterile plant with pollen of a different line assures that the genome of the male sterile but female fertile plant will uniformly obtain properties of both parental lines. Thus, the transgenic seeds and plants according to the invention can be used for the breeding of improved plant lines which for example increase the effectiveness of conventional methods such as herbicide or pesticide treatment or allow to dispense with said methods due to their modified genetic properties. Alternatively new crops with improved stress tolerance can be obtained which, due to their optimized genetic "equipment", yield harvested product of better quality than products which were not able to tolerate comparable adverse developmental conditions.

### VI. A Computer Readable Medium

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The invention also provides a computer readable medium having stored thereon a data structure containing nucleic acid sequences having at least 70% sequence identity to a nucleic acid sequence selected from those listed in SEQ ID Nos: 1-795 as well as complementary, ortholog, and variant sequences thereof. Storage and use of nucleic acid sequences on a computer readable medium is well known in the art. (See for example U.S. Patent Nos. 6,023,659; 5,867,402; 5,795,716) Examples of such medium include, but are not limited to, magnetic tape, optical disk, CD-ROM, random access memory, volatile memory, nonvolatile memory and bubble memory. Accordingly, the nucleic acid sequences contained on the computer readable medium may be compared through use of a module that receives the sequence information and compares it to other sequence information. Examples of other sequences to which the nucleic acid sequences of the invention may be compared include those maintained by the National Center for Biotechnology Information (NCBI)(http://www.ncbi.nlm.nih.gov/) and the Swiss Protein Data Bank. A computer is an example of such a module that can read and compare nucleic acid sequence information. Accordingly, the invention also provides the method of comparing a nucleic acid sequence of the invention to another sequence. For example, a sequence of the invention may be submitted to the NCBI for a Blast search as described herein where the sequence is compared to sequence information contained within the NCBI database and a comparison is returned. The invention also provides nucleic acid sequence information in a computer readable medium that allows the encoded polypeptide to be optimized for a desired property. Examples of such properties include, but are not limited to, increased or decreased: thermal stability, chemical stability, hydrophylicity, hydrophobicity, and the like. Methods for the use of computers to model polypeptides and polynucleotides having altered activities are well known in the art and have been reviewed. (Lesyng et al., 1993; Surles et al., 1994; Koehl et al., 1996; Rossi et al., 2001).

The invention will be further described by the following examples which is not intended to limit the scope of the invention.

#### Example 1

#### GeneChip Standard Protocol

## Quantitation of total RNA

Total RNA from plant tissue is extracted and quantified.

5 1. Quantify total RNA using GeneQuant

1OD<sub>260</sub>=40 mg RNA/ml; A260/A280=1.9 to about 2.1

2. Run gel to check the integrity and purity of the extracted RNA

#### Synthesis of double-stranded cDNA

10 Gibco/BRL SuperScript Choice System for cDNA Synthesis (Cat#1B090-019)

was employed to prepare cDNAs. T7-(dT)<sub>24</sub> oligonucleotides were prepared

and purified by HPLC. (5'-

15 GGCCAGTGAATTGTAATACGACTCACTATAGGGAGGCGG-(dT)<sub>24</sub>-3'(SEQ ID NO:800).

#### Step 1. Primer hybridization:

Incubate at 70°C for 10 minutes

Quick spin and put on ice briefly

20 <u>Step 2. Temperature adjustment:</u>

I Incubate at 42°C for 2 minutes

#### Step 3. First strand synthesis:

DEPC-water- 1 µ1

RNA (10 μg final)-10 μl

25 T7=(dT)<sub>24</sub> Primer (100 pmol final)-1 μl pmol

5X 1st strand cDNA buffer-4 µl

0.1M DTT (10 mM final)- 2 μl

10 mM dNTP mix (500 µM final)-1 µl

Superscript II RT 200 U/µl- 1 µl

30 Total of 20 μl

Mix well

Incubate at 42°C for 1 hour

#### Step 4. Second strand synthesis:

Place reactions on ice, quick spin

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DEPC-water- 91 μl

5X 2nd strand cDNA buffer- 30 μl

mM dNTP mix (250 mM final) - 3 μl

E. coli DNA ligase (10 U/μl)-1 μl

E. coli DNA polymerase 1-10 U/μl- 4 μl

RnaseH 2U/μl -1 μl

T4 DNA polymerase 5 U/μl-2 μl

0.5 M EDTA (0.5 M final)--10 μl

Total 162 ul

10 Mix/spin down/incubate 16°C for 2 hours

## Step 5. Completing the reaction:

Incubate at 16°C for 5 minutes

#### Purification of double stranded cDNA

- Centrifuge PLG (Phase Lock Gel, Eppendorf 5 Prime, Inc., PI-188233)
   at 14,000X, transfer 162 μl of cDNA to PLG
  - 2. Add 162 μl of Phenol:Chloroform:Isoamyl alcohol (pH 8.0), centrifuge 2 minutes
  - 3. Transfer the supernatant to a fresh 1.5 ml tube, add
- 20 Glycogen (5 mg/ml)

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0.5 M NH4OAC (0.75xVol)

120

ETOH (2.5xVol, -20 °C)

400

- 4. Mix well and centrifuge at 14,000X for 20 minutes
- 5. Remove supernatant, add 0.5 ml 80% EtOH (-20°C)
- 25 6. Centrifuge for 5 minutes, air dry or by speed vac for 5-10 minutes
  - 7. Add 44 μl DEPC H<sub>2</sub>O

Analyze of quantity and size distribution of cDNA Run a gel using 1 µl of the double-stranded synthesis product

# 30 Synthesis of biotinylated cRNA

(use Enzo BioArray High Yield RNA Transcript Labeling Kit Cat#900182)

Purified cDNA

22 µl

10X Hy buffer

4 µl

10X biotin ribonucleotides

 $4 \mu l$ 

10X DTT
4 μl
10X Rnase inhibitor mix
4 μl
20X T7 RNA polymerase
2 μl

Total
40 μl

Centrifuge 5 seconds, and incubate for 4 hours at 37°C
Gently mix every 30-45 minutes

## Purification and quantification of cRNA

(use Qiagen Rneasy Mini kit Cat# 74103)

10 Determine concentration and dilute to 1 μg/μl concentration

#### Fragmentation of cRNA

cRNA (1 μg/μl) 15 μl 5X Fragmentation Buffer\* 6 μl 15 DEPC  $H_2O$  9 μl 30 μl

### \*5x Fragmentation Buffer

1M Tris (pH8.1) 4.0 ml

20 MgOAc

0.64 g

KOAC

0.98 g

DEPC H<sub>2</sub>O

Total

20 ml

Filter Sterilize

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#### Array wash and staining

Stringent Wash Buffer\*\*

Non-Stringent Wash Buffer\*\*\*

SAPE Stain\*\*\*\*

30 Antibody Stain\*\*\*\*

Wash on fluidics station using the appropriate antibody amplification protocol \*\*Stringent Buffer: 12X MES 83.3 ml, 5 M NaCl 5.2 ml, 10% Tween 1.0 ml, H<sub>2</sub>O 910 ml,

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Filter Sterilize

\*\*\*Non-Stringent Buffer: 20X SSPE 300 ml, 10% Tween 1.0 ml, H<sub>2</sub>O 698 ml, Filter Sterilize, Antifoam 1.0.

\*\*\*\*SAPE stain: 2X Stain Buffer 600 μl, BSA 48 μl, SAPE 12μl, H<sub>2</sub>O 540 μl.

\*\*\*\*Antibody Stain: 2X Stain Buffer 300 μl, H<sub>2</sub>O 266.4 μl, BSA 24 ul, Goat
IgG 6 μl, Biotinylated Ab 3.6 μl

#### Example 2

## Identification of Arabidopsis Genes Induced by Peronospora Infection

To define the transcriptional profile for *Peronospora*-induced and/or *RPP7*- and *RPP8*- dependent genes, four *Arabidopsis* lines were infected with one of two different *P. parasitica* isolates (see Table 2). One *Arabidopsis* line was Col-0::*RPP8* which carries the cloned *RPP8* genomic clone (*RPP8* mediates resistance of the ecotype La-er against Emco5; McDowell et al. 1998) and is Emco5 resistant. Wild-type Col-0 plants are Emco5 susceptible. *RPP7* mediates resistance of *Arabidopsis* against the *Peronospora* isolate Hiks1, and occurs naturally in Col-0. Another *Arabidopsis* line had a loss of function *rpp7* mutant allele (Col-0 (*rpp7*)) (McDowell et al., 2000). Finally, three mutants in the signaling pathway for RPP7, i.e., they lack a *RPP7*-mediated response, were identified, i.e., *edm1*, *edm2* and *edm3*, and a line with one of the mutant alleles, i.e., *edm1*, was used.

RNA samples were collected from two week old seedlings at three time points, 0, 12 and 24 hours post infection with 100,000 spores/ml. Twelve hours post-inoculation was determined to be an appropriate time point for analysis of both incompatible interactions, based on trypan blue staining of HR sites and PR-1 and sen1 induction. Forty-eight hours post-inoculation was determined to be a time point where both compatible interactions exhibit dense hyphal growth and both PR-1 and sen1 expression. Three independent sets of RNA were pooled.

Table 2

	Plant	Peronospora isolate	Time of RNA collection	
	Col-0 (RPP7, incomp.)	Hiksl	0, 12, 48 hours	
35	Col-0 (rpp7, comp.)	Hiks1	0, 12, 48 hours	

 Col-0 (edm1, comp.)
 Hiks1
 0, 12, 48 hours

 Col-0 (tgRPP8, incomp.)
 Emco5
 0, 12, 48 hours

 Col-0 (rpp8, comp.)
 Emco5
 0, 12, 48 hours

PCT/US01/28506

- RNAs were labeled as described in Example 1 and hybridized to an Affymetrix Gene Chip having sequences corresponding to about 8,200 *Arabidopsis* genes. The data collected from these chips was employed to determine:
  - 1) the difference, if any, between RPP7 and RPP8 triggered signaling (and how these profiles compare to other profiles, for example, to the profiles of RPM1 and RPS2);
  - 2) which portion of the transcriptional response is R dependent;
  - which genes are either commonly or uniquely induced during two different compatible interactions;
- 4) which genes are specifically activated during the incompatible interactions
   15 (gleaned by comparison with isogenic compatible controls) and, hence, may be important for the plant's defense against *Peronospora*;
  - which genes whose expression is a prerequisite for induced defense reactions against *Peronospora*; and
  - 6) which genes are specifically repressed during the incompatible interactions, which may be required for successful Oomycete infections.

#### Results

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### Genes Expressed Early and Transiently After Infection

The normalized "average difference" (AD) from the chip experiments were processed as follows. All values together with the respective gene identifiers were loaded in EXCEL. Genes having, for a given treatment, AD values of 25 or less were considered as being not expressed. To reduce spurious background fluctuation, this cut off level was elevated to 75 for most of the analyses and normalized all AD values of 75 or less. For all five infection time courses, the AD values were divided by the corresponding 0 hour time point AD value. To visualize differences in gene expression ground states between the four experimental *Arabidopsis* lines, the following ratios were calculated for the 0 hour AD values: Col-0 (from Emco5 series)/Col-0-RPP8 and rpp7/Col-0 (from Hiks1 series) and edm1/Col-0 (from Hiks1 series). These ratios indicate whether

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a given gene is up- or down-regulated by the RPP7 or RPP8 pathway, respectively. The resulting set of 18 expression ratio series was then loaded into CLUSTER (Eisen et al., 1998). The AD values were log transformed (base 2) and filtered to include only genes in the final data set that showed at least a 3-fold expression change (which equals a 1.6-fold change of the log transformed values). These criteria were fulfilled by 464 genes that define the final data set. With this data set Average linkage clustering was performed (uncentered correlation). The resulting clustergram was displayed using TREEVIEW (Eisen et al., 1998). A second clustergram was generated in the same way applying a cut off value of 50 on the normalized AD values. This clustergram includes 619 genes (probe sets).

Based on these clustergrams, gene clusters showing distinct expression behaviors were defined. Cluster cR7/R8 (see Table 3) contained genes that are early and transiently upregulated early and transiently after *P. parasitica* infection of *Arabidopsis* in an *RPP7/8* dependent manner. Upregulation by the *RPP8* pathway was generally stronger. Strong induced expression of cR7/R8 genes was detectable 12 hours after Emco5 infection in Col-0(tgRPP8) plants (incompatible interaction), whereas no elevated expression can be observed 48 hours post infection in the same plants (as compared to untreated control plants). Only weakly elevated or no elevated expression was detected at all tested time points in Col-0 plants (compatible interaction). After Hiks1 infection, elevated expression of these genes was only observed in Col-0 plants (incompatible interaction) after 12 hours. At no other time point following Hiks1 infection was substantially elevated expression of these genes observed. The expression ground state of these genes is similar in all four *Arabidopsis* lines.

#### Table 3

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20245 s at cut off = 75
18716 At
18022 at
14609 at
17014 s at
17051 s at
19640 at
14248 at
13176\_at

	•
15978 at	cut off +50
17014 s at	
14609 at	
13764 at	
16649 s at	Subcluster A
13215 s at	
17653 at	
17008 at	
15042 at	
12778_r_at	
14614 at	
17051 sat	
14248 at	Subcluster B
20245 s at	
19640 at	
18716 at	
18022_at	•
17500 s at	•
13217 s at	-
18928 at	

The cR7/R8 cluster was originally defined using the clustergram based on a cut off value of 75 and comprised only 9 genes. Applying a cut off value of 50, the cluster became larger, comprising 21 genes, which could be divided into two subclusters (Table 4). Subcluster cR7/R8 A includes genes strongly and transiently upregulated by the RPP8 pathway, but either not, or only faintly, upregulated by the RPP7 pathway. Members of subcluster B showed pronounced upregulation by both RPP pathways after 12 hours and also elevated expression during compatible Emco5 infections in Col-0.

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#### Table 4

Subcluster A	
Gene-identifier (probe set)	Related to
15978_at	X68592.6 at
17014_s_at*	atu05206 s at
14609_at -	AC002340.147 at
16649_s_at+	athorf s at
	CafferoylCoAmethyltrans s a
13215_s_at <sup>+</sup>	t
17653_at	AL035679.144 at
17008_at	AC006585.212 at

AL021961.3 at

	11D021701.5_at
12778_r_at	AC006577.16_r_at
Subcluster B Gene-identifier (probe set) 14614_at 17051_s_at 14248_at 20245_s_at 19640_at 18716_at 18022_at 17500_s_at 13217_s_at 18928_at	Related to AC004165.66_at af098947_s_at PAD3_at AC005309.97_s_at AC004561.78_at X91916_at AJ010971_at athcallga_s_at calmodulinlike_s_at AC002333.181_at

<sup>\*</sup>Genes printed in **bold** were identified using a cut off value of 75; all others were identified using a cut off value of 50.

†The probe sets 16649 and 13215 as well as 17500 and 13217 correspond to

the same genes.

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15042 at

Thus, the predominant feature of genes of the cR7/R8 cluster is more pronounced expression along with RPP7 and RPP8 mediated Peronospora resistance in comparison to the corresponding compatible interactions. Expression of these genes peaks in around 12 hours post infection and returns to ground state by 48 hours post infection. Hence, increased transcript levels of these genes coincide with the time window of decision between disease or resistance. Within 48 hours after infection with each tested Peronospora isolate, the plant either responds with HR and hinders development of oomycete hyphae or allows successful infection.

# Identification of Conserved Sequence Motifs Within cR7/R8 Upstream Regions

To identify potential *cis*-elements involved in the specific regulation of cR7/R8 genes, stretches comprising 1.1 to 1.2 kb localized directly upstream of the translation start sites of coding regions from this cluster were collected from genomic *Arabidopsis* sequences (see Figure 2). Two motif search algorithms were used to recognize conserved motifs: AlignAce (Roth et al., 1998) and MEME (Bailey and Elkan, 1994). A search with AlignAce (default settings) revealed two conserved motifs within a set of 8 promoters comprising 7 from the originally defined cR7R8 using a cut off value of 75 and the promoter of a gene encoding a putative Myb-like transcription factor (Martin &Paz-Ares, 1997)

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(probe set 13176, SEQ ID NO:36), which shows a similar expression behavior as cR7/R8. The AlignAce output is shown below in Table 5:

Table 5

Motif 1	MAP Score:	SEQ ID NO.	Corresponding
** * * * * * * *	6.87857	÷.	Probe Set
CAACAATTAT	0 36 1	685	17014
CAACTATGTC	0 981 1	686	17014
CAACAATGAC	1 934 0	687	20245
CAACAATGAC	2 271 1	688	14609
AAACAATGAT	2 1184 1	689	14609
CAACATTGAC	3 163 0	690	17051
CAACATTGTT	3 298 0	691	17051
CAACAATGTT	4 613 1	692	19640
CAACTATGAT	4 791 0	693	19640
CAACAATGAT	4 1064 0	694	19640
CAAAAATGAT	7 397 0	695	17008
CAACATTGAT	7 415 1	696	17008
CATATCA/TA/TTGAC (Consensus)		697	N.A.

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Motif 2  **** * * **	MAP Score: 2.22382	SEQ ID NO.	Corresponding Probe Set
TTTGGGCAAAA	1 368 0	698	20245
GTTGGGTCAAA	1 739 0	699	20245
GATGGGCACAA	2 691 0	700	14609
GATGGGCCAAA	3 700 0	701	17051
GATGGGTATAA	4 344 0	702	19640
GATGGGACTAA	5 98 1	703	14248
TTTGGGCCTAA	5 971 1	704	14248
TATGGGCCCAA	5 1012 0	705	14248
CTTGGGACAAA	7 579 1	706	17008
GATGGGTCAAC	7 1009 0	707	17008
CATGGGAATAA	7 1072 1	708	17008
NATTGGGNC/ANAA (Consensus)		709	N.A.

For each motif, examples from cR7/R8 promoters are listed. Nucleotides evaluated by AlignAce as being conserved are indicated by a "\*" above the examples. Besides motifs 1 and 2, a variety of additional ones were found by AlignAce, which, however, consisted almost exclusively of Ts and As. These motifs probably do not constitute functional cis-elements. Promoter sequences

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are generally enriched in As or Ts (A/T content in cR7R8 promoters is 70%) and regulatory cis-elements usually contain also Cs or Gs.

Motif 1 (CAACA/TA/TTGA/TC/T, SEQ ID NO:710) resembles the type I Myb binding site consensus (MBS;T/CAACT/GG), which is bound by animal and some plant Myb transcription factors (Yang & Klessig, 1996). The 3' portion of the motif 1 consensus allows overlap with binding sites of WRKY (TTTGACC/T; Eulgem et al., 2000) or TGA-bZIP transcription factors (TGACG; Schindler et al. 1992). Interestingly, a gene encoding a putative Myblike transcription factor (gene identification number 13176) is upregulated along with cR7/R8 genes in an RPP7/RPP8-dependent, early and transient manner. The factor encoded by this gene may be a regulator of cR7/R8 genes. Motif 1 is particularly enriched in promoters of the originally defined cR7/R8 cluster (cut off = 75). This motif, which consists of six strictly defined positions and four positions allowing two base pairs each, should occur by random roughly once per 20,000 bp in each of both possible orientations (assuming a T/A content of 70% and a G/C content of 30%. In the 8,441 bp of cR7/R8 promoter sequences, the motif occurs 9 times (roughly 20 times the statistically expected frequency). Consistent with the statistical expectation, motif 1 occurs only two times within 15,873 bp from a set of control promoters of genes showing no defense related expression (data not shown).

In the extended set of cR7/R8 genes (cut off = 50), the enrichment of motif 1 is less striking. It occurs only 9 times within 16,519 bp in total and, more importantly, none of the promoters added to the set by lowering the cut off value contains this motif. Thus, if motif 1 is involved in the specific regulation of cR7/R8 genes it cannot alone be responsible for the particular expression features of these genes, and may rather be responsible for the more pronounced R7/R8-triggered expression of members of the originally defined cluster (cut off = 75).

Motif 2 (A/TTGGGNC/ANAA; SEQ ID NO:711) does not resemble any other known plant *cis*-element. The consensus sequence consists of six strictly defined positions, two positions with two alternatives and two positions with four alternatives; and, hence, should occur by random once per 10,000 bp. Motif 2 can be found 10 times within 8,441 bp of the promoters from the originally defined cR7/R8 (cut off = 75) and 14 times within 16,519 bp of the extended

cR7/R8 promoter set. Thus, its occurrence in cR7/R8 promoters is roughly 10 times the statistical expectation. Motif 2 is absent in the 15,873 bp of promoters of genes showing no defense related expression (data not shown). It is, however, also enriched in the promoters of previously defined clusters of defense related genes, such as the PR1 regulon (SAR genes) and SOMc3 (genes up regulated in Ws-0 after infection with avirulent Noco2; data not shown). The latter two clusters may also include genes sharing expression features with cR7/R8 genes. In addition to AlignAce, a motif 2-like sequence was also recognized by MEME. Within the extended set of cR7/R8 promoters, MEME found the following

10 sequence stretch to be conserved:

	oits 2.2									•
	2.0									
	1.7			•	•					
15	1.5			•	•					
Inform	nation 1.3					•	*	•		
conter	it 1.1					*	*			
(13.41	oits) 0.9		***	**.	*	*		**		
	0.7		***	***	*	*		**	*	
20 .	0.4	**	***	***	*	*		**	*	*
	0.2	**	***	* ***	*	*	*	**	*	*
	0.0					•				

Multilevel Consensus sequence T/C TTT G/C A/T/G C/T

25 CCAAG/A/CAA/GAC/A (SEQ ID NO:712)

> The reverse complement of this consensus sequence is (positions with three alternatives were given as "N") given below with motif 2 for comparison:

30 MEME motif: G/T T C/T T N T T G G G N G/C A A A A/C (SEQ ID

NO:713)

Motif 2: TGGGNAN/CAA(SEQIDNO:714)

MEME was run using the default settings of "advanced MEME" choosing the options "one motif per sequence", "narrow motifs" and "additional strand: reverse complement".

To confirm these results, Northern blots using probes of representatives of each cluster are performed. In addition, T-DNA lines are screened for knock out in Myb-like gene (AL031394.56\_at) and selected members of cR7/R8, such as RNS1 (atu05206\_s\_at) or the gene encoding a calmodulin-like protein (identification number 13217\_s\_at). Also, yeast one hybrid screens are performed to clone transcription factors binding to motifs 1 and 2, or other identified motifs. Transgenic *Arabidopsis* plants carrying fusions of GUS with a representative promoter from cR7/R8 can be used to further evaluate the functional significance of motifs 1 and 2, and to perform a genetic screening for mutants impaired in regulation of cR7/R8 genes.

# 15 Genes Induced by *Peronospora* Infection in a Manner that Requires *RPP7* or *RPP8*

The data sets described above were employed to identify genes that were induced by *Peronospora* infection and were RPP7/8-dependent. All expression values less than 5 were set to 5. Using ratios calculated in Excel, and database sorting using Access, a subset of genes was identified whose expression depends on RPP7 or RPP8 using the following criteria:

Col Hiks1 12 hour/Col untreated > 2.5 AND Col Hiks1 12 hour > 30 AND Col Hiks1 12 hour/Col rpp7 Hiks1 12 hour > 2 OR

Col Hiks1 48 hour/Col untreated > 2.5 AND Col Hiks1 48 hour > 30
AND Col Hiks1 48 hour/Col rpp7 Hiks1 48 hour > 2
OR

Col RPP8 Emco5 12 hour/Col RPP8 no treatment > 2.5 AND Col RPP8 Emco5 12 hour > 30 AND Col RPP8 Emco5 12 hour/Col (rpp8) Emco5

30 12 hour > 2

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OR -

Col RPP8 Emco5 48 hour/Col RPP8 no treatment AND Col RPP8
Emco5 48 hour > 30 AND Col RPP8 Emco5 48 hour/Col (rpp8) Emco5
48 hour > 2

This yielded 217 probe sets ("RPP7 or 8") which correspond to sequences, the expression of which were increased by at least 2.5 fold and were RPP7- or RPP8-dependent (Table 6).

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# Table 6

ProbeSet	Description
12007_at (Z99708.249_AT)	emb CAB16829.1  (Z99708) putative protein [Arabidopsis thaliana]
12091_at (AC004450.116_AT)	gb AAC64313.1  (AC004450) unknown protein [Arabidopsis thaliana]
 12115_at (AL033545.26_AT)	emb CAA22152.1  (AL033545) extensin-like protein [Arabidopsis thaliana]
12240_at (AL078579.130_AT)	emb CAB43974.1  (AL078579) putative protein [Arabidopsis thaliana]
12278_at (AJ011674.2_AT)	emb CAA09731.1  (AJ011674) receptor-like protein kinase, RLK3 [Arabidopsis thaliana] gb AAD12037.1  (AC002392) putative
12307_at (AC002392.162_AT)	receptor-like protein kinase [Arabidopsis thaliana] dbj BAA82810.1  (AB023448) basic
12332_s_at (AB023448.2_S_AT)	endochitinase [Arabidopsis thaliana] emb CAA16619.1  (AL021637) vacuolar
12341_s_at (AL021637.176_S_AT)	sorting receptor-like protein [Arabidopsis thaliana]
12349_s_at (X84728.6_S_AT)	gb AAA17993.1  (M91192) phenylalanine ammonia-lyase [Trifolium subterraneum] emb CAA18468.1  (AL022347)
12360_at (AL022347.131_AT)	serine/threonine kinase-like protein [Arabidopsis thaliana] gb AAC32192.1  (AF081067) IAA-Ala
12500_s_at (AF081067.3_S_AT)	hydrolase; IAA-amino acid hydrolase [Arabidopsis thaliana]
12521_at (AF049236.28_AT)	gb AAC14413.1  (AF049236) unknown [Arabidopsis thaliana]
12538_at (AF033205.2_AT)	gb AAC02973.1  (AF033205) putative pectin methylesterase [Arabidopsis thaliana] emb CAB45330.1  (AL079344) nucleotide
12556_at (AL079344.155_AT)	pyrophosphatase-like protein [Arabidopsis thaliana]
125/4_at (X82624.2_AT)	emb CAA57944.1  (X82624) SRG2At [Arabidopsis thaliana]
12630_at (AF024504.13_AT)	gb AAF18681.1 AF024504_11 (AF024504) unknown protein [Arabidopsis thaliana]
12642_at (AC006920.138_AT)	gb AAD22285.1 AC006920_9 (AC006920) unknown protein [Arabidopsis thaliana]

gb AI099570 and gb T22281 come from this gene. [Arabidopsis thaliana] gb AD25772.1 AC006577 8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolas with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AD25772.1 AC006577 8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolas with GDSL-motif family. ESTs gb T44453, gb AD09570 and gb T22281 come from this gene. [Arabidopsis thaliana] gb AAD25772.1 AC006577 8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolas with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb A1099570 and gb T22281 come from this gene. [Arabidopsis thaliana] gb AAC49282.1] (U40856) AIGI [Arabidopsis thaliana] gb AAC49282.1] (U40856) AIGI [Arabidopsis thaliana] gb AAC49282.1] (U40856) AIGI [Arabidopsis thaliana] gb AAC63850.1] (U73786) ACC synthase [Arabidopsis thaliana] gb AAC63850.1] (U73786) ACC synthase [Arabidopsis thaliana] db BAA32419.1] (AB008104) ethylene responsive element binding factor 2 [Arabidopsis thaliana] db BAA32422.1] (AB008107) ethylene responsive element binding factor 5 [Arabidopsis thaliana] gb AAC13947.1] (AF021244) coronatine-induced protein 1 [Arabidopsis thaliana] emb CAA65420.1] (X96600) pathogenesis-related protein 1 [Arabidopsis thaliana] gb AAC26690.1] (AC004077) putative cytochrome P450 [Arabidopsis thaliana] gb AAC24083.1] (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U26344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519 come from this gene. [Arabidopsis thaliana] gb AAC26690.1] (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and gb AAC06158.1] (AC003680) putative		gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138,
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Belongs to the PF 00657 Lipase/Acylhydrolas with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb T04815, gb T45993, gb R30138, gb Al099570 and gb T22281 come from this gene. [Arabidopsis thaliana] gb AAC49282.1  (U40856) AIG1 [Arabidopsi thaliana] gb AAC49282.1  (U40856) AIG1 [Arabidopsi thaliana] gb AAC49282.1  (U40856) AIG1 [Arabidopsi thaliana] gb AAC63850.1  (U73786) ACC synthase [Arabidopsis thaliana] gb AAC63850.1  (U73786) ACC synthase [Arabidopsis thaliana] db]BAA32419.1  (AB008104) ethylene responsive element binding factor 2 [Arabidopsis thaliana] db]BAA32422.1  (AB008107) ethylene responsive element binding factor 5 [Arabidopsis thaliana] gb AAC13947.1  (AF021244) coronatine-induced protein 1 [Arabidopsis thaliana] emb CAA65420.1  (X96600) pathogenesis-related protein 1 [Arabidopsis thaliana] gb AAC24083.1  (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AAC042519 come from this gene. [Arabidopsis thaliana] gb AAC06158.1  (AC003680) putative	12778_r_at (AC006577.16_R_AT)	with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene. [Arabidopsis thaliana]
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[Arabidopsis thaliana]  gb AAC13947.1  (AF021244) coronatine- induced protein 1 [Arabidopsis thaliana]  emb CAA65420.1  (X96600) pathogenesis- related protein 1 [Arabidopsis thaliana]  gb AAC26690.1  (AC004077) putative  cytochrome P450 [Arabidopsis thaliana]  gb AAC24083.1  (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519  come from this gene. [Arabidopsis thaliana] gb AAC06158.1  (AC003680) putative	12905_s_at (ATERF2_S_AT)	[Arabidopsis thaliana] dbj BAA32422.1  (AB008107) ethylene
induced protein 1 [Arabidopsis thaliana] emb CAA65420.1  (X96600) pathogenesis- related protein 1 [Arabidopsis thaliana] gb AAC26690.1  (AC004077) putative cytochrome P450 [Arabidopsis thaliana] gb AAC24083.1  (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519 come from this gene. [Arabidopsis thaliana] gb AAC06158.1  (AC003680) putative	12908_s_at (ATERF5_S_AT)	responsive element binding factor 5 [Arabidopsis thaliana]
related protein 1 [Arabidopsis thaliana] gb AAC26690.1  (AC004077) putative cytochrome P450 [Arabidopsis thaliana] gb AAC24083.1  (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519 come from this gene. [Arabidopsis thaliana] gb AAC06158.1  (AC003680) putative	12916_s_at (ATHCOR1_S_AT)	induced protein 1 [Arabidopsis thaliana]
gb AAC24083.1  (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519 come from this gene. [Arabidopsis thaliana] gb AAC06158.1  (AC003680) putative	12933_r_at (ATPR1TAN_R_AT)	related protein 1 [Arabidopsis thaliana]
calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519 come from this gene. [Arabidopsis thaliana] gb AAC06158.1  (AC003680) putative	12989_s_at (AC004077.149_S_AT)	cytochrome P450 [Arabidopsis thaliana]
come from this gene. [Arabidopsis thaliana] gb AAC06158.1  (AC003680) putative		calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719,
gb AAC06158.1  (AC003680) putative cytochrome P450 [Arabidopsis thaliana]	13067_s_at (AC003114.16_S_AT)	come from this gene. [Arabidopsis thaliana]
gb AAB60774.1  (AC000375) ESTs	13100_at (AC003680.50_AT)	cytochrome P450 [Arabidopsis thaliana]
gb U75592,gb T13956,gb T43869 come from from this gene. [Arabidopsis thaliana]	13115_at (AC000375.44_AT)	gb U75592,gb T13956,gb T43869 come from

	13154_s_at (AC002333.210_S_AT)	gb AAB64047.1  (AC002333) putative endochitinase [Arabidopsis thaliana]
	13176_at (AL031394.56_AT)	emb CAA20567.1  (AL031394) putative protein [Arabidopsis thaliana]
	13187_i_at (ATTHIRED4_I_AT)	gb AAC49356.1  (U35829) thioredoxin h [Arabidopsis thaliana]
	13188_r_at (ATTHIRED4_R_AT)	gb AAC49356.1  (U35829) thioredoxin h [Arabidopsis thaliana]
	13189_s_at (ATTHIRED4_S_AT)	gb AAC49356.1  (U35829) thioredoxin h [Arabidopsis thaliana]
	13198_i_at (ATTS0190_I_AT)	emb CAA16892.1  (AL021749) 12S cruciferin seed storage protein [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563)
	13215_s_at (CAFFEROYLCOAMETHYLTRA NS_S_AT)	putative S-adenosyl-L-methionine:trans- caffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
	13217_s_at (CALMODULINLIKE_S_AT)	emb CAB42906.1  (AL049862) calmodulin- like protein [Arabidopsis thaliana]
	13273_s_at (HSF4_S_AT)	gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
	13277_i_at (HSP176A_I_AT)	emb CAA74399.1  (Y14070) Heat Shock Protein 17.6A [Arabidopsis thaliana]
	13285_s_at (HSP83_S_AT)	gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAD22369.1 AC006580 1 (AC006580)
٠.	13381_at (AC006580.8_AT)	NAM (no apical meristem)-like protein [Arabidopsis thaliana] gb AAC49775.1  (AF003102) AP2 domain
	13435_at (AF003102.3_AT)	containing protein RAP2.9 [Arabidopsis thaliana]
	13588_at (AL021961.24_AT)	emb CAA17552.1  (AL021961) Phosphoglycerate dehydrogenase - like protein [Arabidopsis thaliana] emb CAA23036.1  (AL035394) putative
	13627_at (AL035394.196_AT)	Na+/H+-exchanging protein [Arabidopsis thaliana] gb AAB82634.1  (AC002387) putative
	13631_at (AC002387.185_AT)	transketolase precursor [Arabidopsis thaliana] emb CAA18462.1  (AL022347)
	13659_at (AL022347.46_AT)	serine/threonine kinase-like protein [Arabidopsis thaliana] gb AAD25552.1 AC005850_9 (AC005850)
		Highly Similiar to Mlo proteins [Arabidopsis thaliana]
	13696_at (NII15_AT)	No hits found less than or equal to 1e-15.
	13697_at (NI16_AT)	No hits found.

13751_at (NOVARTIS127_AT)	gb AAF16751.1 AC010155_4 (AC010155) F3M18.8 [Arabidopsis thaliana]
13764_at (NOVARTIS22_AT)	gb AAD39641.1 AC007591_6 (AC007591) F9L1.6 [Arabidopsis thaliana] gb AAD15433.1  (AC006218) putative
13818_s_at (AC006218.175_S_AT)	aspartate aminotransferase [Arabidopsis thaliana] emb CAB39611.1  (AL049480) possible
13880_s_at (AL049480.183_S_AT)	apospory-associated like protein [Arabidopsis thaliana]
13966_at (AL022023.172_AT)	emb CAA17775.1  (AL022023) putative protein [Arabidopsis thaliana] gb AAC78535.1  (AC005662) putative
14083_at (AC005662.56_AT)	embryo-abundant protein [Arabidopsis thaliana]
14096_at (AC002291.12_AT)	No hits found.
14110_i_at (AL035528.279_I_AT)	emb CAB36854.1  (AL035528) putative disease resistance protein [Arabidopsis thaliana] emb CAB36854.1  (AL035528) putative
14111_s_at (AL035528.279_S_AT)	disease resistance protein [Arabidopsis thaliana] gb AAC26243.1  (AF077407) contains
*	similarity to sugar transporters (Pfam: sugar_tr.hmm, score: 395.39) [Arabidopsis
14116_at (AF077407.30_AT)	thaliana]
14141_at (NOVARTIS31_AT)	No hits found less than or equal to 1e-15. dbj BAA22813.1  (D26015) CND41, chloroplast nucleoid DNA binding protein
14145_at (NOVARTIS35_AT)	[Nicotiana tabacum]
14146_at (NOVARTIS36_AT)	gb AAD25550.1 AC005850_7 (AC005850) Hypothetical protein [Arabidopsis thaliana]
14148_at (NOVARTIS38_AT)	gb AAF34713.1 AF224762_1 (AF224762) SigA binding protein [Arabidopsis thaliana]
14201_at (NOVARTIS73_AT)	No hits found less than or equal to 1e-15.
14202_at (NOVARTIS73_RC_AT)	No hits found less than or equal to 1e-15.
14223_at (NOVARTIS9_AT)	emb CAA19683.1  (AL024486) putative protein [Arabidopsis thaliana]
14232_at (NOVARTIS95_AT)	gb AAF16756.1 AC010155_9 (AC010155) F3M18.20 [Arabidopsis thaliana]

14248_at (PAD3_AT)	gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to gb X97864 cytochrome P450 from Arabidopsis thaliana and is a member of the PF 00067 Cytochrome P450 family. ESTs gb N65665, gb T14112, gb T76255, gb T20906 and gb AI100027 come from this gene.
14249_i_at (PAD4_I_AT)	emb CAB43438.1  (AL050300) putative protein [Arabidopsis thaliana]
14250_r_at (PAD4_R_AT)	emb CAB43438.1  (AL050300) putative protein [Arabidopsis thaliana]
14254_s_at (PAL1-MRNA_S_AT)	gb AAD18156.2  (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis
14256_f_at (PAL1-INTRON_F_AT)	gb AAD18156.2  (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis thaliana]
14278_at (AL022373.295_AT)  14320_at (AC005956.54_AT)	emb CAA18503.1  (AL022373) hypothetical protein [Arabidopsis thaliana] gb AAD23719.1 AC005956_8 (AC005956) putative RING zinc finger protein [Arabidopsis thaliana]
14381_at (AC003930.34_AT)  14381_at (AC002521.68_AT)  14443_at (AC000348.23 AT)	thaliana] gb AAC05341.1  (AC002521) unknown protein [Arabidopsis thaliana] gb AAB61498.1  (AC000348) T7N9.22 [Arabidopsis thaliana]
14609_at (AC002340.147_AT)	gb AAC02748.1  (AC002340) putative cytochrome P450 [Arabidopsis thaliana]
14614_at (AC004165.66_AT)	gb AAC16958.1  (AC004165) putative glucosyltransferase [Arabidopsis thaliana] gb AAA32835.1  (M96073) phosphoribosylanthranilate transferase
14620_s_at (PAT1_S_AT)	[Arabidopsis thaliana]
14621_at (PDF1.2_AT)	gb AAC31244.1  (AC004747) putative antifungal protein [Arabidopsis thaliana] gb AAC69381.1  (AC005398) pathogenesis- related PR-1-like protein [Arabidopsis
14635_s_at (PR.1_S_AT)	thaliana] emb CAA50677.1  (X71794) peroxidase
14638_s_at (PRXCB_S_AT)	[Arabidopsis thaliana]
14672_s_at (TSA1_S_AT)	gb AAC49117.1  (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana] gb AAA32879.1  (M81620) tryptophan synthase beta-subunit [Arabidopsis thaliana]
14673_s_at (TSB2_S_AT)	thaliana]
14682_i_at (WT1012A_RC_I_AT)	No hits found.
14691_at (WT1096_AT)	No hits found.

14704_s_at (WT768_RC_S_AT)	gb AAD15461.1  (AC006067) unknown protein [Arabidopsis thaliana]
14709_at (WT788_AT)	No hits found less than or equal to 1e-15. emb CAA60521.1  (X86958) protein kinase
14763_at (X86958.1_AT)	catalytic domain (fragment) [Arabidopsis thaliana] gb AAA32835.1  (M96073)
14838_s_at (M96073.6_S_AT)	phosphoribosylanthranilate transferase [Arabidopsis thaliana]
14882_at (AL022605.63_AT)	emb CAA18753.1  (AL022605) putative protein [Arabidopsis thaliana] emb CAA17549.1  (AL021961) cinnamyl alcohol dehydrogenase - like protein
15042_at (AL021961.3_AT)	[Arabidopsis thaliana]
15116_f_at (AF121356_F_AT)	gb AAD28243.1 AF121356_1 (AF121356) peroxiredoxin TPx2 [Arabidopsis thaliana] gb AAB53975.1  (U90522) lysine-ketoglutarate
15161_s_at (ATU90522_S_AT)	reductase/saccharopine dehydrogenase [Arabidopsis thaliana] gb AAB63077.1  (U93215) unknown protein
15366_at (U93215.38_AT)	[Arabidopsis thaliana]
15415_at (AF000657.28_AT)	gb AAB72169.1  (AF000657) hypothetical protein [Arabidopsis thaliana]
15431_at (AL030978.64_AT)	emb CAA19722.1  (AL030978) putative protein [Arabidopsis thaliana]
15523_s_at (AL078637.213_S_AT)	emb CAB45071.1  (AL078637) putative protein [Arabidopsis thaliana]
15532_r_at (AL078637.191_R_AT)	emb CAB45069.1  (AL078637) putative protein [Arabidopsis thaliana]
15616_s_at (ATHPRO25A_S_AT)	dbj BAA24440.1  (AB010407)
15629_s_at (AB003280_S_AT)	phosphoglycerate dehydrogenase [Arabidopsis thaliana] gb AAD10829.1  (AF117063) putative inositol
15641_s_at (AF117063_S_AT)	polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] gb AAB80922.1  (AF022658) putative c2h2
15665_s_at (AF022658_S_AT)	zinc finger transcription factor [Arabidopsis thaliana] emb CAA67234.1  (X98676) zinc finger
15778_at (X98676.2_AT)	protein [Arabidopsis thaliana]
15779_g_at (X98676.2_G_AT)	emb CAA67234.1  (X98676) zinc finger protein [Arabidopsis thaliana] gb AAD15461.1  (AC006067) unknown
15846_at (AC006067.63_AT)	protein [Arabidopsis thaliana]

15847_g_at (AC006067.63_G_AT)	gb AAD15461.1  (AC006067) unknown protein [Arabidopsis thaliana]
15859_at (AC006587.164_AT)	gb AAD21491.1  (AC006587) unknown protein [Arabidopsis thaliana]
15970_s_at (X71794.2_S_AT)	emb CAA50677.1  (X71794) peroxidase [Arabidopsis thaliana]
15978_at (X68592.6_AT) 16053_i_at (Y14251.4_I_AT)	emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] emb CAA74639.1  (Y14251) glutathione S- transferase [Arabidopsis thaliana]
16061_s_at (AB004796_S_AT)  16083_s_at (AF153283 S AT)	gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAD34615.1 AF153283_1 (AF153283) putative progesterone-binding protein homolog [Arabidopsis thaliana]
16091_s_at (ATHHSP83_S_AT)	gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana]
16105_s_at (ATU68017_S_AT)	gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
16150_s_at (ATHPEAR_S_AT)	emb CAB41718.1  (AL049730) pEARLI 1 [Arabidopsis thaliana]
16232_s_at (AL080252.77_S_AT)	emb CAB45796.1  (AL080252) putative protein [Arabidopsis thaliana] gb AAC32915.1  (AC004138) putative
16257_at (AC004138.105_AT)	nucleoside triphosphatase [Arabidopsis thaliana]
16298_at (AL021890.71_AT)	emb CAA17152.1  (AL021890) putative protein [Arabidopsis thaliana]
16299_at (AL024486.185_AT)	emb CAA19705.1  (AL024486) putative protein [Arabidopsis thaliana] gb AAD40144.1 AF149413_25 (AF149413) contains similarity to protein kinase domains (Pfam F00069, Score=162.6, E=6.8e-45, N=1) and leucien rich repeats (Pfam PF00560, Score=210.7, F=222.50) N=10 ft. 10 ft.
16357_at (AF149413.38_AT)	Score=210.7, E=2.2e-59, N=10) [Arabidopsis thaliana] emb CAA20203.1  (AL031187) receptor-like
16360_at (AL031187.126_AT)	serine/threonine protein kinase ARK3 [Arabidopsis thaliana]
16365_at (AC003974.136_AT)	gb AAC04495.1  (AC003974) putative disease resistance protein [Arabidopsis thaliana]
16522_at (X77500.2_AT)	emb CAA54631.1  (X77500) amino acid transporter [Arabidopsis thaliana] dbj BAA32422.1  (AB008107) ethylene
16536_s_at (AB008107_S_AT)	responsive element binding factor 5 [Arabidopsis thaliana]

16578_s_at (ATHRPRP1B_S_AT)  16609_s_at (AB008104_S_AT)  16649_s_at (ATHORF_S_AT)	emb CAB68132.1  (AL137080) beta-1, 3-glucanase 2 (BG2) [Arabidopsis thaliana] dbj BAA32419.1  (AB008104) ethylene responsive element binding factor 2 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
16817_s_at (AL096882.91_S_AT)	emb CAB51412.1  (AL096882) ACC synthase (AtACS-6) [Arabidopsis thaliana]
16914_s_at (AL049500.57_S_AT)	emb CAB39936.1  (AL049500) osmotin precursor [Arabidopsis thaliana]
16981_s_at (U35829.2_S_AT)	gb AAC49356.1  (U35829) thioredoxin h [Arabidopsis thaliana]
16995_at (AC002391.188_AT)	gb AAB87114.1  (AC002391) unknown protein [Arabidopsis thaliana] gb AAD23027.1 AC006585_22 (AC006585) putative tyrosine aminotransferase
17008_at (AC006585.212_AT)	[Arabidopsis thaliana]
17014_s_at (ATU05206_S_AT)	gb AAC48925.1  (U05206) ribonuclease [Arabidopsis thaliana]
17051_s_at (AF098947_S_AT)	gb AAD09952.1  (AF098947) CTF2B [Arabidopsis thaliana]
17068_s_at (ATHRLPKA_S_AT)	gb AAA32857.1  (M84658) receptor-like protein kinase [Arabidopsis thaliana] gb AAD25838.1 AC006951_17 (AC006951) putative indole-3-glycerol phosphate synthase
17083_s_at (ATU18770_S_AT)	[Arabidopsis thaliana] gb AAC49697.1  (U66345) calreticulin
17097_s_at (ATU66345_S_AT)	[Arabidopsis thaliana]
17128_s_at (ATHRPRP1A_S_AT)	gb AAC69381.1  (AC005398) pathogenesis- related PR-1-like protein [Arabidopsis
17278_at (AC000107.5_AT)	gb AAD36959.1 AC000107_5 (AC000107) F17F8.5 [Arabidopsis thaliana]
17413_s_at (AJ006961.4_S_AT)	emb CAA67551.1  (X99097) peroxidase [Arabidopsis thaliana] gb AAB60752.1  (AC000132) Similar to A. thaliana receptor-like protein kinase (gb RLK5_ARATH). ESTs gb ATTS0475,gb ATTS4362 come from this
17464_at (AC000132.72_AT)	gene. [Arabidopsis thaliana] emb CAB10405.1  (Z97340) beta-1, 3- glucanase class I precursor [Arabidopsis
17485_s_at (Z97340.345_S_AT)	thaliana]

17487_s_at (U18993.2_S_AT)	gb AAC49117.1  (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
17499_s_at (AF107726_S_AT)	gb AAD19610.1  (AF107726) cyclic nucleotide gated channel [Arabidopsis thaliana]
17500_s_at (ATHCALLGA_S_AT)	emb CAB42906.1  (AL049862) calmodulin- like protein [Arabidopsis thaliana] gb AAB71482.1  (AC002294) similar to S-
17511_s_at (AF067605_S_AT)	linalool synthase gp U58314 1491939 [Arabidopsis thaliana] dbj BAA28538.1  (D78606) cytochrome P450
17522_s_at (D78606_S_AT)	monooxygenase [Arabidopsis thaliana] gb AAC49282.1  (U40856) AIG1 [Arabidopsis
17544_s_at (ATU40856_S_AT)	thaliana]
17653_at (AL035679.144_AT)	emb CAB38823.1  (AL035679) putative protein [Arabidopsis thaliana]
17744_s_at (AC004684.168_S_AT)	gb AAC23646.1  (AC004684) putative alcohol dehydrogenase [Arabidopsis thaliana] gb AAC28500.1  (AC004392) Similar to glucose-6-phosphate/phosphate-translocator
17775_at (AC004392.2_AT)	(GPT) gb AF020814 from Pisum sativum. [Arabidopsis thaliana] gb AAB64049.1  (AC002333) putative
17840_s_at (AC002333.223_S_AT)	endochitinase [Arabidopsis thaliana]
17843_s_at (AC002391.150_S_AT)	gb AAB87109.1  (AC002391) putative cytochrome P450 [Arabidopsis thaliana]
17899_at (Z97339.197_AT)	emb CAB10339.1  (Z97339) hypothetical protein [Arabidopsis thaliana] emb CAA07352.1  (AJ006960) peroxidase
17930_s_at (AJ006960.4_S_AT)	[Arabidopsis thaliana] emb CAB59428.1  (AJ002295) inositol-1,4,5-
18012_s_at (AJ002295_S_AT)	trisphosphate 5-Phosphatase [Arabidopsis thaliana] emb CAB52675.1  (AJ010971) glucose-6-phosphate 1-dehydrogenase [Arabidopsis
18022_at (AJ010971_AT)	thaliana]
18054_at (AJ238846_AT)	emb CAB54517.1  (AJ238846) SGP1 monomeric G-protein [Arabidopsis thaliana] gb AAF24959.1 AC012375 22 (AC012375)
18216_at (X95573.2_AT)	T22C5.18 [Arabidopsis thaliana]
18217_g_at (X95573.2_G_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana] emb CAA17150.1  (AL021890) putative
18224_s_at (AL021890.57_S_AT)	protein [Arabidopsis thaliana] gb AAB87112.1  (AC002391) putative
18551_at (AC002391.163_AT)	cytochrome P450 [Arabidopsis thaliana]
18567_at (AC004411.25_AT)	gb AAC34217.1  (AC004411) putative alcohol dehydrogenase [Arabidopsis thaliana]

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18590_at (AJ222713.4_AT)	emb CAA10955.1  (AJ222713) unnamed protein product [Arabidopsis thaliana]
18591_at (X74756.2_AT)	emb CAA52772.1  (X74756) ATAF2 [Arabidopsis thaliana] gb AAC72125.1  (AC005278) ESTs gb H36966, gb R65511, gb T42324 and
18625_at (AC005278.22_AT)	gb[T20569 come from this gene. [Arabidopsis thaliana]
18716_at (X91916_AT)	gb AAF26754.1 AC007396_3 (AC007396) T4O12.6 [Arabidopsis thaliana] gb AAC33239.1  (AC005315) putative ligand-
18844_at (AC005315.131_AT)	gated ion channel protein [Arabidopsis thaliana]
18908_i_at (AF055848.2_I_AT)	gb AAC62611.1  (AF055848) subtilisin-like protease [Arabidopsis thaliana]
18909_s_at (AF055848.2_S_AT)	gb AAC62611.1  (AF055848) subtilisin-like protease [Arabidopsis thaliana] gb AAC16927.1  (AC002338) putative laccase
18920_at (AC002338.11_AT)	[Arabidopsis thaliana]
18946_at (Y11788.1_AT)	emb CAA72484.1  (Y11788) peroxidase ATP24a [Arabidopsis thaliana] gb AAD45127.1 AF163823_1 (AF163823)
18968_at (AF163823.4_AT)	endoxyloglucan transferase [Arabidopsis thaliana] gb AAD45127.1 AF163823_1 (AF163823)
18969_g_at (AF163823.4_G_AT)	endoxyloglucan transferase [Arabidopsis thaliana]
18983_s_at (AL049730.104_S_AT)	emb CAB41722.1  (AL049730) pEARLI 1-like protein [Arabidopsis thaliana] emb CAA65053.1  (X95738) proline
19158_at (X95738.2_AT)	transporter 2 [Arabidopsis thaliana] gb AAB64325.1  (AC002335) putative trypsin
19171_at (AC002335.160_AT)	inhibitor [Arabidopsis thaliana]
19177_at (X99923.1_AT)	emb CAA68191.1  (X99923) male sterility 2-like protein [Arabidopsis thaliana] emb CAA21214.1  (AL031804) putative
19182_at (AL031804.245_AT)	protein [Arabidopsis thaliana]
19229_at (AC003027.39_AT)	gb AAD10694.1  (AC003027) lcl prt_seq No definition line found [Arabidopsis thaliana]
19247_at (AF071527.44_AT)	gb AAD11587.1 AAD11587 (AF071527) hypothetical protein [Arabidopsis thaliana] gb AAC27173.1  (AC003028) putative
19284_at (AC003028.196_AT)	anthocyanidin synthase [Arabidopsis thaliana]
19415_at (AL080253.32_AT)	emb CAB45805.1  (AL080253) putative protein [Arabidopsis thaliana]
19594_i_at (X98321.2_I_AT)	emb CAA66965.1  (X98321) peroxidase [Arabidopsis thaliana]

19640_at (AC004561.78_AT)	gb AAC95192.1  (AC004561) putative glutathione S-transferase [Arabidopsis thaliana] emb CAB43638.1  (AL050351) NAD(P)H
19664_at (AL050351.172_AT)	oxidoreductase, isoflavone reductase-like protein [Arabidopsis thaliana] gb[AAC36019.1] (AC005687) RAP2.6
19672_at (AC005687.19_AT)	[Arabidopsis thaliana] emb[CAB36812.1] (AL035527) peptide
19762_at (AL035527.204_AT)	transporter-like protein [Arabidopsis thaliana]
19892_at (AC005770.30_AT)	gb AAC79626.1  (AC005770) putative protease inhibitor [Arabidopsis thaliana] emb CAA05025.1  (AJ001809) succinate dehydrogenase flavoprotein alpha subunit
19894_at (AJ001809.1_AT)	[Arabidopsis thaliana] gb AAC79593.1  (AC005727) unknown
19914_at (AC005727.175_AT)	protein [Arabidopsis thaliana]
19951_at (AC005395.47_AT)	gb AAC42241.1  (AC005395) unknown protein [Arabidopsis thaliana] gb AAC17040.1  (AC002986) Similarity to A. thaliana gene product F21M12.20,
19982_at (AC002986.28_AT)	gb AC000132. EST gb Z25651 comes from this gene. [Arabidopsis thaliana] gb AAD21459.1  (AC007017) similar to
19991_at (AC007017.124_AT)	harpin-induced protein hin1 from tobacco [Arabidopsis thaliana] gb AAD32864.1 AC005489_2 (AC005489)
20189_at (AC005489.2_AT)	F14N23.2 [Arabidopsis thaliana] emb CAA18469.1  (AL022347) serine/threonine kinase-like protein
20223_at (AL022347.145_AT)	[Arabidopsis thaliana] emb CAA18460.1  (AL022347) protein kinase-
20232_s_at (AL022347.12_S_AT)	like protein [Arabidopsis thaliana]
20238_at (X74514.2_AT)	emb CAA52619.1  (X74514) beta- fructofuranosidase [Arabidopsis thaliana] emb CAA52619.1  (X74514) beta-
20239_g_at (X74514.2_G_AT)	fructofuranosidase [Arabidopsis thaliana] emb CAA05625.1  (AJ002584) AtMRP4
20245_s_at (AC005309.97_S_AT)	[Arabidopsis thaliana] thaliana] gb AAC95354.1  (AF084037) receptor-like
20246_s_at (AF084037.3_S_AT)	protein kinase [Arabidopsis thaliana] gb AAB82640.1  (AC002387) putative
20269_at (AC002387.237_AT)	pectinesterase [Arabidopsis thaliana] emb CAA74930.1  (Y14590) class IV chitinase
20287_at (Y14590.5_AT)	[Arabidopsis thaliana] emb CAA74930.1  (Y14590) class IV chitinase
20288_g_at (Y14590.5_G_AT)	[Arabidopsis thaliana]
20291_s_at (M92353.4_S_AT)	gb AAA32738.1  (M92353) anthranilate

synthase alpha subunit [Arabidopsis thaliana] gb|AAD23617.1|AC007168 8 (AC007168) putative aspartate aminotransferase 20331 at (AC007168.86 AT) [Arabidopsis thaliana] gb|AAD25552.1|AC005850 9 (AC005850) Highly Similar to Mlo proteins [Arabidopsis 20365\_s\_at (AC005850.19 S AT) thaliana] gb|AAC36163.1| (AC005314) putative serpin 20368 at (AC005314.38 AT) [Arabidopsis thaliana] emb|CAA19698.1| (AL024486) putative 20420 at (AL024486.131 AT) chitinase [Arabidopsis thaliana] emb|CAB10219.1| (Z97336) hypothetical 20429 s at (Z97336.167 S AT) protei [Arabidopsis thaliana] gb|AAC98070.1| (AC005896) putative C2H2-20620 g at (AC005896.161 G AT) type zinc finger protein [Arabidopsis thaliana] emb|CAB41131.1| (AL049658) hypothetical 20625 at (AL049658.102 AT) protein [Arabidopsis thaliana]

The functions of RPP7 and RPP8 are independent of PAD4, NDR1. NahG, COII, and EDSI. Consequently, it is unlikely that genes whose expression is reduced in pad4 or NahG backgrounds are important for RPP7- or RPP8- mediated resistance. Although data from Peronospora infection of pad4 or nahG plants was not available, data from Pseudomonas syringae (ES4326) infection of pad4 and NahG plants was available and employed to determine which genes depended on PAD4 or were interfered with by NahG. Resistance to P. syringae mediated by RPS2 requires NDR1 function. Genes whose expression 10 requires RPS2 are less likely to be important for RPP7 or RPP8 mediated resistance than genes whose expression is independent of RPS2. Gene expression data was available for wild-type plants infected with Pseudomonas syringae pv. tomato DC3000 avrRpt2, and rps2 mutant plants infected with Pseudomonas syringae pv. tomato DC3000 avrRpt2, 6 hours after infection. 15 EDM1 is required for resistance mediated by RPP7. Thus, genes whose expression requires EDM1 are likely to be important for RPP7-mediated resistance. Gene expression data was available for an edml mutant (see above).

The data from the 7 additional data sets (Col Psm ES4326 30 hour; Col Mock 30 hour; Col NahG Psm ES4226 30 hour; Col pad4 Psm ES4326 30 hour; Col water control; Col DC3000 avrRpt2 and rps2 DC3000 avrPrt2 was added to the 217 probe sets in "RPP7 or 8". The following sets were identified: edm1:

20

Col edm1 Hiks1 12 hour/Col Hiks1 12 hour < 0.5 OR Col edm1 Hiks1 48 hour/Col Hiks1 48 hour < 0.5.

#### rps2:

5

Col DC3000 avrRpt2 6 hour/Col water control > 2 AND rps2 DC3000 avrRpt2 6 hour/Col DC3000 avrRpt2 6 hour < 0.5.

#### pad4 or NahG:

Col Psm ES4326 30 hour/Col Mock 30 hour > 2 AND Col pad4 Psm

10 ES4326 30 hour/Col Psm ES4326 30 hour < 0.5.

OR

Col Psm ES4326 30 hour/Col Mock 30 hour > 2 AND Col NahG Psm ES4326 30 hour/Col Psm ES4326 30 hour < 0.5.

There are 8 possible combinations of these three sets. Access was used to divide the 217 probe sets into 8 sets, numbered 2-9 (considering the whole set of 217 probe sets as set 1) as shown in Tables 7 and 8:

#### Table 7

20	<u>Set</u>	Content	Number of probe
	sets		
	2	RPP7 or 8 not edm1 not rps2 not pad4 o rNahG	62 probe sets
	3	edm1 not rps2 not pad4 or NahG	50 probe sets
	4	rps2 not edm1 not pad4 or NahG	14 probe sets
25	5	pad4 or NahG not edm1 not rps2	21 probe sets
	6	edm1 and rps2 not pad4 or NahG	23 probe sets
	7	edm1 and pad4 or NahG not rps2	25 probe sets
	8	rps2 and pad4 or NahG not edm1	8 probe sets
	9	edm1 and rps2 and pad4 or NahG	14 probe sets
30	Tota	al :	217 probe sets

#### Table 8

## 2 RPP7 or 8 not edm1 not rps2 not pad4orNahG

ProbeSet	Description
12007_at (Z99708.249_AT)	emb CAB16829.1  (Z99708) putative protein [Arabidopsis thaliana]
12240_at (AL078579.130_AT)	emb CAB43974.1  (AL078579) putative protein [Arabidopsis thaliana]
12349_s_at (X84728.6_S_AT)	gb AAA17993.1  (M91192) phenylalanine ammonia-lyase [Trifolium subterraneum]
12538_at (AF033205.2_AT)	gb AAC02973.1  (AF033205) putative pectin methylesterase [Arabidopsis thaliana] emb CAB45330.1  (AL079344) nucleotide
12556_at (AL079344.155_AT)	pyrophosphatase-like protein [Arabidopsis thaliana]
12642_at (AC006920.138_AT)	gb AAD22285.1 AC006920_9 (AC006920) unknown protein [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657
12777_i_at (AC006577.16_I_AT)	Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene. [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif
12779_f_at (AC006577.16_F_AT)	family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene. [Arabidopsis thaliana]
12889_s_at (ASA1_S_AT)	gb AAA32738.1  (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana] dbj BAA32422.1  (AB008107) ethylene responsive element binding factor 5
12908_s_at (ATERF5_S_AT)	[Arabidopsis thaliana]
12933_r_at (ATPR1TAN_R_AT)	emb CAA65420.1  (X96600) pathogenesis- related protein 1 [Arabidopsis thaliana] emb CAA20567.1  (AL031394) putative
13176_at (AL031394.56_AT)	protein [Arabidopsis thaliana]
13696_at (NI115_AT)	No hits found less than or equal to 1e-15. emb CAB39611.1  (AL049480) possible
13880_s_at (AL049480.183_S_AT)	apospory-associated like protein [Arabidopsis thaliana]
13966_at (AL022023:172_AT)	emb CAA17775.1  (AL022023) putative protein [Arabidopsis thaliana] emb CAB36854.1  (AL035528) putative
14111_s_at (AL035528.279_S_AT)	disease resistance protein [Arabidopsis thaliana]

14145_at (NOVARTIS35 AT)	dbj BAA22813.1  (D26015) CND41, chloroplast nucleoid DNA binding protein [Nicotiana tabacum]
11115_at (110 VAR(11555_A1)	[Nicotiana taoacum]
14256_f_at (PAL1- INTRON_F_AT)	gb AAD18156.2  (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis thaliana]
14278_at (AL022373.295_AT)	emb CAA18503.1  (AL022373) hypothetical protein [Arabidopsis thaliana] gb AAD23719.1 AC005956_8 (AC005956)
14320_at (AC005956.54_AT)	putative RING zinc finger protein [Arabidopsis thaliana]
14381_at (AC002521.68_AT)	gb AAC05341.1  (AC002521) unknown protein [Arabidopsis thaliana]
14443_at (AC000348.23_AT)	gb AAB61498.1  (AC000348) T7N9.22 [Arabidopsis thaliana] gb AAA32835.1  (M96073)
14620_s_at (PAT1_S_AT)	phosphoribosylanthranilate transferase [Arabidopsis thaliana]
14621_at (PDF1.2_AT)	gb AAC31244.1  (AC004747) putative antifungal protein [Arabidopsis thaliana] emb CAA17549.1  (AL021961) cinnamyl
15042_at (AL021961.3_AT)	alcohol dehydrogenase - like protein [Arabidopsis thaliana] gb AAB63077.1  (U93215) unknown protein
15366_at (U93215.38_AT)	[Arabidopsis thaliana]
15415_at (AF000657.28_AT)	gb AAB72169.1  (AF000657) hypothetical protein [Arabidopsis thaliana]
15523_s_at (AL078637.213_S_AT)	emb CAB45071.1  (AL078637) putative protein [Arabidopsis thaliana] gb AAD34615.1 AF153283_1 (AF153283)
16083_s_at (AF153283_S_AT)	putative progesterone-binding protein homolog [Arabidopsis thaliana] gb AAC32915.1  (AC004138) putative
16257_at (AC004138.105_AT)	nucleoside triphosphatase [Arabidopsis thaliana]
16298_at (AL021890.71_AT)	emb CAA17152.1  (AL021890) putative protein [Arabidopsis thaliana] emb CAA19705.1  (AL024486) putative
16299_at (AL024486.185_AT)	protein [Arabidopsis thaliana] emb CAA54631.1  (X77500) amino acid
16522_at (X77500.2_AT)	transporter [Arabidopsis thaliana] dbj BAA32422.1  (AB008107) ethylene
16536_s_at (AB008107_S_AT)	responsive element binding factor 5 [Arabidopsis thaliana] gb AAD09952.1  (AF098947) CTF2B
17051_s_at (AF098947_S_AT)	[Arabidopsis thaliana]

17083_s_at (ATU18770_S_AT) 17464_at (AC000132.72_AT)	gb AAD25838.1 AC006951_17 (AC006951) putative indole-3-glycerol phosphate synthase [Arabidopsis thaliana] gb AAB60752.1  (AC000132) Similar to A. thaliana receptor-like protein kinase (gb RLK5_ARATH). ESTs gb ATTS0475,gb ATTS4362 come from this gene. [Arabidopsis thaliana]
17487_s_at (U18993.2_S_AT)	gb AAC49117.1  (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
18216_at (X95573.2_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana]
18217_g_at (X95573.2_G_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana] emb CAA52772.1  (X74756) ATAF2
18591_at (X74756.2_AT)	[Arabidopsis thaliana] gb AAC72125.1  (AC005278) ESTs gb H36966, gb R65511, gb T42324 and
18625_at (AC005278.22_AT)	gb T20569 come from this gene. [Arabidopsis thaliana] gb AAC33239.1  (AC005315) putative ligand-
18844_at (AC005315.131_AT)	gated ion channel protein [Arabidopsis thaliana]
18908_i_at (AF055848.2_I_AT)	gb AAC62611.1  (AF055848) subtilisin-like protease [Arabidopsis thaliana] gb AAC62611.1  (AF055848) subtilisin-like
18909_s_at (AF055848.2_S_AT)	protease [Arabidopsis thaliana] gb AAC16927.1  (AC002338) putative laccase
18920_at (AC002338.11_AT)	[Arabidopsis thaliana]
18983_s_at (AL049730.104_S_AT)	emb CAB41722.1  (AL049730) pEARLI 1- like protein [Arabidopsis thaliana]
19158_at (X95738.2_AT)	emb CAA65053.1  (X95738) proline transporter 2 [Arabidopsis thaliana]
19229_at (AC003027.39_AT)	gb AAD10694.1  (AC003027) lcl prt_seq No definition line found [Arabidopsis thaliana] emb CAB45805.1  (AL080253) putative
19415_at (AL080253.32_AT)	protein [Arabidopsis thaliana]
19594_i_at (X98321.2_I_AT)	emb CAA66965.1  (X98321) peroxidase [Arabidopsis thaliana] emb CAB43638.1  (AL050351) NAD(P)H
.19664_at (AL050351.172_AT)	oxidoreductase, isoflavone reductase-like protein [Arabidopsis thaliana] emb CAA05025.1  (AJ001809) succinate
19894_at (AJ001809.1_AT)	dehydrogenase flavoprotein alpha subunit [Arabidopsis thaliana]

gb|AAC17040.1| (AC002986) Similarity to A. thaliana gene product F21M12.20, gb|AC000132. EST gb|Z25651 comes from 19982 at (AC002986.28 AT) this gene. [Arabidopsis thaliana] gb|AAD32864.1|AC005489 2 (AC005489) 20189\_at (AC005489.2\_AT) F14N23.2 [Arabidopsis thaliana] emb|CAA52619.1| (X74514) beta-20238\_at (X74514.2 AT) fructofuranosidase [Arabidopsis thaliana] emb|CAA52619.1| (X74514) beta-20239\_g\_at (X74514.2 G AT) fructofuranosidase [Arabidopsis thaliana] gb|AAC95354.1| (AF084037) receptor-like 20246\_s\_at (AF084037.3\_S\_AT) protein kinase [Arabidopsis thaliana] gb|AAA32738.1| (M92353) anthranilate 20291\_s\_at (M92353.4\_S\_AT) synthase alpha subunit [Arabidopsis thaliana] gb|AAD23617.1|AC007168\_8 (AC007168) putative aspartate aminotransferase 20331\_at (AC007168.86\_AT) [Arabidopsis thaliana] gb|AAC36163.1| (AC005314) putative serpin 20368\_at (AC005314.38 AT) [Arabidopsis thaliana] 20620 g at gb|AAC98070.1| (AC005896) putative C2H2-(AC005896.161 G AT) type zinc finger protein [Arabidopsis thaliana] **Duplicates** 12777 and 12779 (lipase) 12889 and 20291 (ASA1) 12908 and 16536 (ERF5) 17464 and 19982 (RLK5) 18216 and 18217 (T22C5.18) 18908 and 18909 (subtilisin-like protease) 20238 and 20239 (betafructofuranosidase) 3 edm1 without rps2 without

# 3 edm1 without rps2 without pad4orNahG ProbeSet

Description gb|AAC64313.1| (AC004450) unknown

12091\_at (AC004450.116\_AT)

protein [Arabidopsis thaliana]

en

emb|CAA22152.1| (AL033545) extensin-like

12115\_at (AL033545.26\_AT)

protein [Arabidopsis thaliana]

	gb AAC32192.1  (AF081067) IAA-Ala
	hydrolase; IAA-amino acid hydrolase
12500_s_at (AF081067.3_S_AT	)[Arabidopsis thaliana]
	gb AAC14413.1  (AF049236) unknown
12521_at (AF049236.28_AT)	[Arabidopsis thaliana]
	emb CAA57944.1  (X82624) SRG2At
12574_at (X82624.2_AT)	[Arabidopsis thaliana]
<b>–</b> ,	gb AAD25772.1 AC006577_8 (AC006577)
	Belongs to the PF 00657
	Lipase/Acylhydrolase with GDSL-motif
	family. ESTs gb T44453, gb T04815,
	gb T45993, gb R30138, gb A1099570 and
12778_r_at	gb T22281 come from this gene. [Arabidopsis
(AC006577.16_R_AT)	thaliana]
	gb AAC63850.1  (U73786) ACC synthase
12891_at (ATACS6 AT)	[Arabidopsis thaliana]
_ ` /	gb AAC63850.1  (U73786) ACC synthase
12892 g at (ATACS6 G AT)	[Arabidopsis thaliana]
_0_ (	dbj BAA32419.1  (AB008104) ethylene
	responsive element binding factor 2
12905_s_at (ATERF2_S_AT)	[Arabidopsis thaliana]
(	gb AAC13947.1  (AF021244) coronatine-
12916_s_at (ATHCOR1 S AT)	induced protein 1 [Arabidopsis thaliana]
13188 r at	gb AAC49356.1  (U35829) thioredoxin h
(ATTHIRED4_R_AT)	[Arabidopsis thaliana]
(030 21111)	emb CAA16892.1  (AL021749) 12S cruciferin
13198_i_at (ATTS0190 I AT)	seed storage protein [Arabidopsis thaliana]
	gb AAC49775.1  (AF003102) AP2 domain
	containing protein RAP2.9 [Arabidopsis
13435_at (AF003102.3 AT)	thaliana]
10 100_11 (11 000102.5_111)	gb AAB82634.1  (AC002387) putative
13631_at (AC002387.185 AT)	transketolase precursor [Arabidopsis thaliana]
10001_41 (110002507.105_111)	transactorase precursor [Arabidopsis thanana]
14096_at (AC002291.12_AT)	No hits found.
	emb CAA19683.1  (AL024486) putative
14223_at (NOVARTIS9 AT)	protein [Arabidopsis thaliana]
- in (and 11111)	gb AAF16756.1 AC010155_9 (AC010155)
14232_at (NOVARTIS95 AT)	F3M18.20 [Arabidopsis thaliana]
14254 s at (PAL1-	gb AAD18156.2  (AC006260) phenylalanine
MRNA S AT)	ammonia lyase (PAL1) [Arabidopsis
	gb[AAC16958.1] (AC004165) putative
14614_at (AC004165.66 AT)	glucosyltransferase [Arabidopsis thaliana]
14691 at (WT1096 AT)	No hits found.
11021_at (W 11030_A1)	
•	gb AA32835.1  (M96073)
14838_s_at (M96073.6_S_AT)	phosphoribosylanthranilate transferase
11030_3_at (111300/3.0_5_A1)	[Arabidopsis thaliana]
·	gb AAB53975.1  (U90522) lysine-
15161 c at (ATTIONS 22 C ATT	ketoglutarate reductase/saccharopine
15161_s_at (ATU90522_S_AT)	dehydrogenase [Arabidopsis thaliana]

15532_r_at	emb CAB45069.1  (AL078637) putative
(AL078637.191_R_AT)	protein [Arabidopsis thaliana]
	dbj BAA24440.1  (AB010407)
	phosphoglycerate dehydrogenase [Arabidopsis
15629_s_at (AB003280_S_AT)	thaliana]
	gb AAB80922.1  (AF022658) putative c2h2
·	zinc finger transcription factor [Arabidopsis
15665_s_at (AF022658 S AT)	thaliana]
16232 s at	•
	emb CAB45796.1  (AL080252) putative
(AL080252.77_S_AT)	protein [Arabidopsis thaliana]
	dbj BAA32419.1  (AB008104) ethylene
16600+ (AD000104 G AT)	responsive element binding factor 2
16609_s_at (AB008104_S_AT)	[Arabidopsis thaliana]
16817_s_at	emb CAB51412.1  (AL096882) ACC synthase
(AL096882.91_S_AT)	(AtACS-6) [Arabidopsis thaliana]
	gb AAC49356.1  (U35829) thioredoxin h
16981_s_at (U35829.2_S_AT)	[Arabidopsis thaliana]
	gb AAD23027.1 AC006585_22 (AC006585)
•	putative tyrosine aminotransferase
17008_at (AC006585.212_AT)	[Arabidopsis thaliana]
	gb AAC48925.1  (U05206) ribonuclease
17014_s_at (ATU05206_S_AT)	[Arabidopsis thaliana]
17500_s_at	emb CAB42906.1  (AL049862) calmodulin-
(ATHCALLGA_S_AT)	like protein [Arabidopsis thaliana]
	dbj BAA28538.1  (D78606) cytochrome P450
17522_s_at (D78606_S_AT)	monooxygenase [Arabidopsis thaliana]
	emb CAB38823.1  (AL035679) putative
17653_at (AL035679.144 AT)	protein [Arabidopsis thaliana]
17744 s at	gb AAC23646.1  (AC004684) putative
(AC004684.168_S_AT)	alcohol dehydrogenase [Arabidopsis thaliana]
17843 s at	gb AAB87109.1  (AC002391) putative
(AC002391.150 S AT)	cytochrome P450 [Arabidopsis thaliana]
:	emb CAB52675.1  (AJ010971) glucose-6-
	phosphate 1-dehydrogenase [Arabidopsis
18022_at (AJ010971_AT)	thaliana]
10022_40 (12010)/1_111)	gb AAB87112.1  (AC002391) putative
18551_at (AC002391.163 AT)	cytochrome P450 [Arabidopsis thaliana]
10351_ut (11C002351.103_A1)	
18716_at (X91916_AT)	gb AAF26754.1 AC007396_3 (AC007396)
10/10_at (X91910_A1)	T4012.6 [Arabidopsis thaliana]
19046 of (VI11700 1 AT)	emb CAA72484.1  (Y11788) peroxidase
18946_at (Y11788.1_AT)	ATP24a [Arabidopsis thaliana]
10171 -+ (4 0000225 1 0 4 50)	gb AAB64325.1  (AC002335) putative trypsin
19171_at (AC002335.160_AT)	inhibitor [Arabidopsis thaliana]
10100 . 0700000	emb CAA68191.1  (X99923) male sterility 2-
19177_at (X99923.1_AT)	like protein [Arabidopsis thaliana]
	emb CAA21214.1  (AL031804) putative
19182_at (AL031804.245_AT)	protein [Arabidopsis thaliana]
•	gb AAC27173.1  (AC003028) putative
19284_at (AC003028.196_AT)	anthocyanidin synthase [Arabidopsis thaliana]

gene.

19672\_at (AC005687.19 AT)

19762\_at (AL035527.204\_AT)

19892 at (AC005770.30 AT) 20232 s at

(AL022347.12 S AT)

20429\_s\_at (Z97336.167 S AT) protei [Arabidopsis thaliana]

20641\_at (X91919.1 AT)

gb|AAC36019.1| (AC005687) RAP2.6 [Arabidopsis thaliana] emb|CAB36812.1| (AL035527) peptide transporter-like protein [Arabidopsis thaliana] gb|AAC79626.1| (AC005770) putative protease inhibitor [Arabidopsis thaliana] emb|CAA18460.1| (AL022347) protein kinase-like protein [Arabidopsis thaliana] emb|CAB10219.1| (Z97336) hypothetical emb|CAA63012.1| (X91919) LEA76 homologue typel [Arabidopsis thaliana] Arabidopsis thaliana. ESTs gb|N97082,

gb|Z27056 and gb|Z29902 come from this

Duplicates -12905 and 16609 (ERF2)

13188 and 16981 (thioredoxin h) 12891 and 12892 (ACC synthase) 14838 (phosphoribosyl anthranilate transferase) has a duplicate in set 2 (14620). Counted in set 2

14254 (PAL1) has a duplicate in set 2 (14256) Counted in set 2. 12778 (lipase) has duplicates in set 2 (12777 and 12779) Counted in set 2.

# 4 rps2 without edm1 without pad4orNahG

ProbeSet Description

emb|CAA09731.1| (AJ011674) receptor-like 12278 at (AJ011674.2 AT) protein kinase, RLK3 [Arabidopsis thaliana]

> emb|CAA18468.1| (AL022347) serine/threonine kinase-like protein

12360 at (AL022347.131 AT) [Arabidopsis thaliana]

13154 s at

(AC002333.210 S AT)

13273\_s\_at (HSF4\_S\_AT)

gb|AAB64047.1| (AC002333) putative endochitinase [Arabidopsis thaliana]

gb[AAC31756.1] (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]

gb|AAD25552.1\AC005850 9 (AC005850) Highly Simlilar to Mlo proteins [Arabidopsis

13685 s\_at (MLOLIKE2\_S\_AT) thaliana]

14141\_at (NOVARTIS31\_AT) No hits found less than or equal to 1e-15.

emb|CAA19722.1| (AL030978) putative 15431\_at (AL030978.64 AT) protein [Arabidopsis thaliana] emb|CAA74639.1| (Y14251) glutathione S-16053 i at (Y14251.4 I AT) transferase [Arabidopsis thaliana] gb[AAB87114.1] (AC002391) unknown 16995 at (AC002391.188 AT) protein [Arabidopsis thaliana] emb|CAB54517.1| (AJ238846) SGP1 18054 at (AJ238846 AT) monomeric G-protein [Arabidopsis thaliana] 18224 s at emb|CAA17150.1| (AL021890) putative (AL021890.57 S AT) protein [Arabidopsis thaliana] gb|AAC42241.1| (AC005395) unknown 19951\_at (AC005395.47 AT) protein [Arabidopsis thaliana] emblCAA18469.1| (AL022347) serine/threonine kinase-like protein [Arabidopsis thaliana] gb|AAD25552.1|AC005850 9 (AC005850) Highly Simlilar to Mlo proteins [Arabidopsis thalianal

20223\_at (AL022347.145 AT)

20365 s at (AC005850.19 S AT)

#### **Duplicates**

12360 and 20223 (serine threonine kinase) 13685 and 20365 (similar to Mlo)

# 5 pad4orNahG without rps2 without edm1

ProbeSet

12307 at (AC002392.162 AT) 12332 s at (AB023448.2\_S\_AT)

13067 s at (AC003114.16 S AT)

13381\_at (AC006580.8 AT)

13588 at (AL021961.24 AT)

13751\_at (NOVARTIS127\_AT) F3M18.8 [Arabidopsis thaliana]

13764\_at (NOVARTIS22\_AT)

Description

gb|AAD12037.1| (AC002392) putative receptor-like protein kinase [Arabidopsis thaliana dbj|BAA82810.1| (AB023448) basic endochitinase [Arabidopsis thaliana] gb|AAC24083.1| (AC003114) Match to calreticulin (AtCRTL) mRNA gblU27698 and DNA gb|U66344. ESTs gb|T45719, gb|T22451, gb|H36323 and gb|AA042519 come from this gene. [Arabidopsis thaliana] gb[AAD22369.1]AC006580\_1 (AC006580) NAM (no apical meristem)-like protein [Arabidopsis thaliana] emb|CAA17552.1| (AL021961)

Phosphoglycerate dehydrogenase - like

protein [Arabidopsis thaliana]

gb|AAF16751.1|AC010155 4 (AC010155)

gb|AAD39641.1|AC007591\_6 (AC007591)

F9L1.6 [Arabidopsis thaliana]

gb|AAD25550.1|AC005850 7 (AC005850) 14146 at (NOVARTIS36 AT) Hypothetical protein [Arabidopsis thaliana] emb|CAA67234.1| (X98676) zinc finger 15778\_at (X98676.2\_AT) protein [Arabidopsis thaliana] emb|CAA67234.1| (X98676) zinc finger 15779 g at (X98676.2 G AT) protein [Arabidopsis thaliana] gb|AAD21491.1| (AC006587) unknown 15859\_at (AC006587.164 AT) protein [Arabidopsis thaliana] emb|CAA20203.1| (AL031187) receptor-like serine/threonine protein kinase ARK3 16360\_at (AL031187.126\_AT) [Arabidopsis thaliana] 16578 s at emb|CAB68132.1| (AL137080) beta-1, 3-(ATHRPRP1B\_S AT) glucanase 2 (BG2) [Arabidopsis thaliana] 16914 s at emb|CAB39936.1| (AL049500) osmotin (AL049500.57 S AT) precursor [Arabidopsis thaliana] gb|AAC49697.1| (U66345) calreticulin 17097 s at (ATU66345 S AT) [Arabidopsis thaliana] gb|AAD36959.1|AC000107 5 (AC000107) 17278\_at (AC000107.5 AT) F17F8.5 [Arabidopsis thaliana] emb|CAB10405.1| (Z97340) beta-1, 3glucanase class I precursor [Arabidopsis 17485\_s at (Z97340.345 S AT) thalianal gb|AAD11587.1|AAD11587 (AF071527) 19247 at (AF071527.44 AT) hypothetical protein [Arabidopsis thaliana] 20245 s at emb|CAA05625.1| (AJ002584) AtMRP4 (AC005309.97 S AT) [Arabidopsis thaliana] thaliana] emb|CAA19698.1| (AL024486) putative 20420 at (AL024486.131 AT) chitinase [Arabidopsis thaliana] emb|CAB41131.1| (AL049658) hypothetical 20625\_at (AL049658.102 AT) protein [Arabidopsis thaliana] Duplicates: 15778 and 15779 (zinc finger protein) 6 edm1 and rps2 without pad4orNahG ProbeSet Description gb|AAF18681.1|AF024504 11 (AF024504) 12630 at (AF024504.13 AT) unknown protein [Arabidopsis thaliana] gb|AAC06158.1| (AC003680) putative 13100 at (AC003680.50 AT)

cytochrome P450 [Arabidopsis thaliana] gb|AAB60774.1| (AC000375) ESTs gb|U75592,gb|T13956,gb|T43869 come from 13115 at (AC000375.44 AT) from this gene. [Arabidopsis thaliana] gb|AAC49356.1| (U35829) thioredoxin h (ATTHIRED4 I AT) [Arabidopsis thaliana] gb|AAC49356.1| (U35829) thioredoxin h (ATTHIRED4 S AT) [Arabidopsis thaliana]

13187 i at

13189 s at

13215_s_at (CAFFEROYLCOAMETHYLT RANS_S_AT)	gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:trans- caffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
13217_s_at (CALMODULINLIKE_S_AT)	emb CAB42906.1  (AL049862) calmodulin- like protein [Arabidopsis thaliana]
13277_i_at (HSP176A_I_AT)	emb CAA74399.1  (Y14070) Heat Shock Protein 17.6A [Arabidopsis thaliana]
13285_s_at (HSP83_S_AT)	gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC78535.1  (AC005662) putative
14083_at (AC005662.56_AT)	embryo-abundant protein [Arabidopsis thaliana]
	gb AAC26243.1  (AF077407) contains similarity to sugar transporters (Pfam:
14116_at (AF077407.30 AT)	sugar_tr.hmm, score: 395.39) [Arabidopsis thaliana]
	gb AAD31062.1 AC007357_11 (AC007357)
	Strong similarity to gb X97864 cytochrome P450 from Arabidopsis thaliana and is a
	member of the PF 00067 Cytochrome P450
	family. ESTs gb N65665, gb T14112,
14248_at (PAD3_AT)	gb T76255, gb T20906 and gb AI100027 come
	from this gene.
1400Z 1 at	
14682_i_at (WT1012A_RC_I_AT)	No hits found.
	gb AAD10829.1  (AF117063) putative
(WT1012A_RC_I_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2
	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine
(WT1012A_RC_I_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:trans-
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)  16105_s_at (ATU68017_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcrifeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)  16105_s_at (ATU68017_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1  (AC002294) similar to S-
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)  16105_s_at (ATU68017_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1  (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)  16105_s_at (ATU68017_S_AT)  16649_s_at (ATHORF_S_AT)  17511_s_at (AF067605_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1  (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana] emb CAB10339.1  (Z97339) hypothetical
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)  16105_s_at (ATU68017_S_AT)  16649_s_at (ATHORF_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1  (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana] emb CAB10339.1  (Z97339) hypothetical protein [Arabidopsis thaliana]
(WT1012A_RC_I_AT)  15641_s_at (AF117063_S_AT)  15978_at (X68592.6_AT)  16061_s_at (AB004796_S_AT)  16091_s_at (ATHHSP83_S_AT)  16105_s_at (ATU68017_S_AT)  16649_s_at (ATHORF_S_AT)  17511_s_at (AF067605_S_AT)	gb AAD10829.1  (AF117063) putative inositol polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] emb CAA48579.1  (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1  (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1  (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana] emb CAB10339.1  (Z97339) hypothetical

19640\_at (AC004561.78\_AT)

gb|AAC95192.1| (AC004561) putative glutathione S-transferase [Arabidopsis thaliana]

#### **Duplicates**

13187 and 13189 (thioredoxin h) There are two duplicates of this in set 3 (13188 and 16981) Count it in set 3.

13215 and 16649 (putative S-adenosyl-L-methionine:trans-caffeoyl-Coenzyme A 3-O-methyltransferase)
13285 and 16091 (heat shock protein 83)
13217 (calmodulin-like) has a duplicate in set 3 (17500). Count in set 3

16105 (hsf4) has a duplicate in set 4 (13273) Count in set 4

# 7 pad4orNahG and edm1 without rps2

ProbeSet

14110\_i\_at (AL035528.279\_I\_AT)

14148\_at (NOVARTIS38\_AT)

14249\_i\_at (PAD4\_I\_AT)

14250\_r\_at (PAD4\_R\_AT)

14672\_s\_at (TSA1\_S\_AT)

14673\_s\_at (TSB2\_S\_AT)

14704\_s\_at (WT768\_RC\_S\_AT) protein [Arabidopsis thaliana] 14709\_at (WT788\_AT) No hits found less than or equ

14882\_at (AL022605.63\_AT) 15616\_s\_at (ATHPRO25A S AT)

Description emb|CAB36854.1| (AL035528) putative disease resistance protein [Arabidopsis thaliana gb|AAF34713.1|AF224762 1 (AF224762) SigA binding protein [Arabidopsis thaliana] emb|CAB43438.1| (AL050300) putative protein [Arabidopsis thaliana] emb|CAB43438.1| (AL050300) putative protein [Arabidopsis thaliana] gb|AAC49117.1| (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana] gb|AAA32879.1| (M81620) tryptophan synthase beta-subunit [Arabidopsis thaliana] thaliana] gb|AAD15461.1| (AC006067) unknown No hits found less than or equal to 1e-15. emb|CAA18753.1| (AL022605) putative protein [Arabidopsis thaliana] emb|CAA08794.1| (AJ009696) wallassociated kinase 1 [Arabidopsis thaliana]

	•
15946 (4 500 60 65 60 450	gb AAD15461.1  (AC006067) unknown
15846_at (AC006067.63_AT)	protein [Arabidopsis thaliana]
15847_g_at	gb AAD15461.1  (AC006067) unknown
(AC006067.63_G_AT)	protein [Arabidopsis thaliana]
'-	emb CAA50677.1  (X71794) peroxidase
15970_s_at (X71794.2_S_AT)	[Arabidopsis thaliana]
	emb CAB41718.1  (AL049730) pEARLI 1
16150_s_at (ATHPEAR_S_AT)	[Arabidopsis thaliana]
	gb AAD40144.1 AF149413_25 (AF149413)
	contains similarity to protein kinase domains
•	(Pfam F00069, Score=162.6, E=6.8e-45, N=1)
	and leucien rich repeats (Pfam PF00560,
	Score=210.7, E=2.2e-59, N=10) [Arabidopsis
16357_at (AF149413.38_AT)	thaliana]
_ (	gb AAC04495.1  (AC003974) putative disease
16365_at (AC003974.136 AT)	resistance protein [Arabidopsis thaliana]
17068 s at	gb AAA32857.1  (M84658) receptor-like
(ATHRLPKA_S_AT)	protein kinase [Arabidopsis thaliana]
(	
17413_s_at (AJ006961.4_S_AT)	emb CAA67551.1  (X99097) peroxidase
17115_5_at (13000501.4_B_A1)	
	gb[AAD19610.1] (AF107726) cyclic
17499_s_at (AF107726_S_AT)	nucleotide gated channel [Arabidopsis
17422_3_at (AP107720_3_A1)	thaliana]
•	gb AAC28500.1  (AC004392) Similar to
	glucose-6-phosphate/phosphate-translocator
17775 at (AC004202 a AT)	(GPT) gb AF020814 from Pisum sativum.
17775_at (AC004392.2_AT)	[Arabidopsis thaliana]
19500 -4 (4 1999712 4 477)	emb CAA10955.1  (AJ222713) unnamed
18590_at (AJ222713.4_AT)	protein product [Arabidopsis thaliana]
	gb AAD45127.1 AF163823_1 (AF163823)
10000	endoxyloglucan transferase [Arabidopsis
18968_at (AF163823.4_AT)	thaliana]
100.00	gb AAD45127.1 AF163823_1 (AF163823)
18969_g_at	endoxyloglucan transferase [Arabidopsis
(AF163823.4_G_AT)	thaliana]
	emb CAA74930.1  (Y14590) class IV
20287_at (Y14590.5_AT)	chitinase [Arabidopsis thaliana]
	emb CAA74930.1  (Y14590) class IV
20288_g_at (Y14590.5_G_AT)	chitinase [Arabidopsis thaliana]
Duplicates	
14249 and 14250 (PAD4)	
•	· X ·
14704 and 15846 and 15847	
(unknown protein)	
•	

18968 and 18969 (endoxyloglucan transferase)

20287 and 20288 (class IV chitinase) 14110 (putative diseaseresistance gene) is an incomplete probe set with very low signal. It has a duplicate (14111) in set 2. Count in set 2. 14672 (trp synthase alpha) has a duplicate in set 2 (17487) Counted in set 2.

## 8 rps2 and pad4orNahG without edm1

ProbeSet Description emb|CAA23036.1| (AL035394) putative Na+/H+-exchanging protein [Arabidopsis 13627\_at (AL035394.196\_AT) thaliana emb|CAA18462.1| (AL022347) serine/threonine kinase-like protein 13659\_at (AL022347.46 AT) [Arabidopsis thaliana] gb|AAD15433.1| (AC006218) putative 13818 s at aspartate aminotransferase [Arabidopsis (AC006218.175\_S\_AT) thalianal gb|AAC69381.1| (AC005398) pathogenesisrelated PR-1-like protein [Arabidopsis 14635\_s\_at (PR.1\_S\_AT) thalianal 17128 s at gb|AAC69381.1| (AC005398) pathogenesis-(ATHRPRP1A S AT) related PR-1-like protein [Arabidopsis . emb|CAA07352.1| (AJ006960) peroxidase 17930\_s\_at (AJ006960.4\_S AT) [Arabidopsis thaliana] gb|AAC79593.1| (AC005727) unknown 19914\_at (AC005727.175 AT)

protein [Arabidopsis thaliana]

[Arabidopsis thaliana]

gb|AAD21459.1| (AC007017) similar to harpin-induced protein hin1 from tobacco

Duplicates:

14635 and 17128 (PR-1)

19991\_at (AC007017.124 AT)

### 9 edm1 and rps2 and pad4orNahG

ProbeSet Description emb|CAA16619.1| (AL021637) vacuolar 12341 s at sorting receptor-like protein [Arabidopsis (AL021637.176 S AT) thaliana gb|AAC49282.1| (U40856) AIG1 12879\_s at (AIG1\_S AT) [Arabidopsis thaliana]

12989 s at gb[AAC26690.1] (AC004077) putative (AC004077.149 S\_AT) cytochrome P450 [Arabidopsis thaliana] 13697 at (NI16 AT) No hits found. 14201 at (NOVARTIS73 AT) No hits found less than or equal to 1e-15. 14202 at (NOVARTIS73 RC AT) No hits found less than or equal to 1e-15. gb|AAC02748.1| (AC002340) putative 14609 at (AC002340.147\_AT) cytochrome P450 [Arabidopsis thaliana] emb|CAA50677.1| (X71794) peroxidase 14638\_s\_at (PRXCB\_S\_AT) [Arabidopsis thaliana] emb|CAA60521.1| (X86958) protein kinase catalytic domain (fragment) [Arabidopsis 14763 at (X86958.1 AT) thalianal gb|AAD28243.1|AF121356 1 (AF121356) 15116 f at (AF121356 F AT) peroxiredoxin TPx2 [Arabidopsis thaliana] gb|AAC49282.1| (U40856) AIG1 17544\_s at (ATU40856 S AT) [Arabidopsis thaliana] gb|AAB64049.1| (AC002333) putative 17840 s at (AC002333.223\_S\_AT) endochitinase [Arabidopsis thaliana] gb[AAC34217.1] (AC004411) putative 18567 at (AC004411.25 AT) alcohol dehydrogenase [Arabidopsis thaliana] gblAAB82640.1| (AC002387) putative 20269 at (AC002387.237 AT) pectinesterase [Arabidopsis thaliana]

Duplicates 12879 and 17544 (AIG1)

10

14201 and 14202 (Novartis 73) 14638 (peroxidase) has a duplicate in set 7 (15970). Counted in set 7.

The first set (set 2 above) do not require *EDM1* and are not affected by *rps2*, *pad4* or *NahG*. These genes are less likely to be important for resistance mediated by *RPP7* and *RPP8* than the genes in the second set (set 3 above). The genes in the second set require *EDM1* but are not affected by *rps2*, *pad4* or *NahG*. These genes are more likely to be important for resistance mediated by *RPP7* and *RPP8*. The genes in the third set (set 4 above) are affected by *rps2*, do not require *EDM1*, and are not affected by *pad4* or *NahG*. The genes in the fourth (set 5 above), seventh (set 8 above) and eight (set 9 above) sets include genes that are affected by *pad4* or *NahG*, and so are less likely to be important for resistance mediated by *RPP7* and *RPP8*. The genes in the sixth set (set 7 above) require *RPP7* or *RPP8* for increased expression after infection by

*Peronospora* and are affected by *rps2*. These genes are also less likely to be important for resistance mediated by *RPP7* and *RPP8*.

Duplicates were identified by sorting "RPP7 or 8" by description, which put duplicated genes adjacent to one another. Most duplicates were in the same set, but there were nine cases of duplicates that had one probe set in one set, and another in another set. Examination of the data revealed that in these cases, the expression values were close to the cut-offs used to define the various sets. If one copy met the criteria, e.g., for *rps2*-dependent, but the other copy did not, the gene was classified as not *rps2*-dependent. Duplicates are noted. One hundred eighty four unique genes remained (SEQ ID NOs:1-211 and 792, Table 9 and Table 10).

#### Table 9

5

10

SEQ ID		
NO	ProbeSet	Description
	12007_at	emb CAB16829.1  (Z99708) putative protein
1	(Z99708.249_AT)	[Arabidopsis thaliana]
	12091_at	gb AAC64313.1  (AC004450) unknown
2 .	(AC004450.116_AT)	
	12115_at	emb CAA22152.1  (AL033545) extensin-like
3	(AL033545.26_AT)	protein [Arabidopsis thaliana]
4	12240_at	emb CAB43974.1  (AL078579) putative
4	(AL078579.130_AT)	protein [Arabidopsis thaliana]
	12278 at	emb CAA09731.1  (AJ011674) receptor-like
5	(AJ011674.2_AT)	protein kinase, RLK3 [Arabidopsis thaliana]
-	(	gb AAD12037.1  (AC002392) putative
•	12307_at	receptor-like protein kinase [Arabidopsis
6	(AC002392.162_AT)	thaliana]
	12332_s_at	dbj BAA82810.1  (AB023448) basic
7	(AB023448.2_S_AT)	
	12341_s_at	emb CAA16619.1  (AL021637) vacuolar
	(AL021637.176_S_A	
8	T)	thaliana]
	12349_s_at	gb AAA17993.1  (M91192) phenylalanine
9	(X84728.6_S_AT)	ammonia-lyase [Trifolium subterraneum]
	10060	emb CAA18468.1  (AL022347)
10	12360_at	serine/threonine kinase-like protein
10	(AL022347.131_AT)	
	20222 at	emb CAA18468.1  (AL022347)
11	20223_at	serine/threonine kinase-like protein
11	(AL022347.145_AT)	[Arabidopsis thaliana]

12 13	12500_s_at (AF081067.3_S_AT) 12521_at (AF049236.28_AT)	gb AAC32192.1  (AF081067) IAA-Ala hydrolase; IAA-amino acid hydrolase [Arabidopsis thaliana] gb AAC14413.1  (AF049236) unknown [Arabidopsis thaliana]
14	12538_at (AF033205.2_AT)	gb AAC02973.1  (AF033205) putative pectin methylesterase [Arabidopsis thaliana] emb CAB45330.1  (AL079344) nucleotide
15	12556_at (AL079344.155_AT) 12574 at	pyrophosphatase-like protein [Arabidopsis thaliana] emb CAA57944.1  (X82624) SRG2At
16	(X82624.2_AT)	[Arabidopsis thaliana]
17	12630_at (AF024504.13_AT) 12642_at	gb AAF18681.1 AF024504_11 (AF024504) unknown protein [Arabidopsis thaliana] gb AAD22285.1 AC006920_9 (AC006920)
18	(AC006920.138_AT)	unknown protein [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577)
		Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb A1099570 and
19	12777_i_at (AC006577.16_I_AT)	gb T22281 come from this gene. [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577)
	10770	Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815,
20 .	12778_r_at (AC006577.16_R_AT )	gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene. [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577)
		Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815,
	12779_f_at	gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene.
21	(AC006577.16_F_AT) 12879 s at	[Arabidopsis thaliana] gb AAC49282.1  (U40856) AIG1
22	(AIG1_S_AT) 17744 s at	[Arabidopsis thaliana]
23	— <b>—</b>	gb AAC23646.1  (AC004684) putative alcohol dehydrogenase [Arabidopsis thaliana]
24		gb AAA32738.1  (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana]
25	20291_s_at (M92353.4_S_AT)	gb AAA32738.1  (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana]

	•	
26	12891_at (ATACS6_AT)	gb AAC63850.1  (U73786) ACC synthase [Arabidopsis thaliana]
	12892 g at	gb AAC63850.1  (U73786) ACC synthase
27	(ATACS6_G_AT)	[Arabidopsis thaliana] dbj BAA32419.1  (AB008104) ethylene
	12905_s_at	responsive element binding factor 2
28	(ATERF2_S_AT)	[Arabidopsis thaliana]
,	16600	dbj BAA32419.1  (AB008104) ethylene
29	16609_s_at (AB008104_S_AT)	responsive element binding factor 2 [Arabidopsis thaliana]
2)	(11D000104_5_A1)	dbj BAA32422.1  (AB008107) ethylene
	12908_s_at	responsive element binding factor 5
30	(ATERF5_S_AT)	[Arabidopsis thaliana]
	16536	dbj BAA32422.1  (AB008107) ethylene
31	16536_s_at (AB008107_S_AT)	responsive element binding factor 5 [Arabidopsis thaliana]
•	12916 s at	gb AAC13947.1  (AF021244) coronatine-
32	(ATHCOR1_S_AT)	induced protein 1 [Arabidopsis thaliana]
22	12933_r_at	emb CAA65420.1  (X96600) pathogenesis-
33	(ATPRITAN_R_AT) 12989 s at	related protein 1 [Arabidopsis thaliana]
	<del></del>	gb AAC26690.1  (AC004077) putative
34	T)	cytochrome P450 [Arabidopsis thaliana]
		gb AAC24083.1  (AC003114) Match to
	•	calreticulin (AtCRTL) mRNA gb U27698
	13067 s at	and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519
35		come from this gene. [Arabidopsis thaliana]
•	13100_at	gb AAC06158.1  (AC003680) putative
36	(AC003680.50_AT)	cytochrome P450 [Arabidopsis thaliana]
		gb AAB60774.1  (AC000375) ESTs
37	13115_at (AC000375.44_AT)	gb U75592,gb T13956,gb T43869 come from
31	13154 s at	from this gene. [Arabidopsis thaliana]
		gb AAB64047.1  (AC002333) putative
38	T)	endochitinase [Arabidopsis thaliana]
39	13176_at	emb CAA20567.1  (AL031394) putative
39		protein [Arabidopsis thaliana] gb AAC49356.1  (U35829) thioredoxin h
40	(ATTHIRED4_I_AT)	[Arabidopsis thaliana]
	13188_r_at	gb AAC49356.1  (U35829) thioredoxin h
41	(ATTHIRED4_R_AT)	[Arabidopsis thaliana]
42	13189_s_at	gb AAC49356.1  (U35829) thioredoxin h
74	(ATTHIRED4_S_AT) 16981 s at	gb AAC49356.1  (U35829) thioredoxin h
43	(U35829.2_S_AT)	[Arabidopsis thaliana]
		emb CAA16892.1  (AL021749) 12S
11	13198_i_at	cruciferin seed storage protein [Arabidopsis
44	(ATTS0190_I_AT)	thaliana]

45	13215_s_at (CAFFEROYLCOAM ETHYLTRANS_S_A T)	gb AAF16576.1 AC012563_29 (AC012563)  I putative S-adenosyl-L-methionine:trans- caffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
46	16649_s_at (ATHORF_S_AT) 13217 s at	gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:trans- caffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
47	(CALMODULINLIK E_S_AT) 17500_s_at	emb CAB42906.1  (AL049862) calmodulin- like protein [Arabidopsis thaliana]
48	(ATHCALLGA_S_A T)	emb CAB42906.1  (AL049862) calmodulin- like protein [Arabidopsis thaliana]
49	13273_s_at (HSF4_S_AT)	gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
50	16105_s_at (ATU68017_S_AT) 13277 i at	gb AAC31756.1  (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] emb CAA74399.1  (Y14070) Heat Shock
51	(HSP176A_I_AT)	Protein 17.6A [Arabidopsis thaliana]
52	13285_s_at (HSP83_S_AT) 16091_s_at	gb AAA32822.1  (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAA32822.1  (M62984) heat shock
53	(ATHHSP83_S_AT)	protein 83 [Arabidopsis thaliana] gb AAD22369.1 AC006580_1 (AC006580)
54	13381_at (AC006580.8_AT)	NAM (no apical meristem)-like protein [Arabidopsis thaliana]
	13435_at (AF003102.3_AT)	gb AAC49775.1  (AF003102) AP2 domain containing protein RAP2.9 [Arabidopsis thaliana]
55	13588_at (AL021961.24_AT)	emb CAA17552.1  (AL021961) Phosphoglycerate dehydrogenase - like protein [Arabidopsis thaliana] emb CAA23036.1  (AL035394) putative
56		Na+/H+-exchanging protein [Arabidopsis thaliana] gb AAB82634.1  (AC002387) putative
57	(AC002387.185_AT)	transketolase precursor [Arabidopsis thaliana] emb CAA18462.1  (AL022347)
58	13659_at (AL022347.46_AT)	serine/threonine kinase-like protein [Arabidopsis thaliana] gb AAD25552.1 AC005850_9 (AC005850)
59	13685_s_at	Highly Similar to Mlo proteins [Arabidopsis thaliana]

	•	
•	0004	gb AAD25552.1 AC005850_9 (AC005850)
	20365_s_at	Highly Simlilar to Mlo proteins [Arabidopsis
60	(AC005850.19_S_AT	
61	13696_at (NII15_AT)	No hits found less than or equal to 1e-15.
no cds	13697_at (NI16_AT)	
	13751_at	gb AAF16751.1 AC010155_4 (AC010155)
62		F3M18.8 [Arabidopsis thaliana]
·a -	13764_at	gb AAD39641.1 AC007591_6 (AC007591)
63	(NOVARTIS22_AT)	F9L1.6 [Arabidopsis thaliana]
	13818_s_at	gb AAD15433.1  (AC006218) putative
	(AC006218.175_S_A	aspartate aminotransferase [Arabidopsis
64	T)	thaliana]
	13880_s_at	emb CAB39611.1  (AL049480) possible
	(AL049480.183_S_A	
65	T)	[Arabidopsis thaliana]
	13966_at	emb CAA17775.1  (AL022023) putative
66 -	(AL022023.172_AT)	protein [Arabidopsis thaliana]
		gb AAC78535.1  (AC005662) putative
	14083_at	embryo-abundant protein [Arabidopsis
67	(AC005662.56_AT)	thaliana]
	14096_at	
68	(AC002291.12_AT)	No hits found.
	14110_i_at	emb CAB36854.1  (AL035528) putative
	(AL035528.279_I_AT	disease resistance protein [Arabidopsis
69	)	thaliana]
	14111_s_at	emb CAB36854.1  (AL035528) putative
	(AL035528.279_S_A	disease resistance protein [Arabidopsis
70	T)	thaliana]
		gb AAC26243.1  (AF077407) contains
		similarity to sugar transporters (Pfam:
· .	14116_at	sugar_tr.hmm, score: 395.39) [Arabidopsis
71	(AF077407.30_AT)	thaliana]
	14141_at	•
72	(NOVARTIS31_AT)	No hits found less than or equal to 1e-15.
•		dbj BAA22813.1  (D26015) CND41,
	14145_at	chloroplast nucleoid DNA binding protein
73	(NOVARTIS35_AT)	[Nicotiana tabacum]
	1/11/6 -4	114 4705550 114 5005050 7 (4 5005050)
74	14146_at	gb AAD25550.1 AC005850_7 (AC005850)
/4	(NOVARTIS36_AT)	Hypothetical protein [Arabidopsis thaliana]
•	14148 at	gb AAF34713.1 AF224762_1 (AF224762)
75	<del></del>	SigA binding protein [Arabidopsis thaliana]
. •	14201 at	organ omaing protein [Arabidopsis diamana]
76		No hits found less than or equal to 1e-15.
, 0	14202 at	140 lins found less than of equal to 1e-15.
	(NOVARTIS73_RC	
77		No hits found less than an agual to 15 15
	14223 at	No hits found less than or equal to 1e-15.
78	<del>-</del>	emb CAA19683.1  (AL024486) putative protein [Arabidopsis thaliana]
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7 <u>.</u>	14232_at (NOVARTIS95_AT)	gb AAF16756.1 AC010155_9 (AC010155) F3M18.20 [Arabidopsis thaliana]
	,	gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to gb X97864 cytochrome P450 from Arabidopsis thaliana and is a member of the PF 00067 Cytochrome P450 family. ESTs gb N65665, gb T14112, gb T76255, gb T20906 and gb AI100027
80	14248_at (PAD3_AT) 14249 i at	come from this gene. emb CAB43438.1  (AL050300) putative
81	(PAD4_I_AT) 14250_r_at	protein [Arabidopsis thaliana] emb CAB43438.1  (AL050300) putative
82	(PAD4_R_AT)	protein [Arabidopsis thaliana]
83	14254_s_at (PAL1- MRNA_S_AT)	gb AAD18156.2  (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis gb AAD18156.2  (AC006260) phenylalanine
84	14256_f_at (PAL1- INTRON_F_AT)	ammonia lyase (PAL1) [Arabidopsis thaliana]
	14278 at	emb CAA18503.1  (AL022373) hypothetical
85	(AL022373.295_AT)	protein [Arabidopsis thaliana] gb AAD23719.1 AC005956_8 (AC005956)
	14320_at	putative RING zinc finger protein
86	(AC005956.54_AT)	[Arabidopsis thaliana]
0.5	14381_at	gb AAC05341.1  (AC002521) unknown
87	(AC002521.68_AT)	protein [Arabidopsis thaliana]
88	14443_at (AC000348.23 AT)	gb AAB61498.1  (AC000348) T7N9.22
00	14609 at	[Arabidopsis thaliana] gb AAC02748.1  (AC002340) putative
89	(AC002340.147_AT)	cytochrome P450 [Arabidopsis thaliana]
	14614_at	gb AAC16958.1  (AC004165) putative
90	(AC004165.66_AT)	glucosyltransferase [Arabidopsis thaliana]
		gb AAA32835.1  (M96073)
Λ1	14620 s_at	phosphoribosylanthranilate transferase
91	(PATI_S_AT)	[Arabidopsis thaliana]
	14838 s at	gb AAA32835.1  (M96073) phosphoribosylanthranilate transferase
92	(M96073.6_S_AT)	[Arabidopsis thaliana]
	14621 at	gb AAC31244.1  (AC004747) putative
93	(PDF1.2_AT)	antifungal protein [Arabidopsis thaliana]
	_ ,	gb AAC69381.1  (AC005398) pathogenesis-
4.	14635_s_at	related PR-1-like protein [Arabidopsis
94		thaliana]
	17128_s_at	gb AAC69381.1  (AC005398) pathogenesis-
95	(AIHKPKPIA_S_AT	related PR-1-like protein [Arabidopsis
) )	14638 s at	thaliana]
96	(PRXCB_S_AT)	emb CAA50677.1  (X71794) peroxidase [Arabidopsis thaliana]
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97	15970_s_at (X71794.2_S_AT)	emb CAA50677.1  (X71794) peroxidase [Arabidopsis thaliana]
98	14672_s_at (TSA1_S_AT)	gb AAC49117.1  (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
99	17487_s_at (U18993.2_S_AT)	gb AAC49117.1  (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana] gb AAA32879.1  (M81620) tryptophan
100	14673_s_at (TSB2_S_AT) 14682 i at	synthase beta-subunit [Arabidopsis thaliana] thaliana]
101	(WT1012A_RC_I_A7	Γ΄ No hits found.
102	14691_at (WT1096_AT)	No hits found.
103	14704_s_at (WT768_RC_S_AT) 15846_at	gb AAD15461.1  (AC006067) unknown protein [Arabidopsis thaliana] gb AAD15461.1  (AC006067) unknown
104	(AC006067.63_AT) 15847_g_at	protein [Arabidopsis thaliana]
105	)	gb AAD15461.1  (AC006067) unknown protein [Arabidopsis thaliana]
106	14709_at (WT788_AT)	No hits found less than or equal to 1e-15.
107	14763_at (X86958.1_AT)	emb CAA60521.1  (X86958) protein kinase catalytic domain (fragment) [Arabidopsis thaliana]
108	14882_at (AL022605.63_AT)	emb CAA18753.1  (AL022605) putative protein [Arabidopsis thaliana] emb CAA17549.1  (AL021961) cinnamyl
109	15042_at (AL021961.3_AT)	alcohol dehydrogenase - like protein [Arabidopsis thaliana]
110	15116_f_at (AF121356_F_AT)	gb AAD28243.1 AF121356_1 (AF121356) peroxiredoxin TPx2 [Arabidopsis thaliana] gb AAB53975.1  (U90522) lysine-
111	15161_s_at (ATU90522_S_AT) 15366 at	ketoglutarate reductase/saccharopine dehydrogenase [Arabidopsis thaliana]
112	(U93215.38_AT) 15415_at	gb AAB63077.1  (U93215) unknown protein [Arabidopsis thaliana] gb AAB72169.1  (AF000657) hypothetical
113	(AF000657.28_AT) 15431_at,	protein [Arabidopsis thaliana] emb CAA19722.1  (AL030978) putative
114	(AL030978.64_AT) 15523_s_at	protein [Arabidopsis thaliana]
115	(ALU/8637.213_S_A T) 15532 r at	emb CAB45071.1  (AL078637) putative protein [Arabidopsis thaliana]
116		emb CAB45069.1  (AL078637) putative protein [Arabidopsis thaliana]

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	15616_s_at (ATHPRO25A_S_AT	emb CAA08794.1  (AJ009696) wall-
117	)	associated kinase 1 [Arabidopsis thaliana]
	15629 s at	dbj BAA24440.1  (AB010407)
118	(AB003280_S_AT)	phosphoglycerate dehydrogenase [Arabidopsis thaliana]
	(	gb AAD10829.1  (AF117063) putative
	15641_s_at	inositol polyphosphate 5-phosphatase At5P2
119	(AF117063_S_AT)	[Arabidopsis thaliana]
		gb AAB80922.1  (AF022658) putative c2h2
100- 600	15665_s_at	zinc finger transcription factor [Arabidopsis
120, 082	(AF022658_S_AT)	thaliana]
121	15778_at (X98676.2 AT)	emb CAA67234.1  (X98676) zinc finger
121	15779_g_at	protein [Arabidopsis thaliana] emb CAA67234.1  (X98676) zinc finger
122	(X98676.2 G AT)	protein [Arabidopsis thaliana]
		gb AAD15461.1  (AC006067) unknown
123	(AC006587.164_AT)	protein [Arabidopsis thaliana]
	15978_at	gb AAD15461.1  (AC006067) unknown
124	(X68592.6_AT)	protein [Arabidopsis thaliana]
105	16053_i_at	emb CAA74639.1  (Y14251) glutathione S-
125	(Y14251.4_I_AT) 16061_s at	transferase [Arabidopsis thaliana]
126	(AB004796_S_AT)	gb AAB97145.1  (AF000977) MEK1
120	(AD004790_3_A1)	[Arabidopsis thaliana] thaliana] gb AAD34615.1 AF153283_1 (AF153283)
	16083_s_at	putative progesterone-binding protein
127	(AF153283_S_AT)	homolog [Arabidopsis thaliana]
	16150_s_at	emb CAB41718.1  (AL049730) pEARLI 1
128	(ATHPEAR_S_AT)	[Arabidopsis thaliana]
100	16232_s_at	emb CAB45796.1  (AL080252) putative
129	(AL080252.77_S_AT)	protein [Arabidopsis thaliana]
	16257 at	gb AAC32915.1  (AC004138) putative
130	(AC004138.105_AT)	nucleoside triphosphatase [Arabidopsis thaliana]
200	16298_at	emb CAA17152.1  (AL021890) putative
131	(AL021890.71 AT)	protein [Arabidopsis thaliana]
	16299_at	emb CAA19705.1  (AL024486) putative
132	(AL024486.185_AT)	protein [Arabidopsis thaliana]
		gb AAD40144.1 AF149413 25 (AF149413)
		contains similarity to protein kinase domains
		(Pfam F00069, Score=162.6, E=6.8e-45,
		N=1) and leucien rich repeats (Pfam
122	16357_at	PF00560, Score=210.7, E=2.2e-59, N=10)
133	(AF149413.38_AT)	[Arabidopsis thaliana]
	16360 at	emb CAA20203.1  (AL031187) receptor-like
134	(AL031187.126_AT)	serine/threonine protein kinase ARK3
'	(	fr manachara menenal

135	16365_at (AC003974.136_AT) 16578 s at	gb AAC04495.1  (AC003974) putative disease resistance protein [Arabidopsis thaliana]
136		emb CAB68132.1  (AL137080) beta-1, 3-glucanase 2 (BG2) [Arabidopsis thaliana]
137	16817_s_at (AL096882.91_S_AT) 16914_s_at	emb CAB51412.1  (AL096882) ACC synthase (AtACS-6) [Arabidopsis thaliana] emb CAB39936.1  (AL049500) osmotin
138		precursor [Arabidopsis thaliana] gb AAB87114.1  (AC002391) unknown
139	•	protein [Arabidopsis thaliana] gb AAD23027.1 AC006585_22 (AC006585)
140	17008_at (AC006585.212_AT) 17014_s at	putative tyrosine aminotransferase [Arabidopsis thaliana] gb[AAC48925.1] (U05206) ribonuclease
141	(ATU05206_S_AT) 17051 s at	[Arabidopsis thaliana] gb AAD09952.1  (AF098947) CTF2B
142	(AF098947_S_AT) 17068_s_at	[Arabidopsis thaliana] gb AA32857.1  (M84658) receptor-like
143	(ATHRLPKA_S_AT)	protein kinase [Arabidopsis thaliana]
144	17083_s_at (ATU18770_S_AT)	gb AAD25838.1 AC006951_17 (AC006951) putative indole-3-glycerol phosphate synthase [Arabidopsis thaliana]
145	17097_s_at (ATU66345_S_AT) 17278 at	gb AAC49697.1  (U66345) calreticulin [Arabidopsis thaliana] gb AAD36959.1 AC000107_5 (AC000107)
146	(AC000107.5_AT) 17413_s at	F17F8.5 [Arabidopsis thaliana] emb CAA67551.1  (X99097) peroxidase
147	(AJ006961.4_S_AT)	[Arabidopsis thaliana] gb AAB60752.1  (AC000132) Similar to A. thaliana receptor-like protein kinase
1.40	17464_at	(gb RLK5_ARATH). ESTs gb ATTS0475,gb ATTS4362 come from this
148	(AC000132.72_AT)	gene. [Arabidopsis thaliana] emb CAB10405.1  (Z97340) beta-1, 3-
149		glucanase class I precursor [Arabidopsis thaliana] gb AAD19610.1  (AF107726) cyclic
150	(AF107726_S_AT)	nucleotide gated channel [Arabidopsis thaliana]
151	17511_s_at	gb AAB71482.1  (AC002294) similar to S- linalool synthase gp U58314 1491939 [Arabidopsis thaliana]
152	(D78606_S_AT)	dbj BAA28538.1  (D78606) cytochrome P450 monooxygenase [Arabidopsis thaliana]
153		gb AAC49282.1  (U40856) AIG1 [Arabidopsis thaliana]

154	17653_at (AL035679.144_AT)	emb CAB38823.1  (AL035679) putative protein [Arabidopsis thaliana] gb AAC28500.1  (AC004392) Similar to glucose-6-phosphate/phosphate-translocator
155	17775_at (AC004392.2_AT) 17840 s at	(GPT) gb AF020814 from Pisum sativum. [Arabidopsis thaliana]
156		gb AAB64049.1  (AC002333) putative endochitinase [Arabidopsis thaliana]
157	(AC002391.150_S_A T) 17899 at	gb AAB87109.1  (AC002391) putative cytochrome P450 [Arabidopsis thaliana] emb CAB10339.1  (Z97339) hypothetical
158	(Z97339.197_AT)	protein [Arabidopsis thaliana]
159	17930_s_at (AJ006960.4_S_AT)	emb CAA07352.1  (AJ006960) peroxidase [Arabidopsis thaliana] emb CAB59428.1  (AJ002295) inositol-
160	18012_s_at (AJ002295_S_AT)	1,4,5-trisphosphate 5-Phosphatase [Arabidopsis thaliana] emb CAB52675.1  (AJ010971) glucose-6-
161	18022_at (AJ010971_AT)	phosphate 1-dehydrogenase [Arabidopsis thaliana]
162	18054_at (AJ238846_AT)	emb CAB54517.1  (AJ238846) SGP1 monomeric G-protein [Arabidopsis thaliana]
163	18216_at (X95573.2_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana]
164	18217_g_at (X95573.2_G_AT) 18551_at	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana] gb AAB87112.1  (AC002391) putative
165	(AC002391.163_AT)	cytochrome P450 [Arabidopsis thaliana] gb AAC34217.1  (AC004411) putative
166	18567_at (AC004411.25_AT) 18590 at	alcohol dehydrogenase [Arabidopsis thaliana] emb CAA10955.1  (AJ222713) unnamed
167	(AJ222713.4_AT)	protein product [Arabidopsis thaliana] emb CAA52772.1  (X74756) ATAF2
168; 665	18591_at (X74756.2_AT)	[Arabidopsis thaliana] gb AAC72125.1  (AC005278) ESTs gb H36966, gb R65511, gb T42324 and
169	18625_at (AC005278.22_AT) 18716_at	gb T20569 come from this gene. [Arabidopsis thaliana] gb AAF26754.1 AC007396_3 (AC007396)
170	(X91916_AT)	T4O12.6 [Arabidopsis thaliana] gb AAC33239.1  (AC005315) putative
171	18908_i_at	ligand-gated ion channel protein [Arabidopsis thaliana] gb AAC62611.1  (AF055848) subtilisin-like
172	(AF055848.2_I_AT)	protease [Arabidopsis thaliana]

1		18909_s_at	gb AAC62611.1  (AF055848) subtilisin-like
J	173	(AF055848.2_S_AT)	protease [Arabidopsis thaliana]
1	71	18946_at	emb CAA72484.1  (Y11788) peroxidase
j	174	(Y11788.1_AT)	ATP24a [Arabidopsis thaliana]
		10000	gb AAD45127.1 AF163823_1 (AF163823)
1	175	18968_at	endoxyloglucan transferase [Arabidopsis
	13	(AF163823.4_AT)	thaliana]
,		18969 g at	gb AAD45127.1 AF163823_1 (AF163823)
. 1	76	(AF163823.4_G_AT)	endoxyloglucan transferase [Arabidopsis thaliana]
1	70	18983 s at	diananaj
		- · · · · ·	emb CAB41722.1  (AL049730) pEARLI 1-
1	.77	T)	like protein [Arabidopsis thaliana]
		19158 at	emb CAA65053.1  (X95738) proline
1	78	(X95738.2_AT)	transporter 2 [Arabidopsis thaliana]
		19171 at	gb AAB64325.1  (AC002335) putative
1	79	(AC002335.160 AT)	trypsin inhibitor [Arabidopsis thaliana]
		19177 at	emb CAA68191.1  (X99923) male sterility 2-
1	80	(X99923.1_AT)	like protein [Arabidopsis thaliana]
		19182 at	emb CAA21214.1  (AL031804) putative
1	81	(AL031804.245 AT)	protein [Arabidopsis thaliana]
_		19229_at	gb AAD10694.1  (AC003027) lcl prt_seq No
1	.82	(AC003027.39_AT)	definition line found [Arabidopsis thaliana]
		19247 at	gb AAD11587.1 AAD11587 (AF071527)
1	.83	(AF071527.44_AT)	hypothetical protein [Arabidopsis thaliana]
			gb AAC27173.1  (AC003028) putative
		19284_at	anthocyanidin synthase [Arabidopsis
1	.84	(AC003028.196_AT)	thaliana]
		19415_at	emb CAB45805.1  (AL080253) putative
1	.85	(AL080253.32_AT)	protein [Arabidopsis thaliana]
_	0.0	19594_i_at	emb CAA66965.1  (X98321) peroxidase
1	86	(X98321.2_I_AT)	[Arabidopsis thaliana]
		10640 -4	gb AAC95192.1  (AC004561) putative
1	87	19640_at	glutathione S-transferase [Arabidopsis
1	.07	(AC004561.78_AT)	thaliana]
		19664 at	emb CAB43638.1  (AL050351) NAD(P)H oxidoreductase, isoflavone reductase-like
1	88	(AL050351.172 AT)	protein [Arabidopsis thaliana]
•		19672 at	gb AAC36019.1  (AC005687) RAP2.6
i	89	(AC005687.19 AT)	[Arabidopsis thaliana]
_		(	emb CAB36812.1  (AL035527) peptide
		19762 at	transporter-like protein [Arabidopsis
1	90	(AL035527.204 AT)	
		19892 at	gb AAC79626.1  (AC005770) putative
l	91	(AC005770.30_AT)	protease inhibitor [Arabidopsis thaliana]
			emb CAA05025.1  (AJ001809) succinate
		19894_at	dehydrogenase flavoprotein alpha subunit
1	92	(AJ001809.1_AT)	[Arabidopsis thaliana]

	19914 at	gb AAC79593.1  (AC005727) unknown
193	(AC005727.175 AT)	protein [Arabidopsis thaliana]
	19951 at	gb AAC42241.1  (AC005395) unknown
194	(AC005395.47 AT)	protein [Arabidopsis thaliana]
	_ /	gb AAD21459.1  (AC007017) similar to
	19991 at	harpin-induced protein hin1 from tobacco
195	(AC007017.124 AT)	[Arabidopsis thaliana]
	20189 at	gb AAD32864.1 AC005489 2 (AC005489)
196	(AC005489.2_AT)	F14N23.2 [Arabidopsis thaliana]
		emb CAA18469.1  (AL022347)
	20232_s_at	serine/threonine kinase-like protein
197	(AL022347.12_S_AT)	[Arabidopsis thaliana]
	20238_at	emb CAA52619.1  (X74514) beta-
198	(X74514.2_AT)	fructofuranosidase [Arabidopsis thaliana]
	20239_g_at	emb CAA52619.1  (X74514) beta-
199	(X74514.2_G_AT)	fructofuranosidase [Arabidopsis thaliana]
	20245_s_at	emb CAA05625.1  (AJ002584) AtMRP4
200		[Arabidopsis thaliana] thaliana]
	20246_s_at	gb AAC95354.1  (AF084037) receptor-like
201	(AF084037.3_S_AT)	protein kinase [Arabidopsis thaliana]
000	20269_at	gb AAB82640.1  (AC002387) putative
202	(AC002387.237_AT)	pectinesterase [Arabidopsis thaliana]
.002	20287_at	emb CAA74930.1  (Y14590) class IV
203	(Y14590.5_AT)	chitinase [Arabidopsis thaliana]
204	20288_g_at	emb CAA74930.1  (Y14590) class IV
204	(Y14590.5_G_AT)	chitinase [Arabidopsis thaliana]
	20331 at	gb AAD23617.1 AC007168_8 (AC007168) putative aspartate aminotransferase
205	(AC007168.86 AT)	[Arabidopsis thaliana]
203	20368 at	gb AAC36163.1  (AC005314) putative serpin
206	(AC005314.38 AT)	[Arabidopsis thaliana]
	20420 at	emb CAA19698.1  (AL024486) putative
207	(AL024486.131 AT)	chitinase [Arabidopsis thaliana]
	20429_s_at	emb CAB10219.1  (Z97336) hypothetical
208		protei [Arabidopsis thaliana]
	20620 g at	gb AAC98070.1  (AC005896) putative
	(AC005896.161_G_A	C2H2-type zinc finger protein [Arabidopsis
209	T)	thaliana]
•		
010	20625_at	emb CAB41131.1  (AL049658) hypothetical
210	(AL049658.102_AT)	protein [Arabidopsis thaliana]
		emb CAA63012.1  (X91919) LEA76
		homologue typel [Arabidopsis thaliana]
	20641 at	Arabidopsis thaliana. ESTs gb N97082,
211	(X91919.1_AT)	gb Z27056 and gb Z29902 come from this gene.
211	18224 s at	emb CAA17152.1  (AL021890) putative
789	<del></del>	protein [Arabidopsis thaliana]
. 05	16522 at	emb CAA54631.1  (X77500) amino acid
790	(X77500.2_AT)	transporter [Arabidopsis thaliana]
•	(= · · · · · · · · · · · · · · · · · · ·	

		gb AAC17040.1  (AC002986) Similarity to
		A. thaliana gene product F21M12.20,
	19982 at	gb AC000132. EST gb Z25651 comes from
791	(AC002986.28_AT)	this gene. [Arabidopsis thaliana]
	18920 at	gb AAC16927.1  (AC002338) putative
367	(AC002338.11 AT)	laccase [Arabidopsis thaliana]
212	12324 i at	AC007212
793	12345 at	L36246
213	12500_s_at	AF081067
	12505_s_at	AC005309
215	12608 i at	S70188
216	12642 at	AC006920
217	12746 i at	AL096882
218	12748 f at	AL096882
219	12761 s at	AC006577
220	<del></del>	AC005727
221	12778_r_at	AC006577
222	12778 at	AC003028
223	12802 at	AL022373
223	12842_s_at	No hits found less than or equal to 1e-15
224	12843_s_at	AC007195
227	12845 s at	AJ004810
225	12879 s at	U40856
226	12891 at	U73786
227	12892 g at	U73786
	12904 s at	AB008103
229	12905_s_at	AB008104
230	12908_s_at	AB008107
231	12909 s at	Z97343
232	12905_s_at	X84229
233	12916 s at	AF021244
234	13138 at	AL096882
235	13177_at	AL049640
236	13177_at	U93215
237	13187 i at	U35829
238	13189 s at	U35829
239	13198 i at	AL021749
240	13212 s at	AL137080
241	13217 s at	AL049862
242	13258 s at	AC005309
243	13273 s at	U68017
244	13284 s at	AJ002551
245	13604 at	AC000104
246	13615 at	AC002332
247	13617 at	AC006592
248	13718 at	Z72152
249	13771 at	AC006593
250	13771_at	AC007987
251	14052 at	AC004122
201	14032_at	ACCOUT144

252	14096 at	AC002291
253	14097_at	AC005309
254	14116 at	AF077407
255	14141_at	AC011437
256; 683	14148 at	AF224762
257	14196 at	AC012563
258	14201 at	AL163972
259	14219 at	AC068667
260	14223 at	AL024486
261	14248 at	AC007357
262	14250 r at	AL050300
263	14595 at	AL022580
264	14608 at	AC007357
	14614 at	AC004165
266	14621 at	AC004747
267	14627 i at	X76609
268	14628 r at	X76609
269	14635 s at	AC005398
270	14636 s at	AC013258
	14643 s at	AC006836
272	14672 s at	U18993
273	14675 s at	D85191
274	14691 at	AP002046
275	14704 s at	AC006067
276	14706 r at	AL137189
277	14709 at	AP002046
278	14711 s at	AF085279
279	14731 s at	AF014960
280	14784 at	AC005310
281	14951 at	AL049481
282	15057 at	AL035440
283	15085 s at	AL031018
284	15105_s_at	Z14987
285	15116 f at	AF121356
286	15125 f at	D85190
287	15141 s at	D85191
288	15145 s at	D64155
289	15154 s at	AL096860
290		U90522
	15178 s at	U43489
292	15216 s at	U75198
293	15431 at	AL030978
294	15496 at	AC006282
295	15523 s at	AL078637
296	15593 s at	U54561
297	15611 s at	L22567
298	15616 s at	AJ009696
299	15622 s at	U43945
277	13022 5_al	U43743

300	15629 s at	AB010407
301	15665 s at	AF022658
302	15680 s at	D42061
303	15846 at	AC006067
304	15847_g_at	AC006067
305	15866 s at	AC005770
306	15950 at	AC006429
307	15954 at	U72155
308	15978 at	X68592
309	16038 s at	L04173
310	16063 s at	AB008103
311	16105 s at	U68017
312	16150 s at	AL049730
313	16153 s at	AC013258
314	16393 s at	AC006436
315	16412 s at	AL022603
316	16442 s at	AJ002551
317	16504 s at	Z97335
318	16510 at	AL034567
319	16536 s at	AB008107
320	16539 s at	Z97343
321	16569 s at	L23968
322	16578 s at	AL137080
323	16609 s at	AB008104
324	16620 s at	AF051338
325	16637 s at	Z97336
326	16817 s at	AL096882
327	16864 i at	AL133248
328	16951 i at	AC005662
329	16952 s at	AC005662
330	16981_s at	U35829
331	17014_s_at	U05206
332	17033_s_at	U83179
333	17054 s at	AF134128
334	17073_s_at	AC006836
335	17119_s_at	AF132212
336	17123_s_at	AF106087
337	17128_s_at	AC005398
338	17187_at	AF128396
339; 669	17303_s_at	AC005499
340; 663	17379_at	AC018721
341	17386_at	AC006264
342	17413_s_at	X99097
343	17499_s_at	AF107726
344	17500_s_at	AL049862
345	17544_s_at	U40856
346	17567_at	AL162751
347	17886_at	AC004484

		•
348	17899 at	Z97339
349	17917 s at	AC004261
350	17961 at	AC007323
351	17963 at	AL049730
352	18003 at	AF188334
302	18064 r at	No hits found
	18069 at	No hits found
	18070 r at	No hits found
353	18216 at	AC012375
354	18210_at 18217_g_at	AC012375 AC012375
355	18235 at	•
	-	AC000348
356	18252_at	AL096882
357	18255_at	AC005770
358	18272_at	AF002109
359	18533_at	AL021684
360	18556_at	AC006264
361	18590_at	AJ222713
362	18591_at	X74756
363	18607_s_at	U78721
364	18635_at	AC004005
365	18716_at	AC007396
366	18876_at	AF002109
367	18920_at	AC002338
368	18928_at	AC002338
369	19034_at	AL021768
370	19171_at	AC002335
371	19178_at	AB035137
372	19182_at	AL031804
373	19251_at	AL035538
374	19640 at	AC004561
375	19977 at	AL049659
376	20017 at	AC004521
377	20034 i at	A71607
378	20201 at	AL078470
379	20227 s at	AB027252
380	20269 at	AC002387
381	20297 at	AC007153
382	20314 s at	AL096882
383	20335 s at	Y14208
384	20429 s at	Z97336
385	20585 s at	AC005309
386	20641 at	X91919
387	12333 at	AJ286345
388	14028 at	AJ20034J
389	14026_at	Z54356
390	14293_s_at	
		AC002329
391	15969_s_a	AJ133036
392	15982_s_a	X98190

		•
393	16461 I at	AC004683
394	16462_s_a	AC004683
395	16514 at	AL035538
396	17549_s_a	L37126
397	18706 s a	X75782
398	19594 i at	X98321
399	20555 s a	AL080318
		gb AAD17366.1  (AF128396) similar to
400	16010	human phosphotyrosyl phosphatase activator
400	16212_at	PTPA (GB:X73478) [Arabidopsis thaliana]
704	14005	gb AAC32233.2  (AC005168) unknown
794	14985_s_at	protein [Arabidopsis thaliana]
401	16411	emb CAB71046.1  (AL137898) shaggy-like
401	16411_s_at	kinase beta [Arabidopsis thaliana]
400	15000	gb AAD39561.1 AC007067_1 (AC007067)
402	15920_i_at	T10O24.1 [Arabidopsis thaliana]
400	1.000	emb CAA19705.1  (AL024486) putative
403	16299_at	protein [Arabidopsis thaliana]
	•	gb AAC98458.1  (AC005851) putative
404	18445 at	glucosyltransferase [Arabidopsis thaliana]
		gb AAC23400.1  (AC004005) putative
	• •	methyl chloride transferase [Arabidopsis
405	19215 at	thaliana]
		emb CAA50905.1  (X72022) ORF1
406	16439_at	[Arabidopsis thaliana]
		gb[AAD20710.1] (AC006300) unknown
407	16047_at	protein [Arabidopsis thaliana]
	<del>-</del>	gb AAF01328.1 AF188334_1 (AF188334)
	•	Toll/interleukin-1 receptor-like protein
408	18003 at	[Arabidopsis thaliana]
		emb CAB10219.1  (Z97336) hypothetical
409	20429_s_at	protei [Arabidopsis thaliana]
		gb AAD12002.1  (AC004261) calcium
		binding protein (CaBP-22) [Arabidopsis
410	17917_s_at	thaliana]
		emb CAB41717.1  (AL049730) pEARLI 1-
411	17963_at	like protein [Arabidopsis thaliana]
		emb CAB41718.1  (AL049730) pEARLI 1
412	16150_s_at	[Arabidopsis thaliana]
		emb CAA52619.1  (X74514) beta-
413	20239_g_at	fructofuranosidase [Arabidopsis thaliana]
		emb CAA52619.1  (X74514) beta-
414	20238_at	fructofuranosidase [Arabidopsis thaliana]
		omble A A 0.970.4 11 (A 10.00.000)11
415	15616 a of	emb CAA08794.1  (AJ009696) wall-
417	15616_s_at	associated kinase 1 [Arabidopsis thaliana]
416	18591 at	emb CAA52772.1  (X74756) ATAF2
410	10391_at	[Arabidopsis thaliana]

417	14116_at	gb AAC26243.1  (AF077407) contains similarity to sugar transporters (Pfam: sugar_tr.hmm, score: 395.39) [Arabidopsis thaliana]
		gb AAC72120.1  (AC005278) Strong similarity to gb D14550 extracellular dermal glycoprotein (EDGP) precursor from Daucus carota. ESTs gb 84105 and gb AI100071
418	12759_at	come from this gene. [Arabidopsis thaliana] gb AAC18809.1  (AC003671) Similar to high affinity potassium transporter, HAK1 protein gb U22945 from Schwanniomyces
419	19060_at	occidentalis. [Arabidopsis thaliana]
420	12998_at	emb CAB41863.1  (AL049746) aldose 1-epimerase-like protein [Arabidopsis thaliana]
	4.7.7.	gb AAD30608.1 AC007369_18 (AC007369)
421	13172_s_at	Sugar transporter [Arabidopsis thaliana]
422	14709_at	No hits found less than or equal to 1e-15.
423	15931 at	gb AAD41420.1 AC007727_9 (AC007727) F8K7.9 [Arabidopsis thaliana]
123	19991_41	emb CAB41109.1  (AL049656) ammonium
		transport protein (AMT1) [Arabidopsis
424	20369 s at	thaliana]
425	14201_at	No hits found less than or equal to 1e-15.
426	14691_at	No hits found.
		gb AAF34713.1 AF224762 1 (AF224762)
75	14148_at	SigA binding protein [Arabidopsis thaliana]
	•	emb CAB42872.1  (AJ012423) wall-
426	16140_s_at	associated kinase 2 [Arabidopsis thaliana]
		emb CAB42924.1  (AL049862) putative
427	13625 s at	disease resistance protein [Arabidopsis thaliana]
727	13023_s_at	emb CAA19683.1  (AL024486) putative
428	13702 s at	protein [Arabidopsis thaliana]
		gb AAC14530.1  (AC004484) unknown
429	17886_at	protein [Arabidopsis thaliana]
		gb AAC23641.1  (AC004684) putative
430	12354 g at	receptor-like protein kinase [Arabidopsis
750	12337_ <u>g_</u> ai	thaliana] gb AAC23641.1  (AC004684) putative
		receptor-like protein kinase [Arabidopsis
431	12353_at	thaliana]
,	<del>-</del> .	emb CAB10339.1  (Z97339) hypothetical
432	17899_at	protein [Arabidopsis thaliana]
		·

			emb CAB43665.1  (AL050352) Ca2+-
	433	18894 at	transporting ATPase-like protein
	433	10094_at	[Arabidopsis thaliana]
	434	14978_at	gb AAB64024.1  (AC002333) putative glucosyltransferase [Arabidopsis thaliana] emb CAA19683.1  (AL024486) putative
	435	14223_at	protein [Arabidopsis thaliana] gb[AAC05342.1] (AC002521) putative
	436	16109_s_at	protein kinase [Arabidopsis thaliana] gb AAD28055.1 AC007166_3 (AC007166) putative protein kinase [Arabidopsis
	437	18820_at	thaliana] gb AAC72865.1  (AF104919) similar to class I chitinases (Pfam: PF00182, E=1.2e-142,
	438	20345_at	N=1) [Arabidopsis thaliana] gb AAF29406.1 AC022354 5 (AC022354)
	439	14170_at	unknown protein [Arabidopsis thaliana] gb AAD38519.1 AF138281_1 (AF138281) phospholipase D-gamma-2 [Arabidopsis
	. 440	15143_s_at	thaliana] gb AAD19610.1  (AF107726) cyclic nucleotide gated channel [Arabidopsis
	441	17499_s_at	thaliana] emb CAB37511.1  (AL035540) Phospholipase like protein [Arabidopsis
	442	20590_at	thaliana]
		·	gb AAD31074.1 AC007357_23 (AC007357) Similar to gb AF038007 FIC1 gene from Homo sapiens and is a member of the PF 00122 E1-E2 ATPase family. ESTs gb T45045 and gb AA394473 come from this
	443	14608_at	gene. [Arabidopsis thaliana] emb[CAA18465.1] (AL022347)
-			serine/threonine kinase-like protein
	444	13550_at	[Arabidopsis thaliana]
	445	13355 at	emb CAA71371.1  (Y10342) putative
	443	15555_at	amidase [Arabidopsis thaliana] gb AAC78521.1  (AC005312) putative
	446	13564_at	glutathione S-transferase [Arabidopsis thaliana]
		<del>-</del>	emb CAB37510.1  (AL035540)
	447	19845 <u>g</u> at	monooxygenase 2 (MO2) [Arabidopsis thaliana] gb AAC32912.1  (AC004138) putative
	448	12764_f_at	glutathione S-transferase [Arabidopsis thaliana]
			gb AAB16855.1  (U71122) pyruvate
	449	17922_at	decarboxylase [Arabidopsis thaliana]

450 451	15982_s_at 12227_at	emb CAA66863.1  (X98190) peroxidase ATP2a [Arabidopsis thaliana] gb AAD39285.1 AC007576_8 (AC007576) Unknown protein [Arabidopsis thaliana] emb CAB45975.1  (AL080318) copper amine
452	20555_s_at	oxidase like protein (fragment2) [Arabidopsis thaliana] emb CAB37510.1  (AL035540) monooxygenase 2 (MO2) [Arabidopsis
453	19844_at	thaliana] gb AAC31242.1  (AC004747) unknown
454	13985_s_at	protein [Arabidopsis thaliana]
		gb AAD41421.1 AC007727_10 (AC007727) ESTs gb N96028, gb F14286, gb T20680, gb F14443, gb AA657300 and gb N65244
455	13548_at	come from this gene. [Arabidopsis thaliana] gb AAA32775.1  (L22567) cor78
456	15611_s_at	[Arabidopsis thaliana]
		gb AAD25759.1 AC007060_17 (AC007060) Strong similarity to F19I3.2 gi 3033375 putative berberine bridge enzyme from Arabidopsis thaliana BAC gb AC004238.
457	19840_s_at	EST gb R90518 comes from this gene. emb CAA67340.1  (X98808) peroxidase
458	15985_at	ATP3a [Arabidopsis thaliana] gb AAD32844.1 AC007658 3 (AC007658)
459	14584_at	unknown protein [Arabidopsis thaliana]
460	15422_at	gb AAD36948.1 AF069441_8 (AF069441) hypothetical protein [Arabidopsis thaliana] gb AAB61076.1  (AF007271) A TM021B04.14 gene product [Arabidopsis
461	20150_at	thaliana] gb AAC33239.1  (AC005315) putative
462	18844_at	ligand-gated ion channel protein [Arabidopsis thaliana] emb CAA20203.1  (AL031187) receptor-like serine/threonine protein kinase ARK3
463	16360_at	[Arabidopsis thaliana] gb AAB87113.1  (AC002391) putative
464	20292_at	protein kinase [Arabidopsis thaliana]
465	14554_at	gb AAC18798.1  (AC003671) F1707.4 [Arabidopsis thaliana] gb AAC19273.1  (AF069298) similar to several small proteins (~100 aa) that are
		induced by heat, auxin, ethylene and wounding such as Phaseolus aureus indole-3-
466	18604_at	acetic acid induced protein ARG (SW:32292) [Arabidopsis thaliana]

	•	gb AAB97145.1  (AF000977) MEK1
467	16061 s at	[Arabidopsis thaliana] thaliana]
	. — —	dbj BAA22813.1  (D26015) CND41,
		chloroplast nucleoid DNA binding protein
468	14145 at	[Nicotiana tabacum]
100	11115_ac	dbj BAA04870.1  (D21843) MAP kinase
469	16144 s at	
409	10144_S_at	[Arabidopsis thaliana]
470	15005	gb AAC49697.1  (U66345) calreticulin
470	17097_s_at	[Arabidopsis thaliana]
		gb AAB71447.1  (AC000098) Similar to
	•	Arabidopsis Fe(II) transport protein
471	19718_at	(gb U27590). [Arabidopsis thaliana]
	•	gb[AAC25507.1] (AC003979) T22J18.6
472	14298 g_at	[Arabidopsis thaliana]
		gb AAD12037.1  (AC002392) putative
		receptor-like protein kinase [Arabidopsis
473	12307 at	thaliana]
	12307_41	emb CAA18460.1  (AL022347) protein
474	20232 s at	
4/4	20232_8_at	kinase-like protein [Arabidopsis thaliana]
		gb AAF19738.1 AC012463 12 (AC012463)
475	19322 at	T2E6.14 [Arabidopsis thaliana]
7/3	19322_at	
		gb AAF07386.1 AC010675_9 (AC010675)
176	14004	putative peptide transporter [Arabidopsis
476	14224_at	thaliana]
	•	gb AAD39269.1 AC007203_1 (AC007203)
		Dutativa IIDD almanasatana
177	1.4270 -4	Putative UDP-glucose:sterol
477	14270_at	glucosyltransferase [Arabidopsis thaliana]
	<b>-</b>	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative
477 478	14270_at 15479_at	glucosyltransferase [Arabidopsis thaliana]
	<b>-</b>	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana]
478	_ 15479_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438)
	<b>-</b>	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana]
478 479	15479_at 14090_i_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1
478	_ 15479_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana]
478 479	15479_at 14090_i_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1
478 479 480	15479_at 14090_i_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana]
478 479	15479_at 14090_i_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to
478 479 480	15479_at 14090_i_at 16162_s_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana]
478 479 480	15479_at  14090_i_at  16162_s_at  20149_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana]  gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown
478 479 480 481	15479_at 14090_i_at 16162_s_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana]
478 479 480 481	15479_at  14090_i_at  16162_s_at  20149_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana]  gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative
478 479 480 481 482	15479_at  14090_i_at  16162_s_at  20149_at  12765_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana]  gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein
478 479 480 481	15479_at  14090_i_at  16162_s_at  20149_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana]
478 479 480 481 482	15479_at  14090_i_at  16162_s_at  20149_at  12765_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana]  gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana] gb AAC79588.1  (AC005727) putative RING
478 479 480 481 482 214	15479_at  14090_i_at  16162_s_at  20149_at  12765_at  12505_s_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana]  gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana] gb AAC79588.1  (AC005727) putative RING zinc finger ankyrin protein [Arabidopsis
478 479 480 481 482	15479_at  14090_i_at  16162_s_at  20149_at  12765_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana] gb AAC79588.1  (AC005727) putative RING zinc finger ankyrin protein [Arabidopsis thaliana]
478 479 480 481 482 214	15479_at  14090_i_at  16162_s_at  20149_at  12765_at  12505_s_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana] gb AAC79588.1  (AC005727) putative RING zinc finger ankyrin protein [Arabidopsis thaliana] gb AAC72124.1  (AC005278) ESTs
478 479 480 481 482 214	15479_at  14090_i_at  16162_s_at  20149_at  12765_at  12505_s_at  13140_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana]  gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana] gb AAC79588.1  (AC005727) putative RING zinc finger ankyrin protein [Arabidopsis thaliana] gb AAC72124.1  (AC005278) ESTs gb H37641 and gb AA651422 come from this
478 479 480 481 482 214	15479_at  14090_i_at  16162_s_at  20149_at  12765_at  12505_s_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1  (AL049483) putative protein [Arabidopsis thaliana] gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1  (U39944) BELL1 [Arabidopsis thaliana] gb AAB70415.1  (AC000106) Similar to Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1  (AC006340) unknown protein [Arabidopsis thaliana] gb AAC63643.1  (AC005309) putative CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana] gb AAC79588.1  (AC005727) putative RING zinc finger ankyrin protein [Arabidopsis thaliana] gb AAC72124.1  (AC005278) ESTs

		emb CAB41935.1  (AL049751) putative
485	16306_at	protein [Arabidopsis thaliana]
•	•	gb AAC26246.1  (AF077407) contains
		similarity to phosphoenolpyruvate synthase
		(ppsA) (GB:AE001056) [Arabidopsis
486	18611_at	thaliana]
	,	emb CAB43428.1  (AL050300) putative
487	20199_at	protein [Arabidopsis thaliana]
		emb CAA18626.1  (AL022580) putative
488	14595_at	protein [Arabidopsis thaliana]
		gb AAD31337.1 AC007354 10 (AC007354)
		Strong similarity to gb Y09533 involved in
		starch metabolism from Solanum tuberosum
		and contains a PF 01326 Pyruvate phosphate
		dikinase, PEP/pyruvate binding domain. EST
		gb N96757 comes from this gene.
489	12532_ at	[Arabidopsis thaliana]
105	12552_at	emb[CAB41162.1] (AL049659) putative
490	19977 at	protein [Arabidopsis thaliana]
170	19971_at	gb AAC79594.1  (AC005727) putative
		membrane channel protein [Arabidopsis
491	12773 at	thaliana
171	12115_at	gb AAD32870.1 AC005489 8 (AC005489)
492	19632 at	F14N23.8 [Arabidopsis thaliana]
772	19032_at	emb CAA16957.1  (AL021811) putative
493	19359 s at	protein [Arabidopsis thaliana]
,100	15555_5_41	emb[CAA78712.1] (Z14988) glycine rich
494	14716_f_at	protein [Arabidopsis thaliana] thaliana]
151	11710_1_41	gb AAC19269.1  (AF069298) T14P8.18 gene
495	13648 at	product [Arabidopsis thaliana]
155	150.10_at	gb AAD41977.1 AC006438_9 (AC006438)
496	12768_at	unknown protein [Arabidopsis thaliana]
.,,	12700_ut	gb AAC69134.1  (U78721) putative auxin-
497	16422_at	regulated protein [Arabidopsis thaliana]
12,1	10422_00	gb AAC26203.1  (AF053747) dormancy-
498	15131_s_at	associated protein [Arabidopsis thaliana]
170	13131_3_at	emb CAB38829.1  (AL035679) drought-
		inducible cysteine proteinase RD19A
499	14659 s at	precursor
.,,,	11005_0_4	emb CAB38829.1  (AL035679) drought-
		inducible cysteine proteinase RD19A
500	14658_s_at	precursor [Arabidopsis thaliana]
200	. 1000	emb CAB36513.1  (AL035440) putative
501	1'5057 at	protein [Arabidopsis thaliana]
		emb CAB56039.1  (AJ133786) gigantea
502	17581_g_at	protein [Arabidopsis thaliana]
		gb[AAC34217.1] (AC004411) putative
		alcohol dehydrogenase [Arabidopsis
503	18567 at	thaliana]
	·	

504 505	17047_s_at 15105_s_at	gb AAC68674.1  (AF078825) RING-H2 finger protein RHA3b [Arabidopsis thaliana] emb CAA78711.1  (Z14987) glycine rich protein [Arabidopsis thaliana]
506	14196_at	gb AAF16557.1 AC012563_10 (AC012563) unknown protein [Arabidopsis thaliana] gb AAD50976.1 AF170494_1 (AF170494) ionotropic glutamate receptor ortholog GLR6
507	17599_s_at	[Arabidopsis thaliana]
508	18556_at	gb AAD29803.1 AC006264_11 (AC006264) cyclophilin (CYP2) [Arabidopsis thaliana]
509	16486_at	gb AAB04606.1  (M81130) carboxypeptidase Y-like protein [Arabidopsis thaliana] gb AAB95293.1  (AF002109) unknown
510	18272_at	protein [Arabidopsis thaliana]
795	12219_at	gb AAD31373.1 AC006053_15 (AC006053) unknown protein [Arabidopsis thaliana] gb AAC80600.1  (AC005106) T25N20.21
511	20446_s_at	[Arabidopsis thaliana]
512	12561_at	emb CAA16701.1  (AL021687) putative protein [Arabidopsis thaliana]
		gb AAD46000.1 AC005916_12 (AC005916) Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs
513	12060_at	gb N95317, gb Z29139 and gb Z30853 come from this gene. [Arabidopsis thaliana] gb AAB61480.1  (AC000348) T7N9.4
514	18235_at	[Arabidopsis thaliana]
515	14021_r_at	emb CAA04134.1  (AJ000497) Starch branching enzyme II [Arabidopsis thaliana]
516	14020_i_at	emb CAA04134.1  (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA44630.1  (X62818)
517	16011_s_at	Metallothionein-like protein [Arabidopsis thaliana]
518	17033_s_at	gb AAD09232.1  (U83179) unknown [Arabidopsis thaliana]
519	16050_at	gb AAD24630.1 AC006919_10 (AC006919) putative fructose bisphosphate aldolase [Arabidopsis thaliana]

	520	19692_at	gb AAC14039.1  (AC003981) F22O13.13 [Arabidopsis thaliana] gb AAC39464.1  (AF053065) late
	521	19181_s_at	embryogenesis abundant protein homolog [Arabidopsis thaliana]
	792	13435_at	gb AAC49775.1  (AF003102) AP2 domain containing protein RAP2.9 [Arabidopsis thaliana]
	522	17128_s_at	gb AAC69381.1  (AC005398) pathogenesis- related PR-1-like protein [Arabidopsis gb AAC69381.1  (AC005398) pathogenesis-
	523	14635_s_at	related PR-1-like protein [Arabidopsis thaliana] gb AAD15461.1  (AC006067) unknown
	524	15846_at	protein [Arabidopsis thaliana] gb[AAD15461.1] (AC006067) unknown
	525	15847_g_at	protein [Arabidopsis thaliana] gb AAD15461.1  (AC006067) unknown
	526	14704_s_at	protein [Arabidopsis thaliana]
•	527	17456_at	gb AAB80678.1  (AC002332) putative steroid dehydrogenase [Arabidopsis thaliana]
	528	14895_s_at	emb CAB10562.1  (Z97344) acetylornithine deacetylase [Arabidopsis thaliana] gb AAB95235.1  (AC002130) F1N21.7
	529	19979_at	[Arabidopsis thaliana]
	530	20325_s_at	emb CAA78152.1  (Z12162) protein phosphatase 1A [Arabidopsis thaliana] gb AAB61479.1  (AC000348) T7N9.3
	531	18234_at	[Arabidopsis thaliana] emb CAA35838.1  (X51474) kin1
	532	16474_s_at	[Arabidopsis thaliana] emb CAA38894.1  (X55053) cold regulated
	533	18701_s_at	[Arabidopsis thaliana] gb AAD23000.1 AC007087_19 (AC007087)
	534	13785_at	cold-regulated protein cor15b precursor [Arabidopsis thaliana] gb AAC23422.1  (AC004005) putative
	535	20387_at	methionine aminopeptidase [Arabidopsis thaliana]
	536	13178_at	gb AAB63086.1  (U93215) unknown protein [Arabidopsis thaliana]
	537	12103_at	gb AAD30603.1 AC007369_13 (AC007369) Unknown protein [Arabidopsis thaliana]
	538	13225_s_at	emb CAA42483.1  (X59814) Cold and ABA regulated gene [Arabidopsis thaliana]

		gb AAB95275.1  (AF002109) putative LIM-
539	17003_at	domain protein [Arabidopsis thaliana]
	. —	emb CAA19880.1  (AL031032) putative
540	15878_at	protein [Arabidopsis thaliana]
		gb AAD03574.1  (AC003952) putative
		senescence-related protein [Arabidopsis
541	13004_at	thaliana]
		gb AAC34333.1  (AC004122) Highly Similar
		to branched-chain amino acid
542	14052_at	aminotransferase [Arabidopsis thaliana]
		gb AAC05351.1  (AC002521) putative
		receptor-like protein kinase [Arabidopsis
543	15798_at	thaliana]
	,	gb AAB67985.1  (L36246) anoxia-induced
793	12345_at	protein [Arabidopsis thaliana]
	•	emb CAA20206.1  (AL031187)
		-serine/threonine kinase-like protein
544	16818_s_at	[Arabidopsis
a 4 =	4004	gb AAC62136.1  (AC005169) unknown
545	13916_at	protein [Arabidopsis thaliana]
•		emb CAB41311.1  (AL049711) putative heat
	000.40	shock transcription factor [Arabidopsis
666	20342_at	thaliana]
		emb CAB10242.1  (Z97336) germin
546	20421 -4	precursor oxalate oxidase [Arabidopsis
. 340	20421_at	thaliana]
547	14250 r at	emb CAB43438.1  (AL050300) putative
J+1	14230_1_at	protein [Arabidopsis thaliana]
548	14249 i at	emb CAB43438.1  (AL050300) putative protein [Arabidopsis thaliana]
340	17277_1_al	gb AAC49282.1  (U40856) AIG1
549	17544 s at	[Arabidopsis thaliana]
347	11544_5_at	gb AAC49282.1  (U40856) AIG1
550	12879 s at	[Arabidopsis thaliana]
	.20/7_5_at	gb AAC16079.1  (AC004521) unknown
551	20017 at	protein [Arabidopsis thaliana]
		emb CAB40989.1  (AL049640) growth factor
552	13177 at	like protein [Arabidopsis thaliana]
		gb AAF18611.1 AC005170_1 (AC005170)
		similar to senescence-associated protein
553	19946 at	[Arabidopsis thaliana]
	_	gb AAD08938.1  (AC005724) unknown
554	17894 at	protein [Arabidopsis thaliana]
	<u></u>	gb AAD15572.1  (AC006340) unknown
·555	15855_at	protein [Arabidopsis thaliana]
		emb CAB45807.1  (AL080253) putative
556	15558_r_at	protein [Arabidopsis thaliana]
557	15208_s_at	No hits found less than or equal to 1e-15.
	·· =	•

,	558	16153 s at	gb AAF21072.1 AC013258_10 (AC013258) thaumatin-like protein [Arabidopsis thaliana]
	<i>33</i> 6	10133_s_at	
	559	14636_s_at	gb AAF21072.1 AC013258_10 (AC013258) thaumatin-like protein [Arabidopsis thaliana] dbj BAA86999.1  (AB035137) blue copper
	560	19178_at	binding protein [Arabidopsis thaliana] emb CAB56039.1  (AJ133786) gigantea
·	561	17580_at	protein [Arabidopsis thaliana]
			gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to gb X97864 cytochrome P450 from Arabidopsis thaliana and is a member of the PF 00067 Cytochrome P450 family. ESTs gb N65665, gb T14112, gb T76255, gb T20906 and gb AI100027
	562	14248_at	come from this gene. emb CAA72484.1  (Y11788) peroxidase
•	563	18946_at	ATP24a [Arabidopsis thaliana] emb CAA17138.1  (AL021889) putative
	564	13009_i_at	protein [Arabidopsis thaliana] gb AAA33709.1  (L16797) glutamate
	565	18508_s_at	decarboxylase [Petunia x hybrida] emb[CAB45330.1] (AL079344) nucleotide pyrophosphatase-like protein [Arabidopsis
	566	12556_at	thaliana]
			gb AAB60774.1  (AC000375) ESTs gb U75592,gb T13956,gb T43869 come from
	567	13115_at	from this gene. [Arabidopsis thaliana] gb AAB87120.1  (AC003000) unknown
	568	15046_s_at	protein [Arabidopsis thaliana] gb AAC67339.2  (AC005499) putative
	339	17303_s_at	WRKY-type DNA binding protein [Arabidopsis thaliana] emb CAB45881.1  (AL080282) berberine bridge enzyme-like protein [Arabidopsis
	569	18597_at	thaliana] emb CAB42588.1  (A71590) unnamed
	570	13908_s_at	protein product [Arabidopsis thaliana]. emb CAB41103.1  (AL049655) putative
	571	14553_at	protein [Arabidopsis thaliana] gb AAB64044.1  (AC002333) putative
	572	18928_at	endochitinase [Arabidopsis thaliana]

		gb AAC72119.1  (AC005278) Strong similarity to gb D14550 extracellular dermal glycoprotein (EDGP) precursor from Daucus carota. ESTs gb H37281, gb T44167, gb T21813, gb N38437, gb Z26470, gb R65072, gb N76373, gb F15470,
573	12772_at	gb Z35182, gb H76373, gb Z34678 and gb Z35387> emb CAA18124.1  (AL022141) putative
574	16326_at	receptor protein kinase [Arabidopsis thaliana]
575	20479_i_at	emb CAB38908.1  (AL035708) cytochrome P450-like protein [Arabidopsis thaliana] gb AAD28318.1 AC006436_9 (AC006436) putative receptor-like protein kinase
576	16393_s_at	[Arabidopsis thaliana] emb CAA67551.1  (X99097) peroxidase
- 577	17413_s_at	[Arabidopsis thaliana] gb[AAA32835.1] (M96073)
578	14620_s_at	phosphoribosylanthranilate transferase [Arabidopsis thaliana]
579	20480_s_at	emb CAB38908.1  (AL035708) cytochrome P450-like protein [Arabidopsis thaliana] gb AAC79625.1  (AC005770) unknown
580	15866_s_at	protein [Arabidopsis thaliana] emb CAA21214.1  (AL031804) putative
581	19182_at	protein [Arabidopsis thaliana] gb AAC79625.1  (AC005770) unknown
582	18255_at	protein [Arabidopsis thaliana] emb CAA74639.1  (Y14251) glutathione S-
583	16054_s_at	transferase [Arabidopsis thaliana]
584	14672_s_at	gb AAC49117.1  (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
585	20291_s_at	gb AAA32738.1  (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana]
586	16053_i_at	emb CAA74639.1  (Y14251) glutathione Stransferase [Arabidopsis thaliana]
587	17083_s_at	gb AAD25838.1 AC006951_17 (AC006951) putative indole-3-glycerol phosphate synthase [Arabidopsis thaliana]
588	12889_s_at	gb AAA32738.1  (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana]
589	12642_at	gb AAD22285.1 AC006920_9 (AC006920) unknown protein [Arabidopsis thaliana]
		•

590	17487_s_at	gb AAC49117.1  (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana] gb AAA32835.1  (M96073) phosphoribosylanthranilate transferase
591	14838_s_at	[Arabidopsis thaliana]
592	17104_s_at	dbj BAA13640.1  (D88541) phosphoserine aminotransferase [Arabidopsis thaliana] gb AAC79626.1  (AC005770) putative
593	19892_at	protease inhibitor [Arabidopsis thaliana]
504	15105	gb AAC31756.1  (U68017) heat shock
594	16105_s_at	transcription factor 4 [Arabidopsis thaliana]
		gb AAC31756.1  (U68017) heat shock
664	13273_s_at	transcription factor 4 [Arabidopsis thaliana]
595	12892 g at	gb AAC63850.1  (U73786) ACC synthase [Arabidopsis thaliana]
393	12092_g_at	dbj BAA22096.1  (D85191) vegetative
596	15141 s at	storage protein [Arabidopsis thaliana]
	131-11_3_41	emb CAA55322.1  (X78585) Di21
597	18231 at	[Arabidopsis thaliana]
	10251_41	dbj BAA24440.1  (AB010407)
		phosphoglycerate dehydrogenase
598	15629_s_at	[Arabidopsis thaliana]
		emb CAA48579.1  (X68592) adenosine
		nucleotide translocator [Arabidopsis
599	15978 at	thaliana]
	<b></b> .	gb AAB82640.1  (AC002387) putative
600	20269_at	pectinesterase [Arabidopsis thaliana]
	* * * * * * * * * * * * * * * * * * *	gb AAC16958.1  (AC004165) putative
601	14614_at	glucosyltransferase [Arabidopsis thaliana]
		emb CAA07352.1  (AJ006960) peroxidase
602	17930_s_at	[Arabidopsis thaliana]
		gb AAC78532.1  (AC005662) calmodulin-
603	16952_s_at	like protein [Arabidopsis thaliana]
		gb AAC49679.1  (U77347) lethal leaf-spot 1
604	12930_s_at	homolog [Arabidopsis thaliana]
	12842_s_at	No hits found less than or equal to 1e-15.
	•	dbj BAA32418.1  (AB008103) ethylene
605	1.000	responsive element binding factor 1
605	16063_s_at	[Arabidopsis thaliana]
		dbj BAA32418.1  (AB008103) ethylene
220	12004 a at	responsive element binding factor 1
228	12904_s_at	[Arabidopsis thaliana]
		dbj BAA32422.1  (AB008107) ethylene
606	12908 s at	responsive element binding factor 5 [Arabidopsis thaliana]
	12700_3_at	[Arabidopsis dialialia]

	•	emb CAA17127.1  (AL021889) hypothetical
607	15937_at	protein [Arabidopsis thaliana]
	_	gb AAB87109.1  (AC002391) putative
608	17843 s at	cytochrome P450 [Arabidopsis thaliana]
		gb AAC95196.1  (AC004561) putative
		glutathione S-transferase [Arabidopsis
609	18966 at	thaliana
	_	gb AAC80599.1  (AC005106) T25N20.20
610	20519_at	[Arabidopsis thaliana]
	_	gb AAC95189.1  (AC004561) putative
		glutathione S-transferase [Arabidopsis
611	19641_at	thaliana]
	sir	gb AAD32297.1 AC006533_21 (AC006533)
		putative glucosyltransferase [Arabidopsis
612	17408_at	thaliana]
		gb AAC37474.1  (L42212) serine
613	15646_s_at	acetyltransferase [Arabidopsis thaliana]
		gb AAC49988.1  (AF014960) multidrug
		resistance-associated protein 2; AtMRP2
614	14731_s_at	[Arabidopsis thaliana]
		emb CAB41928.1  (AL049751) short-chain
		alcohol dehydrogenase like protein
615	20685_at	[Arabidopsis thaliana]
		emb CAA17559.1  (AL021961)
		glucosyltransferase -like protein [Arabidopsis
616	16968_at	thaliana]
		gb AAC78440.1  (U92460) 12-
<b>615</b>	10050	oxophytodienoate reductase OPR1
617	18253_s_at	[Arabidopsis thaliana] thaliana]
	•	gb AAD20156.1  (AC006282) putative
618	15496 at	glucosyl transferase [Arabidopsis thaliana]
010	13.130_at	emb CAA52771.1  (X74755) ATAF1
619	19137 at	[Arabidopsis thaliana]
	13131_dt	emb CAA18722.1  (AL022603) putative
		NADPH quinone oxidoreductase
620	19132 s at	[Arabidopsis thaliana]
		gb AAD22649.1 AC007138_13 (AC007138)
		predicted protein of unknown function
621	13656 at	[Arabidopsis thaliana]
		gb AAB67854.1  (U61231) cytochrome P450
622	17024_s at	[Arabidopsis thaliana]
		emb CAB69849.1  (AL137189) anthranilate
		N-benzoyltransferase-like protein
623	14705_i_at	[Arabidopsis thaliana]
	<del></del>	
	1550	emb CAB42906.1  (AL049862) calmodulin-
624	17500_s_at	like protein [Arabidopsis thaliana]
		·

625	13217_s_at	emb CAB42906.1  (AL049862) calmodulin- like protein [Arabidopsis thaliana] gb AAC49573.1  (U43412) 3'-
626	15196_s_at	phosphoadenosine 5'-phosphosulfate reductase [Arabidopsis thaliana] emb CAA10955.1  (AJ222713) unnamed
627	18590 at	protein product [Arabidopsis thaliana]
628		
020	14700_at	No hits found less than or equal to 1e-15.
		emb CAA69879.1  (Y08568) trehalose-6-
629	14665_r at	phosphate synthase [Arabidopsis thaliana]
023	1,005_1_40	phospiano symmaso [i masicopolis chamana]
•		gb AAF18681.1 AF024504 11 (AF024504)
630	12630_at	unknown protein [Arabidopsis thaliana]
	· ·	gb AAC69851.1  (AF077955) branched-
		chain alpha keto-acid dehydrogenase E1
631	18953_at	alpha subunit [Arabidopsis thaliana]
	-	emb CAA16793.1  (AL021713) putative
632	13514 s at	protein [Arabidopsis thaliana]
		gb AAF02787.1 AF195115 7 (AF195115)
		weak similarity to receptor protein kinase
633	12490 at	[Arabidopsis thaliana]
	, , ,	emb CAB10404.1  (Z97340) phytochrome D
634	12246 s at	[Arabidopsis thaliana]
05.	122 10_0_a	emb CAB37488.1  (AL035539) putative
635	20536_s_at	protein [Arabidopsis thaliana]
000	20000_5_4.	gb AAC72122.1  (AC005278) F15K9.14
636	18409_at	[Arabidopsis thaliana]
050	10105_ut	gb AAD21475.1  (AC007017) unknown
637	19387 at	protein [Arabidopsis thaliana]
057	19507_at	gb AAB70244.1  (AF016848) WD-40 repeat
638	16117_s_at	protein [Arabidopsis thaliana]
050	10117_3_4	emb CAA21480.1  (AL031986) putative
639	18347 s at	protein [Arabidopsis thaliana]
033	105 17_5_41	emb CAB38906.1  (AL035708) putative
640	15880 at	protein [Arabidopsis thaliana]
0.10	15000_at	gb AAC49767.1  (AF003094) AP2 domain
		containing protein RAP2.1 [Arabidopsis
667	20471 at	thaliana]
007	20471_at	trantalaj
		gb AAB58497.1  (U81293) UDP-
		glucose:indole-3-acetate beta-D-
641	16603_s_at	glucosyltransferase [Arabidopsis thaliana]
	- <del>-</del>	gb AAC13598.1  (AF058914) F21E10.13
642	12049_at	gene product [Arabidopsis thaliana]
		gb AAC78704.1  (AF001308) predicted
643	12048_at	glycosyl transferase [Arabidopsis thaliana]
		•

		gb AAB61117.1  (AC002062) ESTs
644	14064 at	gb N38288,gb T43486,gb AA395242 come from this gene. [Arabidopsis thaliana]
044	14004_at	gb[AAC04492.1] (AC003974) unknown
645	12149_at	protein [Arabidopsis thaliana]
0.5	12117	emb CAA91183.1  (Z54356) HD-ZIP
646	14295 s at	[Arabidopsis thaliana]
0.0	1 1255_5_ac	emb CAA16930.1  (AL021768) TMV
		resistance protein N-like [Arabidopsis
647	19034 at	thaliana]
		gb AAC27848.1  (AC004218) unknown
648	18624 at	protein [Arabidopsis thaliana]
	_	gb AAC97218.1  (AC005936) unknown
649	13181_at	protein [Arabidopsis thaliana]
		gb AAD10163.1  (AC005917) putative Tal1-
•		like non-LTR retroelement protein
650	18866_at	[Arabidopsis thaliana]
		emb CAB44686.1  (AL078620) cytochrome
651	19502_at	P450-like protein [Arabidopsis thaliana]
		emb CAA19807.1  (AL031018) hypothetical
652	16301_s_at	protein [Arabidopsis thaliana]
		gb AAD32774.1 AC007661 11 (AC007661)
653	19411_at	unknown protein [Arabidopsis thaliana]
	. ~	emb CAA71588.1  (Y10556) CONSTANS
654	20300_g_at	[Arabidopsis thaliana]
		emb CAA71588.1  (Y10556) CONSTANS
655	20299_at	[Arabidopsis thaliana]
	· · · · · · · · · · · · · · · · · · ·	gb AAB57688.1  (U96045) APS reductase
656	18696_s_at	[Arabidopsis thaliana]
		gb AAC26980.1  (AF016283) 5'-
		adenylylsulfate reductase [Arabidopsis
657	15186_s_at	thaliana] [Arabidopsis thaliana]
659	16609_s_at	AtERF2 (, BAA32419.1; AB008104)
660	10000 - 4	EREBP4-like, AtERF6 (, CAB10530.1;
660	12909_s_at	Z97343)
661	16536_s_at	AtERF5 (, BAA32422.1; AB008107)
201	15/65	put. C2H2 zinc finger transcription factor (,
301	15665_s_at	AAB80922.1; AF022658)
668	12176 of	Myb-like (, emb CAA20567.1  (AL031394)
670	13176_at 15778 at	putative protein
671	20619 at	X98676.2_at AC005896.161 at
672	-	<b>—</b>
673	20335 s at	AL023094.197_s_at Y14208.2 s at
674	18949 at	Z54136.1_at
675	13015_s_at	<del>-</del>
013	13012_s_at	X98673.2 s at emb CAB41311.1

		(AL049711) putative heat shock transcription
		factor [Arabidopsis thaliana]
676	19646_s_at	AC005819.55 s at
677	19855_at	AC007260.16_at .
		AC007047.101_at-gb AAC49767.1
	•	(AF003094) AP2 domain containing protein
. 678	18475_at	RAP2.1 [Arabidopsis thaliana]
		unspecified t-factor gb AAC49775.1
		(AF003102) AP2 domain containing protein
679	13001_at	RAP2.9 [Arabidopsis thaliana]
		AC004665.101_at emb CAA67234.1
		(X98676) zinc finger protein [Arabidopsis
680	15219_at	thaliana]
		ATTHIRED4_s_at gb AAC98070.1
		(AC005896) putative C2H2-type zinc finger
681	. 13189_s_at	protein [Arabidopsis thaliana]
•		hsp70_s_at emb CAA18838.1  (AL023094)
		bZIP transcription factor ATB2 [Arabidopsis
684	13284_s_at	thaliana]

Table 10

	<u>Set</u>	Content	Number of unique genes
	2	7 or 8 not edm1 not	55 genes
5		rps2 not pad4 or NahG	
	3	edm1 not rps2 not pad4 or NahG	44 genes
	4	rps2 not edm1 not pad4 or NahG	12 genes
	5	pad4 or NahG not edm1 not rps2	20 genes
	6	edm1 and rps2 not pad4 or NahG	17 genes
10	7	edm1 and pad4 or NahG not rps2	18 genes
	8.	rps2 and pad4 or NahG not edm1	7 genes
	9	edm1 and rps2 and pad4 or NahG	11 genes
		Total	184 genes

15 For engineering resistance to pathogens whose growth is restricted by RPP7 or RPP8 dependent responses, such as oomycete pathogens, all 184 genes are potentially useful. The 128 genes in sets 2, 3, 4, and 6 are more likely to be useful, as pad4 and NahG do not interfere with resistance mediated by RPP7 or RPP8, so genes under their control should not be important for this type of resistance. The 99 genes in sets 2 and 3 are even more likely to be useful, because they not affected by rps2, and RPP7 and RPP8 trigger a different kind of resistance response than the one triggered by RPS2. The 44 genes in set 3 are

most likely to be useful, as they require *EDM1*, and *EDM1* is required for resistance.

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# Example 3

Transcriptional Responses Triggered by the RPP7 Defense Signaling Pathway

The RPP7 defense-signaling pathway (Figure 1) mediates resistance of
the Arabidopsis ecotype Col-0 against the Peronospora isolate Hiks1. In
contrast to conventional R-gene dependent defense signaling pathways in
Arabidopsis, such as the RPP4 pathway, the RPP7 pathway does not essentially
require salicylic acid accumulation or previously described defense regulators,
such as EDS1, NDR1, NPR1, PAD4 and others.

A comparative analysis of transcriptional responses triggered by the RPP7 and RPP8 pathways was performed by gene expression profiling using Affymetrix oligonucleotide chips with roughly 8,200 different Arabidopsis genes, representing roughly a third of the genome. For the analysis, 2 week old seedlings were harvested 0, 12 or 48 hours post infection with a particular Peronospora isolate (Table 11). To examine RPP8 triggered transcriptional responses, a transgenic line Col-0 line carrying the RPP8 resistance gene from the Arabidopsis ecotype Landsberg erecta was infected with the Peronospora isolate Emco5. This interaction is incompatible. The interaction between Emco5 and Col-0 wild type plants served as compatible control. Responses triggered by the RPP7 pathway were analyzed using the incompatible interaction between the *Peronospora* isolate Hiks1 and Col-0 wild type plants, as well as compatible interactions between Hiks1 and the susceptible mutants rpp7, edm1, edm2 and edm3. To compare transcriptional responses triggered by the unconventional RPP7 and RPP8 pathways with those triggered by the more conventional RPP4 pathway, Col-0 wild type plants were infected with the Peronospora isolate Emoy2, which is avirulent on this ecotype. The interactions between Emoy2 and NahG plants, as well as the mutants pad4, ndr1 and npr1, served as compatible controls. Whereas pad4 and NahG plants are fully Emoy2 susceptible, ndr1 and npr1 plants are only partly compromised in Emoy2 resistance.

	Table 11 Plant	Peronospora isolate	Interaction type
	x rant	1 eronospora isolate	interaction type
	Col-0 (tg <i>RPP8</i> )	Emco5	incompatible ·
	Col-0 (rpp8)	Emco5	compatible
5	Col-0 (RPP7)	Hiks1	incompatible (2x)
	Col-0 (rpp7; 3929)	Hiks1	compatible
	Col-0 (edm1)	Hiks1	compatible
	Col-0 (edm2)	Hiksl	compatible
	Col-0 (edm3)	Hiksl	compatible
10	Col-0 (RPP4)	Emoy2	incompatible
	Col-0 (ndr-1-1)	Emoy2	intermediate
	Col-0 (pad4-1)	Emoy2	compatible
	Col-0 (NahG)	Emoy2	compatible
•	Col-0 (npr1-1)	Emoy2	intermediate
			•

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## Results

# <u>Identification of Genes Potentially Required for RPP7 Mediated Peronospora</u> Resistance

Genes that play a role in the establishment of resistance may show 20 differences in their regulation or their expression levels between compatible and incompatible interactions (e.g., genes may show lower expression levels in "loss of Hiks-resistance mutants" as compared to Col-0 wild type plants). Thus, for each gene expression, ratios were calculated between Col-0 and each of the four Hiks response mutants (rpp7, edm1, edm2 or edm3) at each time point (0, 12 and 48 hours). The resulting data set, consisting of 12 expression ratios for each of the 8,775 probe sets on the chip, was first analyzed with "CLUSTER" and "TREE VIEW", two commonly used programs for chip and micro-array data analysis (Eisen et al., 1998). Genes that demonstrated only minor or less significant expression differences between Col-0 wild type and the mutants were 30 excluded from analysis. Genes that showed at least one 3-fold expression difference over all 12 calculated expression ratios were first considered for further analysis.

Table 12 shows 194 probe sets corresponding to genes having SEQ ID NOs: 301-494, the expression of which is altered after infection of *Arabidopsis* with *P. parasitica* (wild-type relative to *rpp7*, *edm1*, *edm2*, or *edm3*). Table 13

depicts about 100 genes (genes comprising SEQ ID NOs: 373, 385, 242, 369, 306, 232, 346, 367, 212, 221, 307, 322, 240, 313, 270, 252, 383, 245, 377, 212, 327, 297, 375, 263, 250, 282, 358, 257, 332, 560, 284, 220, 259, 355, 248, 281, 215, 236, 309, 386, 253, 333, 336, 296, 319, 414, 354, 353, 293, 278, 324, 339, 345, 225, 249, 264, 310, 228, 214, 311, 343, 298, 384, 277, 314, 372, 331, 338, 370, 352, 365, 261, 374, 238, 344, 241, 380, 371, 262, 258, 256, 304, 275, 303, 274, 254, 223, 348, 318, 251, 291, 289, 361, 285, 368, 342, 290, 351, 312, 286, 287, 273, 239, 326, 226, 227, 350, 280, 317, 359, 294, 279, 382, 356, 234, 218, 217, 347, 379, 401, 378, 363, 341, 208, 268, 267, 364, 266, 337, 269, 269, 325, 793 and 224) represented by 137 probe sets that show at least one 3-fold-10 expression difference in comparisons between wild type Arabidopsis Col-0 and mutants rrp7 or edm1, edm2, or edm3 infected with P. parasitica Hiks1 (i.e., probe sets that show at least one 3-fold difference in expression levels over all 12 expression ratios calculated between Col-0 wild type and each of the 4 Hiks 15 response mutants at each of the time points). Genes were grouped together according to similarities of their expression characteristics. Expression ratios were color encoded. Positive expression ratios were red and negative ratios were green. Black indicated no expression difference. Genes were represented in rows and infection treatments in columns. Red generally indicated that a gene at 20 a given time point is more strongly expressed in Col-0 wild type plants as compared to the respective mutant. One cluster represented genes that are upregulated in response to Hiks1 infection by the RPP7 pathway (Table 14; genes comprising SEQ ID NOs: 365, 261, 374, 238, 344, 241, 380, 371, 262, 258, 256, 304, 275, 303, 274, 254, 310, 228, 214, 347 and 225, which are 25 induced by Hiks1-infection and which are RPP7, EDM1, EDM2, EDM3dependent genes and so may encode regulators acting downstream of those genes; see probe sets 12505, 13217 and 12904 which correspond to genes that encode potential regulators of Hiks response pathway). Genes in this cluster represent genes that are more strongly expressed in Col-0 wild type than in all 30 four mutants predominantly at the 12 hour time-point in each comparison. The expression difference is less pronounced in the case of edm2 and edm3, but it is consistent over all four comparisons.

Table 12

Affy ID (Probe Set)	Reference No
12324_i_at	AC007212
12333_at	AJ286345
12345_at	L36246
12500_s_at	AF081067
12505 s at	AC005309
12608 i at	S70188
12642 at	AC006920
12746 i at	AL096882
12748 f at	AL096882
12761_s_at	AC006577
12773_at	AC005727
12778_r_at	AC006577
12798_at	AC003028
12802_at	AL022373
12842_s_at	
12843_s_at	AC007195
12845_s_at	AJ004810
12879_s_at	U40856
12891_at	U73786
12892_g_at	U73786
12904_s_at	AB008103
12905_s_at	AB008104
12908_s_at	AB008107
12909_s_at	Z97343
12911_s_at	X84229
12916_s_at	AF021244
13138_at	AL096882
13177_at	AL049640
13178_at	U93215
13187_i_at	U35829
13189_s_at	U35829
13198_i_at	AL021749
13212_s_at	AL137080
13217_s_at	AL049862
13258_s_at	AC005309
13273_s_at	U68017
13284_s_at	AJ002551
13604_at	AC000104
13615_at	AC002332
13617_at	AC006592
13718_at	Z72152
13771_at	AC006593
13785_at	AC007087
14028_at	AF075597
14052_at	AC004122
14096_at	AC002291
14097_at	AC005309

14116_at	AF077407
14141_at	AC011437
14148_at	AF224762
14196_at	AC012563
14201_at	AL163972
14219_at	AC068667
14223_at	AL024486
14248_at	AC007357
14250 r at	AL050300
14295 s at	Z54356
14595 at	AL022580
14608 at	AC007357
14614 at `	AC004165
14621 at	AC004747
14627 i at	X76609
14628 r at	X76609
14635 s at	AC005398
14636 s at	AC013258
14643 s at	AC006836
14672 s at	U18993
14675 s at	D85191
14691 at	AP002046
14704 s at	AC006067
14704_s_at	AL137189
14709_1_at	AP002046
14711 s at	AF085279
14711_s_at	AF014960
14784 at	AC005310
14951 at	AL049481
14965 at	AC002329
15057 at	AL035440
-	AL033440 AL031018
	Z14987
15105_s_at	
15116_f_at	AF121356
15125_f_at	D85190
15141_s_at	D85191
15145_s_at	D64155
15154_s_at	AL096860
15161_s_at	U90522
15178_s_at	U43489
15216_s_at	U75198
15431_at	AL030978
15496_at	AC006282
15523_s_at	AL078637
15593_s_at	U54561
15611_s_at	L22567
15616_s_at	AJ009696
15622_s_at	U43945

15629_s_at	AB010407
15665 s at	AF022658
15680 s at	D42061
15846 at	AC006067
15847_g_at	AC006067
15866 s at	AC005770
15950 at	AC006429
15954 at	U72155
15969 s a	AJ133036
15978 at	X68592
15982 s a	X98190
16038 s at	L04173
16063 s at	AB008103
16105 s at	U68017
16150 s at	AL049730
16153 s at	AC013258
16393 s at	AC006436
16412 s at	AL022603
16442 s at	AJ002551
16461 I at	AC004683
16462 s a	AC004683
16504 s at	Z97335
16510 at	AL034567
16514 at	AL035538
16536 s at	AB008107
16539 s at	Z97343
16569 s at	L23968
16578 s at	AL137080
16609 s at	AB008104
16620_s_at	AF051338
16637_s_at	Z97336
16817_s_at	AL096882
16864_i_at	AL133248
16951_i_at	AC005662
16952_s_at	AC005662
16981_s_at	U35829
17014_s_at	U05206
17033_s_at	U83179
17054_s_at	AF134128
17073_s_at	AC006836
17119_s_at	AF132212
17123_s_at	AF106087
17128_s_at	AC005398
17187_at	AF128396
17303_s_at	AC005499
17379_at	AC018721
17386_at	AC006264
17413_s_at	X99097

17499_s_at	AF107726
17500_s_at	AL049862
17544_s_at	U40856
17549_s_a	L37126
17567_at	AL162751
17886 at	AC004484
17899 at	Z97339
17917 s at	AC004261
17961 at	AC007323
17963 at	AL049730
18003 at	AF188334
18064_r_at	
18069 at	
18070 r at	
18216 at	AC012375
18217_g_at	AC012375
18235 at	AC000348
18252 at	AL096882
18255 at	AC005770
18272 at	AF002109
18533 at	AL021684
18556 at	AC006264
18590 at	AJ222713
18591 at	X74756
18607 s at	U78721
18635 at	AC004005
18706 s a	X75782
18716 at	AC007396
18876 at	AF002109
18920 at	AC002338
18928 at	AC002338
19034 at	AL021768
19171 at	AC002335
19178 at	AB035137
19182 at	AL031804
19251 at	AL035538
19594 i at	X98321
19640 at	AC004561
19977 at	AL049659
20017 at	AC004521
20034 i at	A71607
20201 at	AL078470
20227 s at	AB027252
20269 at	AC002387
20297 at	AC007153
20314 s at	AL096882
20335 s at	Y14208
20429 s at	Z97336
	. 271000

20555_s_a	AL080318
20585_s_at	AC005309
20641_at	X91919

# Table 13

Probe Set	Description
12324_i_at	AC007212
12345_at	L36246
12505_s_at	AC005309
12608_i_at	S70188
12746_i_at	AL096882
12748_f_at	AL096882
12761_s_at	AC006577
12773_at	AC005727
12778_r_at	AC006577
12798_at	AC003028
12802_at	AL022373
12842_s_at	
12843_s_at	AC007195
12845_s_at	AJ004810
12879_s_at	U40856
12891_at	U73786
12892_g_at	U73786
12904_s_at	AB008103
12908_s_at	AB008107
12911_s_at	X84229
13138_at	AL096882
13178_at	U93215
13189_s_at	U35829
13198_i_at	AL021749
13212_s_at	AL137080
13217_s_at	AL049862
13258_s_at	AC005309
13604_at	AC000104
13718_at	Z72152
13771_at	AC006593
13785_at	AC007087
14052_at	AC004122
14096_at	AC002291
14097_at	AC005309
14116_at	AF077407
14148_at	AF224762
14196_at	AC012563
14201_at	AL163972
14219_at	AC068667
14248_at	AC007357
14250_r_at	AL050300
14595_at	AL022580

14608_at	AC007357
14621_at	AC004747
14627_i_at	X76609
14628 r at	X76609
14635 s at	AC005398
14636_s_at	AC013258
14675_s_at	D85191
14691_at	AP002046
14704_s_at	AC006067
14709_at	AP002046
14711_s_at	AF085279
14731_s_at	AF014960
14784_at	AC005310
14951_at	AL049481
15057_at	AL035440
15105_s_at	Z14987
15116_f_at	AF121356
15125_f_at	D85190
15141_s_at	D85191
15145_s_at	D64155
15154_s_at	AL096860
15161_s_at	U90522
15178_s_at	U43489
15431_at	AL030978
15496_at	AC006282
15593_s_at	U54561
15611_s_at	L22567
15616_s_at	AJ009696
15846_at	AC006067
15847_g_at	AC006067
15950_at	AC006429
15954_at	U72155
16038_s_at	L04173
16063_s_at	AB008103
16105_s_at	U68017
16150_s_at	AL049730
16153_s_at	AC013258
16393_s_at	AC006436
16504_s_at	Z97335
16510_at	AL034567
16536_s_at	AB008107
16578_s_at	AL137080
16620_s_at	AF051338
16637_s_at	Z97336
16817_s_at	AL096882
16864_i_at	AL133248
17014_s_at	U05206
17033_s_at	U83179

WO 02/22675

AF134128
AF106087
AC005398
AF128396
AC005499
AC006264
X99097
AF107726
AL049862
U40856
AL162751
AC004484
Z97339
AC007323
AL049730
AF188334
111 10055 1
AC012375
AC012375
AC000348
AL096882
AF002109
AL021684
AC006264
AJ222713
U78721
AC004005
AC007396
AC002338
AC002338
AL021768
AC002335
AB035137
AL031804
AL035538
AC004561
AL049659
A71607
AL078470
AB027252
AC002387
AL096882
Y14208
Z97336
AC005309
X91919
ハフェブスブ

Table 14

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Probe Set	Description
12505_s_at	AC005309
12879_s_at	U40856
12904_s_at	AB008103
13189_s_at	U35829
13217_s_at	AL049862
14116_at	AF077407
14148_at	AF224762
14201_at	AL163972
14248_at	AC007357
14250_r_at	AL050300
14691_at	AP002046
14704_s_at	AC006067
15846_at	AC006067
15847_g_at	AC006067
16063_s_at	AB008103
17500_s_at	AL049862
17544_s_at	U40856
18716_at	AC007396
19178_at	AB035137
19640_at	AC004561
20269_at	AC002387

Thus, Hiks1 induced upregulation is compromised in all four tested "loss of Hiks1 resistance mutants" and there is a correlation between breakdown of resistance and deregulation of these genes. This may indicate that these genes play important roles in mediating Hiks1 resistance. Furthermore, these genes appear to act downstream of all four genetically defined RPP7 pathway components, RPP7, EDM1, EDM2 and EDM3. Some genes of this set may encode important regulators; whereas others may indicate metabolic processes required for Hiks1 resistance.

Seventy-eight genes (Table 15; genes comprising SEQ ID NOs:330, 292, 311, 243, 237, 302, 315, 283, 300, 372, 308, 335, 272, 305, 357, 213, 362, 331, 254, 384, 277, 343, 298, 349, 314, 265, 262, 258, 256, 303, 321, 304, 275, 274, 323, 238, 374, 241, 344, 244, 365, 261, 380, 371, 295, 255, 316, 233, 345, 225, 260, 229, 324, 301, 235, 340, 278, 320, 231, 319, 230, 354, 353, 293, 376, 247, 246, 366, 216, 214, 276, 299, 310, 334, 271, 381 and 228) that show the same, but less pronounced, expression characteristics were selected using relaxed selection criterion (at least one 2-fold expression difference over all 12 calculated expression ratios). With a lower cutoff criterion of at least one 2-fold-expression difference across all twelve expression ratios, this cluster is larger

than that shown in Table 14. Genes that are part of this less stringently defined cluster were included in further analyses so as to not exclude potentially important genes. Potential regulators in this cluster are listed in Table 16 (genes comprising SEQ ID NOs: 220, 323, 231, 319, 214, 301, 340, 243, 227, 321, 235, 315, 298, 314, 328, 344 and 349). Four different ERF transcription factors genes are upregulated by the *RPP7*-pathway. Genes encoding ERFs are known to be upregulated in response to several stress-related stimuli, such as wounding, pathogens or ethylene. Moreover, ERF transcription factors bind to GCC boxes, which are ethylene, wounding and pathogen-responsive *cis*-elements. This cluster also contains a gene encoding an ACC synthase, a component of the ethylene biosynthetic pathway. Therefore, ethylene may play a role in the Hiks response pathway.

Table 15

Probe Set	Description
12500_s_at	AF081067
12505_s_at	AC005309
12642_at	AC006920
12879_s_at	U40856
12904_s_at	AB008103
12905_s_at	AB008104
12908_s_at	AB008107
12909_s_at	Z97343
12916_s_at	AF021244
13177_at	AL049640
13187_i_at	U35829
13189_s_at	U35829
13217_s_at	AL049862
13273_s_at	U68017
13284_s_at	AJ002551
13615_at	AC002332
13617_at	AC006592
14116_at	AF077407
14141_at	AC011437
14148_at	AF224762
14201_at	AL163972
14223_at	AL024486
14248_at	AC007357
14250_r_at	AL050300
14614_at	AC004165
14643_s_at	AC006836
14672_s_at	U18993

14691 at	AP002046
14704 s at	AC006067
14706 r at	AL137189
14709 at	AP002046
14711 s at	AF085279
15085_s_at	AL031018
15216_s_at	U75198
15431_at	AL030978
15523_s_at	AL078637
15616_s_at	AL078637
15622_s_at	U43945
15629_s_at	AB010407
15665_s_at	AF022658
15680_s_at	D42061
15846_at	AC006067
15847_g_at	AC006067
15866_s_at	AC005770
15978_at	X68592
16063_s_at	AB008103
16105_s_at	U68017
16393_s_at	AB008103
16412_s_at	U68017
16442_s_at	AJ002551
	AB008107
16539_s_at	Z97343
16569_s_at	L23968
16609_s_at	AB008104
16620_s_at	AF051338
16952_s_at	AC005662
16981_s_at	U35829
17073_s_at	AC006836
17119_s_at	AF132212
17379_at	AC018721
17499_s_at 17500 s at	AF107726 AL049862
	U40856
17544_s_at 17917 s at	AC004261
18070 r at	AC004201
18216 at	AC012375
	AC012375
18217_g_at 18255 at	AC005770
18591 at	X74756
18716 at	AC007396
18876 at	AF002109
19178 at	AB035137
19182 at	AL031804
19640 at	AC004561
20017 at	AC004521
-301,_46	

20269\_at AC002387 20297\_at AC007153 20429\_s\_at Z97336

Additionally, a gene encoding a lipoxygenase, a component of the jasmonic acid biosynthetic pathway, is upregulated. Other regulators potentially acting downstream of RPP7, EDM1, EDM2 and EDM3 include some putative

zinc finger transcription factors and protein kinases as well as two calmodulinlike proteins and a calcium binding protein, which may point to a role of calcium in the RPP7-pathway.

### Table 16

- 10 <u>I. Transcriptional regulators:</u>
  - 1.) AtERF1 (12904 s at, BAA32418.1; AB008103)
  - 2.) AtERF2 (16609 s at, BAA32419.1; AB008104)
  - ·3.) EREBP4-like (12909\_s\_at, CAB10530.1; Z97343)
  - 4.) AtERF5 (16536 s at, BAA32422.1; AB008107)
- 5.) putative CONSTANS-like zinc finger (12505\_s\_at, AAC63643.1; AC005309)
  - 6.) putative C2H2 zinc finger transcription factor (15665\_s\_at, AAB80922.1; AF022658)
  - 7.) putative C3H zinc finger protein (17379\_at, AAF18728.1; AC018721)
- 20 8.) heat shock transcription factor 4 (13273\_s\_at, AAC31756.1; U68017)
  - 9.) SigA binding protein (14148\_s\_at, AAF34713.1; AF224762)
  - II. Other signaling proteins
  - 10.) AtACS-6 (12892 g at, CAB51412.1, AL096882, 35400..37154)
  - 11.) lipoxygenase (16569 s at, AAA32749.1; L23968)
- 25 12.) growth factor like protein (13177\_at, CAB40989.1; AL049640)
  - 13.) serine/threonine protein kinase (16412 s at, CAA18704.1; AL022603)
  - 14.) wall associated kinase 1 (15616 s at, CAB08794.1; AJ009696)
  - 15.) putative receptor-like protein kinase (16393\_s\_at, AAD28318.1; AC006436)
- 30 16.) calmodulin-like (16951 i at, AAC78532.1; AC005662)
  - 17.) calmodulin-like (17500 s at; CAB42906.1; AL049862)
  - 18.) calcium binding protein (17917 s at, AAD12002.1; AC004261)

Twenty-six genes (genes comprising SEQ ID NOs: 300, 308, 272, 213, 362, 265, 374, 241, 261, 380, 310, 228, 233, 330, 311, 243, 237, 254, 230, 244, 365, 216, 316, 345, 225, 301, 278, 354, 353, 293, 335, 271 and 321, also shown in Table 15), represented by 33 probe sets, were found to be commonly upregulated in response to both *Peronospora* isolates, Hiks1 and Emco5, in an *RPP7*- or *RPP8*-dependent (and *EDM1*, *EDM2* and *EDM3*-dependent) manner (Table 17). Elevated expression of these genes may be required for resistance against both *Peronospora* isolates or against *Peronospora parasitica* in general.

transcription factors, two proteins potentially involved in calcium signaling, a lipoxygenase, and a cysteine rich antifungal protein.

Among these 26 genes are those encoding ERF1, putative zinc finger

Table 17

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26 Peronospora (Hiks1 and Emco5) induced RPP8, RPP7, EDM1,2,3-dependent genes

Transcription factors

AtERF1

15

Putative salt-tolerance zinc finger transcription factor (18217 g at)

20 (ZFPI) hypothetical Cys-3-His zinc finger protein Putative C2H2 zinc finger transcription factor Heat shock transcription factor 4

# Other potential proteins

Calmodulin-like protein
 Similarity to centrin, Marsilea vestita contains EF-hand calcium-binding domain
 (15431\_at)
 Lipoxygenase

#### 30 Others

heat shock protein 70 putative steroid sulfotransferase putative glucosyltransferase

 phosphoglycerate dehydrogenase ATAF2
 OPDA-reductase homolog coronatine-induced protein I thioredoxin h

40 IAA-amino acid hydrolase

tryptophan synthase alpha chain similar to xyloglucan fucosyltransferase (12642\_at)
Pad3 (Cytochrome P450)
putative pectinesterase
AIG1
putative glutathione S-transferase
adenosine nucleotide translocator
contains similarity to sugar transporters
CYSTEINE-RICH ANTIFUNGAL
PROTEIN 1 PRECURSOR (AFP1)
(18716 at)

Thirteen of these 26 genes (genes comprising SEQ ID NOs: 308, 300, 272, 362, 265, 242, 261, 380, 228, 243, 254, 216 and 225, Peronospora (Hiks1, Emco5 and Emoy2) induced and RPP4, RPP8 and RPP7 dependent) were also found to be more strongly expressed during the incompatible interaction between the Peronospora isolate Emoy2 and Col-0 plants as compared to Emoy2 infections of the compatible or intermediate type (see Table 18). These 13 genes may play an important role in defense against Peronospora parasitica in general. Among these genes is PAD3, which was previously demonstrated to be required for pathogen-induced phytoalexin production. Strikingly, Hiks1 resistance is, partially compromised in the pad1-1/pad3-1 double mutant, whereas Emoy2 resistance is reduced in the pad3-1 single mutant (Glazebrook et al. 1997).

Table 18

-AIG1 (12879 s at)

```
13 Peronospora (Hiks1, Emco5 and Emoy2) induced
15
                RRP4-pathway, RPP7-pathway, and RPP8-dependent genes
     -adenosine nucleotide translocator (15978 at)
     -phosphoglycerate dehydrogenase (15629 s_at)
20
     -tryptophan synthase alpha chain (14672 s at)
     -ATAF2 (18591 at)
25
     -putative glucosyltransferase (14614 at)
      -calmodulin-like protein (13217 s at)
     -pad3 (cytochrome P450)(14248 at)
30
     -putative pectinesterase (20269 at)
     -ethylene responsive element binding factor 1 (12904 s at)
35
     -heat shock transcription factor 4 (13273 s at)
     -sugar transporter-like (14116 at)
     -similar to xyloglucan fucosyltransferase (12642 at)
40
```

Identification of promoter motifs common to RPP7 and RPP8 controlled genes

Sequence motifs conserved in the promoters of genes co-regulated by the RPP7 or RPP8 pathways may lead to information about the types of transcription factors controlling expression of these genes and may serve as starting points for isolating and/or cloning of such factors. Sub-categorization of RPP7-

upregulated genes by K-means clustering, based on the absolute expression levels of the genes, revealed the existence of only a few basic expression profiles. K-means clustering into five gene sets gave the most consistent and tight clusters. For each of the five gene sets (Table 19), the expression changes in two repetitions of the Col-0 wild type/Hiks1 interaction are shown as well as in the rpp7 mutant and edm1, edm2 and edm3.

The first set comprises genes (genes comprising SEQ ID NOs: 292, 302, 315, 300, 308, 213, 265, 374, 241, 344, 261, 299, 365 and 276) that show early and transient upregulation. The genes peak at 12 hours and return to their ground-states at approximately the 48 hours post infection. Genes of the second set (genes comprising SEQ ID NOs: 267, 305, 357, 362, 380, 310, 228, 295, 233, 329, 312, 243, 237, 283, 372, 254, 314, 323, 216, 260, 229 and 235) are also rapidly upregulated, but show a less pronounced or no decline after the peak. In both sets, the response is weaker in the mutants. Only a few genes in the first set respond almost as strongly in *edm2* and *edm3* plants as in wild type plants.

- Genes of the third (genes comprising SEQ ID NOs: 258, 256, 303, 304, 275, 244, 316, 354 and 225) and fifth (genes comprising SEQ ID NOs: 330, 384, 277, 343, 298, 349, 262, 274, 238 and 301) set show an almost linear increase of transcript level in wild type. In the mutants, this increase is delayed and in some cases less steep. In the fourth set (genes comprising SEQ ID NOs: 432, 347,
- 370, 412, 323, 411, 322, 449, 448, 385, 471, and 339) the expression levels rise in the first repetition of the Col-0 wild type/Hiks interaction, but respond very little in the second one. In *edm3*, the transcript levels drop from a high ground state. The probe sets and genes corresponding to each of these K-means cluster sets are shown in Table 19.

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Table 19

K-means set 1	Description	K-means set2	Description
12500_s_at	AF081067	12642_at	AC006920
13217_s_at	AL049862	12904 s at	AB008103
14248_at	AC007357	12905_s_at	AB008104

14614_at	AC004165	12916 s at	AF021244
14706_r_at	AL137189	13177_at	AL049640
15216 s at	U75198	13187_i_at	U35829
15622 s_at	U43945	13273 s at	U68017
15629 s_at	AB010407	14116 at	AF077407
15680_s_at	D42061	14223_at	AL024486
15978_at	X68592	14672_s_at	X76609
16412_s_at	U68017	15085 s at	AL031018
17500_s_at	AL049862	15523 s at	AL078637
18070 r_at		15866 s at	AC005770
18716_at	AC007396	16063_s_at	AB008103
19640_at	AC004561	16105 s at	AL049730
	•	16393_s_at	AC006436
		16609 s at	AB008104
	•	16952 s_at	AC005662
K-means set 3	Description	18255 at	AC005770
12879_s_at	U40856	18591 at	X74756
13284_s_at	AJ002551	19182 at	AL031804
14148_at	AF224762	20269_at	AC002387
14201_at	AC068667	_	
14704_s_at	AC006067		
15846_at	AC006067	K-means set 5	Description
15847 <u>g</u> at	AC006067	13189_s_at	U35829
16442_s_at	AJ002551	14250_r_at	AL050300
17544_s_at	U40856	14691_at	AP002046
		14709_at	AP002046
K-means set4	Description	15616_s_at	AJ009696
12908_s_at	AB008107	15665_s_at	AF022658
	Z97343	16981_s_at	U35829
13617_at	AC006592	17499_s_at	AF107726
14141_at	AC011437	17917_s_at	AC004261
14711_s_at	AF085279	20429_s_at	Z97336
15431_at	AL030978		
16536_s_at	AB008107		
16539_s_at	Z97343		•
17379_at	AC018721		
18216_at	AC012375		
18217_g_at	AC012375		•
20017 at	AC004521		•
2001/_4	AC004321		

Recently, the plant specific family of WRKY transcription factors has been implicated in the regulation of defense-associated genes. Promoters of genes upregulated during systemic acquired resistance were found to be strongly enriched in potential WRKY binding sites. WRKY factors comprise a large family of 72 different members in *Arabidopsis*. Generally these factors appear to bind to sequence motifs containing the core T G A C (W box). It has not yet

been determined whether individual WRKY family members differ in their preferences for distinct W box variations. However, it is probable that a given WRKY protein has a binding preference for a certain variation of the W box motif.

Using the program AlignACE, three variations of potential WRKY binding sites were found to be conserved in sub-sets of RPP7 controlled genes. Two of these motifs are highly enriched in promoters of genes from K-means cluster 3, representing genes that show a linear increase in transcript level after Peronospora infection (Table 20; genes comprising SEQ ID NOs: 384, 298, 349, 262 and 238 for W box I and genes comprising SEQ ID NOs: 384, 349, 262, 238 and 301 for W box II). These motifs are also present in promoters of K-means cluster 5, which represents a similar expression profile. Focusing on sustained up-regulated genes, that show a particularly pronounced expression difference between Col-0 wild type plants and the Hiks1 response mutants, a third variation of potential WRKY binding sites was found to be significantly enriched (Table 21; genes comprising SEQ ID NOs: 397, 371, 238, 262, 256, 275, 254, 214 and 225). In all three cases, sequence conservation clearly exceeds the core motif of WRKY binding sites that appears to be recognized by all members of this family. These additionally conserved positions may confer specificity for certain WRKY family members. They may provide a highly defined binding site preferentially recognized by a distinct WRKY factor or may serve as binding sites for another transcription factor binding closely adjacent to and specifically interacting with a distinct WRKY factor. In any case, the high conservation of extended potential WRKY binding motifs strongly suggests that specific members of this family participate in the regulation of certain sub-sets of RPP7controlled genes.

# Table 20

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	W	box	I	MAP	Score:	8.	9811	. 4			
30				G!	<b>ICATCTT</b>	TTA	ATCI	CTGG	0	638	1
				G'	<b>ICATCTT</b>	TTA	ATCG	CCGG	0.	932	1
				G	CCTTTGC	TTA	TTTA	TAGG	0	1036	5 1
				G!	<b>ICATTTG</b>	TTA	CAAA	GAGG	1	318	1
				G'	<b>ICATTGA</b>	TCA	TAAA	CTGT	1	389	0
35				G	CCACTGC	TGA	ATTG	TCGG	1	494	1
				G!	<b>ICCATTG</b>	TCA	ATAA	ATGG	2	689	1
				G:	<b>ICCATTG</b>	TAA	ATAA	ATGT	2	949	1

```
GTCATCTTTTAATCTCTGT 3 475 0
              GTCACGTATGAATGGAAGG 4 73 1
              GTCAACGTTTAGTTCATGT 4 226 0
              GTCAAGTTTTAAATTGTGG 4 352 1
 5
              GTCAACGTTTAGTTCATGT 4 401 0
                     * * * *
              GTCANNTNTNANTNNNNGG
                     G
                           A
    Expected frequency by chance: 0.02/1kb
10
    Observed frequency:
                                   1.2/1kb
    Enrichment: ~60 fold
    W box II MAP Score: 15.2966
              ATTAAAAGATGAC 0 638 0
              ATTAAAAGATGAC 0 932 0
15
              ATTAAATGCTGTC 2 752 0
              ATTAAATGCTGTC 2 1011 0
              AGCAAAAGCTGAC 2 1092 1
              ATTAAAAGATGAC 3 481 1
              ATCAAAAGTTGTC 3 829 1
20
              ACTAAACGTTGAC 4 232 1
              ACTAAACGTTGAC 4 407 1
              ACTAAAAAGTGAC 5 755 1
               * **** * ****
25
    W II
              ANTAAANGNTGAC
                C
                     A
                          Т
    W I:
              CCNNNNANTNANCNNTGAC
30
    Expected frequency by chance: 0.04/1kb
    Observed frequency:
                                   1.35/1kb
    Enrichment: ~30 fold
    Table 21
    ATAGGTGGTCAAGT 1 106 0
35
    AATTGTGGTCATTT 1 823 0
    ACTTGTGGTCAATT 2 804 0
    AAAAGGGGTCATTT 2 970 1
    ATATGTCGTCTCTT 2 994 0
    AGTTGTGGTCTACC 3 502 0
40
    AAAAGTTGTCAATT 3 732 1
    AGACGTCGTAATTT 4 400 0
    ACGTGGCGTCATAT 5 179 0
    ATGTGGCGTCTCCT 6 249 1
    AGTTGGTGTCACGT 6 925 1
45
    ATTCGTGGTCAACT 7 582 1
    ATATGTCGTCACTT 7 875 1
       *****
    ANNNGTNGTCANNT
         G
```

expected: 0.05/1kb; in random set: 0/1kb; this set: 1/1kb; enrichment: 10-20 fold;

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Novel, conserved promoter motifs were also found. Genes of K-means set 1, which are early and transient *RPP7*-pathway specific genes, share two similar highly conserved promoter motifs, G G T/C C C A (SEQ ID NO:714; genes comprising SEQ ID NOs: 302, 315, 308, 265, 374, 241, 261, 299 and 365) and G N C C A A A (SEQ ID NO:715; genes comprising SEQ ID NOs: 292, 302, 308, 265, 374, 241, 261, 299, 365 and 276) (Table 22). Both motifs are almost perfectly conserved and are represented in nearly all promoters of this gene set. They are compact six- or seven-mers, which is typical for transcription factor binding sites. For the second motif, the permutations G A C C A A A and G T C C A A A are strongly represented. These permutations may constitute the preferred binding site of a so far unknown transcription factor.

#### Table 22

Two related novel motifs are highly enriched in all 11 promoters of K-means set1

```
GGTCCA 1 232 0
    GGCCCA 1 289 1
    GGTCCA 1 597 1
25
    GGTCCA 2 517 0
    GGTCCA 3 211 1
    GGCCCA 3 360 1
    GGTCCA 4 597 1
    GGCCCA 4 681 1
30
    GGTCCA 5 352 0
    GGTCCA 5 1060 1
    GGTCCA 6 358 0
    GGCCCA 7 776 0
    GGCCCA 7 816 0
35
    GGTCCA 8 285 0
    GGTCCA 9 888 1
    *****
    GGTCCA
           C
    Expected frequency by chance: 0.22/1kb
    Observed frequency:
                                   1.2/1kb
    Enrichment: 5.4 fold
    GCCCAAA 0 601 1
    GTCCAAA 10 1186 1
    GTCCAAA 9 562 1
```

```
GACCAAA 8 640 0
    GCCCAAA 7 774 0
    GTCCAAA 7 717 1
    GTCCAAA 6 712 1
5
    GACCAAA 5 970 1
    GTCCAAA 5 350 0
    GACCAAA 4 1164 1
    GACCAAA 4 1072 1
    GTCCAAA 4 784 0
10
    GACCAAA 4 714 0
    GGCCAAA 4 698 0
    GCCCAAA 3 361 1
    GACCAAA 1 920 0
    GTCCAAA 1 230 0
15
    • * * * * *
    GNCCAAA
    Expected frequency by chance: 0.32/1kb
    Observed frequency:
                                    1.3/1kb
    Enrichment: 4 fold
```

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Following a similar approach as for the Hiks1-induced RPP7 pathway controlled genes, a cluster of early and transiently Emco5-induced RPP8dependent genes were defined (Table 23). In Col-0-RPP8 plants these genes (genes comprising SEQ ID NOs: 364, 341, 288, 378, 363, 379 and 219) show a very pronounced upregulation 12 hours post Emco5 infection, after which they return to expression ground states. Using AlignACE, two different motifs (see Example 2) were found to be strongly enriched in the promoters of this cluster. The first motif strongly resembles the type I consensus binding site of Myb transcription factors. This motif is also almost identical to a recently identified cis-element conferring rapid elicitor responsive gene expression in parley cell culture (Kirsch et al., 2001). Interestingly, a gene encoding a Myb-like transcription factor (highlighted in yellow) is co-regulated with other genes of this cluster. The Myb-like transcription factor gene is the only gene of this cluster that does not contain this conserved motif in its promoter. This MYBlike factor could be a regulator of this cluster. The second motif does not resemble any known stress responsive plant cis element or transcription factor binding site. However, the second motif was found to be strongly represented in promoters from other defense related genes, such as SAR genes. As is typical of many transcription factor binding sites, certain permutations of its consensus

sequence may constitute palindromes. This motif was also found to be conserved by the program MEME.

Table 23

5 Glutathinone-conjugate transporter AtMRP4 putative GST putative cytochrome P450 (AC002340) PAD3 (similar to cytochrome P450) CTF2B (similarity to proteins involved in

10 Hydroxylation and oxidation of aromatic rings)Ribonuclease RNS1

Putative Myb-like protein

AlignACE Motif 1:

CAACTTTGAC

AA TT

15 type I Myb binding site:

CAACTG T G

Frequency by chance: 0.05/1kb

Observed frequency in control cluster: 0.13/1kb Observed frequency in this cluster: 1.10/1kb

AlignACE Motif2:

TTGGGNCNAA

A A

**MEME** motif:

20

**GTCTNTTGGGNCAAAA** 

TT AGC

25 Frequency by chance: 0.13/1kb

Observed frequency in control cluster: 0/1kb
Observed frequency in this cluster: 1.2/1kb

Different types of transcription factors may participate in the control of 30 distinct RPP7-pathway dependent expression profiles. Promoters of genes showing a linear sustained increase of transcript are enriched in different variations of WRKY binding sites. However, no WRKY-pathway gene was found to be upregulated by the RPP7 pathway. Therefore, WRKY factors involved in RPP7 mediated up-regulation of K-means cluster 3 and 5 genes may 35 already be pre-formed when the pathogen signal is perceived. However, only 21 WRKY genes out of 72 members of this family in Arabidopsis are represented on the oligonucleotide chip. Upregulated WRKY factors participating in RPP7 mediated gene regulation may not be present on the chip. A novel type of transcription factor may be involved in regulation of early and transient RPP7-40 controlled genes (K-means cluster 1). Promoter stretches containing this motif may be used in yeast one hybrid screenings for novel factors. In addition, transgenic plants carrying appropriate promoter-reporter constructs could be used

as a basis for mutant screenings with the aim of identifying regulators acting on this motif.

A Myb-like transcription factor appears to be involved in the regulation of early and transient *RPP8*-controlled genes. A gene showing a similar pattern of regulation may encode a candidate factor controlling this cluster. The potential role of this factor in regulating *RPP8*-controlled genes may be examined using T-DNA insertion mutants.

# Potential constitutive effects of the RPP7-pathway

In order to focus on genes showing multiple expression differences between Col-0 wild type plants and Hiks response mutants, all genes that showed at least four 1.8-fold expression differences across all comparisons were selected. The resulting clustergram contains the majority of RPP7 pathwaydependent Hiks1-induced genes. Two more clusters of genes behaving consistently in all four tested Hiks1-response mutants were found. The first cluster comprises genes that show at all tested time points (including the 0 hpi time point) elevated transcript levels in Col-0 wild type plants as compared to all four mutants (Table 24; genes comprising SEQ ID NOs: 364, 288, 378, 363, 379, 219, 399, 389 and 390). In most cases, expression levels are not affected by Hiks1 infection. These elevated expression levels may be directed by constitutive (non Hiks1-triggered) signal flow via the RPP7 pathway. This signal flow is disrupted in the rpp7, edm1, edm2 and edm3 mutants. Constitutive expression of these genes may be required for effective disease resistance and may be a prerequisite for induction of subsequent defense responses. A HD-ZIP transcription factor, which is included in this cluster, may act as a pre-formed regulator controlling genes responding to Hiks1-recognition. Its potential role in regulating RPP7 pathway-dependent genes may be examined using T-DNA insertion mutants.

Table 24

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The RPP7 pathway constitutively activates a cluster of genes including a HD-ZIP transcription factor

PROBE SET	DESCRIPTION
14965_at	AC002329
18635_at	AC004005
14295 s at .	Z54356
17386 at	AC006264

15145 s at	D64155
20201_at	AL078470
18607_s_at	U78721
20227 s at	AC007153
12761 s at	AC006577
20555 s a	AL080318

Genes of the second cluster show the opposite behavior (Table 25). Their transcript levels are elevated in all four Hiks1 response mutants as compared to Col-0 wild type plants. Hiks1-infection does also not affect their expression levels. Surprisingly, this cluster contains six genes encoding peroxidases. Since peroxidases are believed to play roles in conferring disease resistance, the biological significance of this phenomenon is obscure. Perhaps the mutant plants "realize" disruption of the RPP7-defense pathway and compensate for their reduced defense capacity by constitutively up-regulating some defense mechanisms. Alternatively, elevated expression of these peroxidases may be advantageous for invading Peronospora hyphae. Therefore, expression of these genes could be constitutively reduced by the RPP7 pathway.

Table 25

5

<b>Probe Set</b>	Description
15982_s_at	X98190
15954_at	U72155
12333_at	AJ286345
19594_i_at	X98321
15969_s_at	AJ133036
16462_s_at	.AC004683
16461 i at	AC004683
12324_i_at	AC007212
12778_r_at	AC006577
16514 at	AL035538
17549 s_at	L37126
14028 at	AF075597
17054_s_at	AF134128

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# Predictions about RPP7-pathway hierarchy

Clustering of the Hiks1-infection data in the "treatment" dimension, instead of the "gene" dimension, revealed that at all three infection time points, gene expression profiles of the *rpp7* mutant resemble most those of the *edm1* mutant. Similarly, expression profiles of the *edm2* mutant resemble those of the

edm3 mutant. This may indicate that RPP7 and EDM1 closely act together at a certain level in the RPP7 signalling cascade, whereas EDM2 and EDM3 may act closely together at a different level of the pathway. Close interaction of RPP7 and EDM1 is also suggested by a T-DNA tagged rpp7 mutant that phenotypically resembles the edm1 mutant. In contrast to rpp7, edm2 and edm3, which appear only to be compromised in resistance against the Peronospora isolate Hiks1, edm1 is also compromised in resistance to other Peronospora isolates that are avirulent on Col-0 plants. Like edm1, the putatively T-DNA tagged rpp7 mutant is also susceptible to a variety of normally Col-0 incompatible Peronospora isolates. This observation may indicate that the EDM1 protein requires a portion of the RPP7 protein for proper function, which is disrupted in the T-DNA tagged rpp7 mutant. Assuming that RPP7, which appears to constitute an NBS-LRR receptor, acts at the top of the Hiks1 recognition pathway hierarchy, EDM1 could act at a high level as well and EDM2 and EDM3 may act more downstream in the cascade.

Table 26

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Hiks1
↑↓

RPP7 / EDM1
↓

EDM2 / EDM3
↓

RESISTANCE

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Table 27 shows the plant and fungal orthologs of the *Arabidopsis* sequences identified herein.

```
Table 27
```

Query= AIG1 s at 12879 s at /id source genbank /description gb|aac49282.1| (u40856) aig1 [arabidopsis thaliana] /blast\_score 1.00e-150

5 (1381 letters) -

60

Database: plantfungal

661,018 sequences; 426,114,510 total letters

10 Searching.....done

> Score Ε

Sequences producing significant alignments:

(bits) Value

15 gb|U64925|NTU64925 Nicotiana tabacum geranylgeranylated protein ... 260 1e-68 emb|AW625701|AW625701 EST319608 tomato radicle, 5 d post-imbibit... 228 6e-59 emblAW720227|AW720227 LjNEST17c4r Lotus japonicus nodule library... 219 4e-56 emb|AW220184|AW220184 EST302667 tomato root during/after fruit s... 208 7e-53 emb[AI774580]AI774580 EST255680 tomato resistant, Cornell Lycope... 173 2e-42 -emb|AW685484|AW685484 NF030E02NR1F1000 Nodulated root Medicago t... 171 2e-41 emb|AI443867|AI443867 sa44d09.yl Gm-c1004 Glycine max cDNA clone... 170 3e-41 emb[AW397252]AW397252 sg76f06.y1 Gm-c1007 Glycine max cDNA clone... 168 1e-40 emblAW033368lAW033368 EST276939 tomato callus, TAMU Lycopersicon... 162 5e-39 emb|AV417858|AV417858 AV417858 Lotus japonicus young plants (two... 153 4e-36 25 emblAI780050|AI780050 EST260929 tomato susceptible, Cornell Lyco... 138 9e-32 emb|AW039095|AW039095 EST281068 tomato mixed elicitor, BTI Lycop... 111 2e-23 emb|AI780139|AI780139 EST261018 tomato susceptible, Cornell Lyco... 86 7e-16 emb|AW164180|AW164180 Ljimpest21-672-c8 Ljimp Lambda HybriZap ... 84 2e-15 emb|AI781596|AI781596 EST262475 tomato susceptible, Cornell Lyco... 75 1e-12 30 emb|AW774764|AW774764 EST333915 KV3 Medicago truncatula cDNA clo... 62 7e-09 emb|AW348781|AW348781 GM210003A22H11R Gm-r1021 Glycine max cDNA ... 54 2e-06 emb|AI491210|AI491210 EST241919 tomato shoot, Cornell Lycopersic... 29 0.010 emb|AW651526|AW651526 EST329980 tomato germinating seedlings, TA... 29 0.011 emblAW220594|AW220594 EST296979 tomato fruit mature green, TAMU ... 29 0.011 emb[AW926585]AW926585 HVSMEg0007J19 Hordeum vulgare pre-anthesis... 29 0.025 emb|AW032321|AW032321 EST275775 tomato callus, TAMU Lycopersicon... 28 0.026 emb|AW736598|AW736598 EST333090 KV3 Medicago truncatula cDNA clo... 37 0.39 emb|AW736597|AW736597 EST333089 KV3 Medicago truncatula cDNA clo... 37 .0.39 emb|AW713727|AW713727 h1f12ne.f1 Neurospora crassa evening cDNA ... 35 1.0 40 emb|AW711542|AW711542 f3g07ne.f1 Neurospora crassa evening cDNA ... 35 1.0 emb|AB009972|AB009972 Aspergillus oryzae gene for beta-1,4-xylos... 35 1.0 emb|AW713709|AW713709 h1e09ne.fl Neurospora crassa evening cDNA ... 35 1.0 emb|AW712721|AW712721 g3a07ne.f1 Neurospora crassa evening cDNA ... 35 1.0 emb|AW709185|AW709185 d3e02ne.f1 Neurospora crassa evening cDNA ... 35 1.0 45 emb[Y09354|SPABC1 S.pombe ABC1 gene. emb|Z99262|SPAC9E9 S.pombe chromosome I cosmid c9E9. 35 1.4 emb|AJ225108|STA225108 Solanum tuberosum (cultivar Bintje) mitoc... 35 1.4 emb|AW933326|AW933326 EST359169 tomato fruit mature green, TAMU ... 26 1.5 emb|Z25870|CACDC10G C.albicans CDC10 gene for cell division cycl... 31 1.5 50 emb[AC008368]AC008368 Trypanosoma brucei chromosome II clone RPC... 35 1.9 emb|AQ942780|AQ942780 Sheared DNA-42B15.TR Sheared DNA Trypanoso... 35 1.9 emb|AQ953508|AQ953508 Sheared DNA-33G8.TR Sheared DNA Trypanosom... 35 1.9 emb|AC009463|AC009463 Trypanosoma brucei chromosome II clone RPC... 35 1.9 emb|AQ950237|AQ950237 Sheared DNA-39C11.TF Sheared DNA Trypanoso... 35 1.9 55 emblAQ643883|AQ643883 RPCI93-DpnII-26C13.TV RPCI93-DpnII Trypano... 35 1.9 emb|AQ948491|AQ948491 Sheared DNA-49H2.TF Sheared DNA Trypanosom... 35 1.9 emb|AW727289|AW727289 GA\_Ea0011H24 Gossypium arboreum 7-10 dpa ... 35 1.9 emb|AW729536|AW729536 GA \_\_Ea0025E24 Gossypium arboreum 7-10 dpa ... 35 1.9 gb|L36856|PEAIAP34A Pisum sativum GTP-binding protein (IAP34) mR... 28 2.1

emb|Z28341|PSCLOEP P.sativum (miranda) mRNA for chloroplast oute... 28 2.1

	emb AW776020 AW776020 EST335085 DSIL Medicago truncatula cDNA cl       28 2.2         emb AW685649 AW685649 NF032G04NR1F1000 Nodulated root Medicago t       28 2.2         emb AW690755 AW690755 NF037H10ST1F1000 Developing stem Medicago       28 2.2
_	emb AW256896 AW256896 EST305033 KV2 Medicago truncatula cDNA clo
5	emb AV422565 AV422565 AV422565 Lotus japonicus young plants (two 28 2.3
	emb AW832303 AW832303 sm07c04.yl Gm-c1027 Glycine max cDNA clone 27 2.3
	emb AL136538 SPAC30 S.pombe chromosome I cosmid c30. 34 2.6
	emb AI724721 AI724721 RHIZ1 26 C05.y2 A001 Rhizomel Sorghum hale 34 2.6
10	emb AW256609 AW256609 EST304746 KV2 Medicago truncatula cDNA clo 34 2.6
10	emb A1775696 A1775696 EST256796 tomato resistant, Cornell Lycope 28 3.1
	emb Z71682 SCYNR067C S.cerevisiae chromosome XIV reading frame O 34 3.6 emb AI776315 AI776315 EST257415 tomato resistant, Cornell Lycope 34 3.6
	emb AF230371 AF230371 Lycopersicon esculentum allene oxide synth 34 3.6 emb AA741645 AA741645 LmLv39p3/255A Leishmania major promastigot 34 3.6
15	
	4 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
	emb AQ399149 AQ399149 mgxb0019C01f CUGI Rice Blast BAC Library P 33 5.0 emb AW615911 AW615911 EST325409 tomato flower buds 0-3 mm, Corne 33 5.0
	emb AW035453 AW035453 EST281191 tomato callus, TAMU Lycopersicon 33 5.0
20	emb AF051695 AF051695 Trypanosoma cruzi sialidase homolog (P85.1 33 5.0
	-emb Z37538 LTGRR4-L-tarentolae mRNA encoding putative NADH subun 28 5.5
	emb X85021 SCXLTORFS S.cerevisiae DNA from left arm of chromosom 33 6.8
	gb L25088 YSCMEF2MIT Saccharomyces cerevisiae nuclear-encoded mi 33 6.8
	emb AQ443826 AQ443826 GSSTc01287 Trypanosoma cruzi random genomi 33 6.8
25	emb AQ502942 AQ502942 V47H4 mTn-3xHA/lacZ Insertion Library Sacc 33 6.8
	emb Z85962 MSZ85962 Musa sp. DNA for sequence tagged microsatell 33 6.8
	emb Z49377 SCYJL102W S.cerevisiae chromosome X reading frame ORF 33 6.8
	emb AW224537 AW224537 EST302980 tomato root, plants pre-anthesis 31 7.5
	emb AW257183 AW257183 EST305320 KV2 Medicago truncatula cDNA clo 32 9.4
30	emb AQ642922 AQ642922 RPCI93-DpnII-27E7.TV RPCI93-DpnII Trypanos 32 9.4
	emb AW738509 AW738509 EST339936 tomato flower buds, anthesis, Co 32 9.4
	emb AW217351 AW217351 EST296174 tomato flower buds 0-3 mm, Corne 32 9.4
	emb AW219255 AW219255 EST301737 tomato root during/after fruit s 32 9.4
٠.	emb AC007865 AC007865 Trypanosoma brucei chromosome II clone RPC 32 9.4
35	emb AW684118 AW684118 NF012F02NR1F1000 Nodulated root Medicago t 32 9.4
	emb AW622239 AW622239 EST313037 tomato root during/after fruit s 32 9.4
	emb AQ651543 AQ651543 Sheared DNA-7C22.TF Sheared DNA Trypanosom 32 9.4
	emb Y11565 NC11565 N.crassa acu-15 gene. 32 9.4
40	emb AW775944 AW775944 EST335009 DSIL Medicago truncatula cDNA cl 32 9.4
40	emb AB014493 AB014493 Gibberella zeae gene for reductase, partia 32 9.4
	Onome ASA1 a at 1980 a at //d assessment /dasaistics
	Query= ASA1_s_at 12889_s_at /id_source genbank /description gb aaa32738.1  (m92353) anthranilate synthase alpha subunit
45	[arabidopsis thaliana] /blast score 0 /ec number /family /chip nova
73	/gb_link /ncgi
	(1788 letters)
	(1700 letters)
	Database: plantfungal
50	661,018 sequences; 426,114,510 total letters
	, , , , , , , , , , , , , , , , , , ,
	Searchingdone
	•
	Score E
<b>55</b>	Sequences producing significant alignments: (bits) Value
	(,
	gb L34344 RTAANTSYNB Ruta graveolens anthranilate synthase alpha 772 0.0
	gb L34343 RTAANTSYNA Ruta graveolens anthranilate synthase alpha 467 0.0
	emb AF079168 AF079168 Nicotiana tabacum feedback-insensitive ant 365 0.0
60	emb AW931942 AW931942 EST357785 tomato fruit mature green, TAMU 356 3e-97
	emb AW218352 AW218352 EST303535 tomato radicle, 5 d post-imbibit 210 1e-73

```
emb|AL031966|SPCC1442 S.pombe chromosome III cosmid c1442.
                                                                      209 6e-69
      emb|AW651095|AW651095 EST329549 tomato germinating seedlings, TA... 222 2e-64
      dbj|D89256|D89256 Schizosaccharomyces pombe mRNA, partial cds, c... 205 5e-64
      emb|AW982499|AW982499 HVSMEg0003G22f Hordeum vulgare pre-anthesi... 215 2e-58
      emb|AW460005|AW460005 si07d11.yl Gm-c1029 Glycine max cDNA clone... 116 1e-57
      gb|U18839|SCE9747 Saccharomyces cerevisiae chromosome V cosmids ... 123 4e-48
      emb|X68327|SCTRP2 S.cerevisiae TRP2 gene for anthranilate syntha... 123 4e-48
      emb|AW719463|AW719463 LjNEST5b1r Lotus japonicus nodule library.... 184 1e-45
      emb|AI736775|AI736775 sb33d01.yl Gm-c1012 Glycine max cDNA clone... 104 2e-40
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      gb|K01388|YSCTRP2 Yeast (S.cerevisiae) TRP2 gene coding for anth... 122 4e-38
      emb|AL032684|SPBP8B7 S.pombe chromosome II p1 p8B7.
      emb[AW509018]AW509018 si39b01.yl Gm-r1030 Glycine max cDNA clone... 113 5e-24
      gb[T14852|T14852 crs299 lambdaZAPST Ricinus communis cDNA clone ... 107 2e-22
      emb|AW223881|AW223881 EST300692 tomato fruit red ripe, TAMU Lyco... 103 4e-21
15
      emb|AF119554|AF119554 Plasmodium falciparum para-aminobenzoic ac... 67 6e-10
      emb|AL111470|CNS019CM Botrytis cinerea strain T4 cDNA library un... 45 2e-05
      emb|AI329873|AI329873 b9g02ne.rl Neurospora crassa evening cDNA ... 48 3e-04
      emb|AW224247|AW224247 EST300974 tomato fruit red ripe, TAMU Lyco... 46 7e-04
      emb|AF149719|AF149719 Aspergillus fumigatus para aminobenzoic ac... 42 0.012
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      emb|AQ448372|AQ448372 mgxb0020C12f CUGI Rice Blast BAC Library P... 39 0.15
      emb|AQ324360|AQ324360 mgxb0018K01r CUGI Rice Blast BAC Library P... 39 0.15
      emb|AQ160089|AQ160089 mgxb0003G09r CUGI Rice Blast BAC Library P... 39 0.15
      emblAW599019|AW599019 gb01b03.y1 Moss EST library PPN Physcomitr... 37 0.38
      emb|AW599000|AW599000 ga99h03.y1 Moss EST library PPN Physcomitr... 37 0.38
25
      emb|AW678847|AW678847 WS1_1_A04.g1_A002 Water-stressed 1 (WS1) S... 35 0.88
      emb|AW680390|AW680390 WS1 52 D12.g1 A002 Water-stressed 1 (WS1) ... 35 0.89
      emb|AW678385|AW678385 WS1_15_H06.g1_A002 Water-stressed 1 (WS1) ... 35 0.89
      emb|AW747146|AW747146 WS1_66 E07.b1 A002 Water-stressed 1 (WS1) ... 35 0.89
      emb|AW678361|AW678361 WS1 15 H06.b1 A002 Water-stressed 1 (WS1) ... 35 0.90
30
      emb|AW745749|AW745749 WS1 37 D12.b1 A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW747427|AW747427 WS1 68 B09.b1 A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW678071|AW678071 WS1 13 E01.b1 A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW747468|AW747468 WS1 68 B09.g1 A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW672427|AW672427 LG1 359 A06.g1 A002 Light Grown 1 (LG1) So... 35 0.90
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      emb|AW744836|AW744836 LG1 384 E07.g1 A002 Light Grown 1 (LG1) So... 35 0.90
      emb|AW746170|AW746170 WS1_39_B05.g1_A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW922317|AW922317 DG1 17 E06.g1 A002 Dark Grown 1 (DG1) Sorg... 35 0.91
      emb|AQ648582|AQ648582 RPCI93-EcoRI-1M22.TP RPCI93-EcoRI Trypanos... 36 0.99
      emb|AQ643551|AQ643551 RPCI93-EcoRI-3124.TJ RPCI93-EcoRI Trypanos... 36 0.99
40
      emb|AI443370|AI443370 sa31b05.x1 Gm-c1004 Glycine max cDNA clone... 35 1.4
      emb|AW101313|AW101313 sd77d08.y1 Gm-c1009 Glycine max cDNA clone... 35 1.4
      emb|AW678030|AW678030 WS1_12 B10.g1_A002 Water-stressed 1 (WS1) ... 35 1.4
      emb|AW349006|AW349006 GM210004A12H10R Gm-r1021 Glycine max cDNA ... 35 1.4
      emb|AW309961|AW309961 sf27b12.x1 Gm-c1028 Glycine max cDNA clone... 35 1.4
45
      emb|AW678582|AW678582 WS1_16_E09.g1_A002 Water-stressed 1 (WS1) ... 35 1.4
      emb|AW317198|AW317198 sf38f03.x1 Gm-c1028 Glycine max cDNA clone... 35 1.4
      emb|AW678305|AW678305 WS1 14 G05.g1 A002 Water-stressed 1 (WS1) ... 35 1.4
      emb|AW680905|AW680905 WS1_8 A08.b1 A002 Water-stressed 1 (WS1) S... 35 1.4
      emb|AW679666|AW679666 WS1_30_B11.g1_A002 Water-stressed 1 (WS1) ... 35 1.4
50
      emb|AL031746|PFMAL1P3 Plasmodium falciparum MAL1P3, complete seq... 35 1.9
      emb|AI959816|AI959816 sc94f02.yl Gm-c1019 Glycine max cDNA clone... 35 1.9
      emb|AW924277|AW924277 WS1_52_D12.b1_A002 Water-stressed 1 (WS1) ... 35 1.9
      gb|J03998|PFAGAR Plasmodium falciparum glutamic acid-rich protei... 35 1.9
      emb|AW696796|AW696796 NF109A07ST1F1052 Developing stem Medicago ... 35 1.9
55
      emb|AW396753|AW396753 sf37c11.x1 Gm-c1028 Glycine max cDNA clone... 35 2.6
      gb|M98871|SOYCHS7A Glycine max chalcone synthase (chs7) gene, co... 35 2.6
      emb|AI460797|AI460797 sa69d02.yl Gm-c1004 Glycine max cDNA clone... 35 2.6
      emb|AW348617|AW348617 GM210002B22G1R Gm-r1021 Glycine max cDNA 3... 35 2.6
      emb|AW310362|AW310362 sf35a09.x1 Gm-c1028 Glycine max cDNA clone... 35 2.6
60
      emb|AI437832|AI437832 sa40c07.yl Gm-c1004 Glycine max cDNA clone... 35 2.6
      gb|BE023927|BE023927 sm94c05.yl Gm-c1015 Glycine max cDNA clone ... 35 2.6
```

	emb AW424189 AW424189 sh62b04.y1 Gm-c1015 Glycine max cDNA clone 35 2.6 emb AW101907 AW101907 sd72d01.y1 Gm-c1008 Glycine max cDNA clone 35 2.6 emb AW102370 AW102370 sd86h01.y1 Gm-c1009 Glycine max cDNA clone 35 2.6
5	emb AW309356 AW309356 sf16d02.x1 Gm-c1028 Glycine max cDNA clone 35 2.6 emb AL355932 NCB5O22 Neurospora crassa DNA linkage group II BAC 34 3.5 emb AQ652663 AQ652663 Sheared DNA-20A9.TR Sheared DNA Trypanosom 34 3.5 emb Z98056 SPAC5D6 S.pombe chromosome I cosmid c5D6. 34 3.5 emb AW348286 AW348286 GM210001B23B6R Gm-r1021 Glycine max cDNA 3 34 4.
10	emb AB018422 AB018422 Pisum sativum mRNA for DNA binding zinc fi 34 4.9 emb AW734949 AW734949 sk93b10.y1 Gm-c1035 Glycine max cDNA clone 34 4.9 emb AW679089 AW679089 WS1_22_A07.g1_A002 Water-stressed 1 (WS1) 34 4.9 emb AW306776 AW306776 sf48c12.y1 Gm-c1009 Glycine max cDNA clone 34 4.9 gb M36941 BLYHORDCA Hordeum vulgare C-hordein gene, complete cds. 29 5.3
15	gb BE034677 BE034677 ML01H08 ML Mesembryanthemum crystallinum cD 33 6.7 emb Z26877 SCDCHR11 S.cerevisiae (S288C) 36.2kb DNA fragment fro 33 6.7 emb AW267708 AW267708 EST305836 DSIR Medicago truncatula cDNA cl 33 6.7 emb Z28162 SCYKL162C S.cerevisiae chromosome XI reading frame OR 33 6.7 emb AW980990 AW980990 EST392143 GVN Medicago truncatula cDNA clo 33 6.7
20	gb BE037412 BE037412 MP20G03 MP Mesembryanthemum crystallinum cD 33 6.7 emb Z28161 SCYKL161C S.cerevisiae chromosome XI reading frame OR 33 6.7 emb AI726247 AI726247 BNLGHi5399 Six-day Cotton fiber Gossypium 33 6.7 emb AE001401 AE001401 Plasmodium chromosome 2, sectio 33 6.7
25	emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb Z98547 PFMAL3P3 Plasmodium falciparum MAL3P3, complete seque 33 9.2 emb AQ659747 AQ659747 Sheared DNA-16J5.TR Sheared DNA Trypanosom 33 9.2 gb BE021269 BE021269 sm56h04.y1 Gm-c1028 Glycine max cDNA clone 33 9.2 emb AW222457 AW222457 EST299268 tomato fruit red ripe, TAMU Lyco 33 9.2 emb AB012116 AB012116 Vigna mungo UFGlyT mRNA for UDP-glycose:fl 33 9.2
30	emb AW725836 AW725836 GA_Ea0019N24 Gossypium arboreum 7-10 dpa 33 9.2 emb AQ324451 AQ324451 mgxb0018L23r CUGI Rice Blast BAC Library P 33 9.2 gb M73492 LEIHSP90 Leishmania donovani heat shock protein 90 mRN 33 9.2 emb AW132634 AW132634 se06h07.y1 Gm-c1013 Glycine max cDNA clone 33 9.2 emb AI166186 AI166186 a032p32u Hybrid aspen plasmid library Popu 33 9.2
35	gb BE053953 BE053953 GAEa0031D23f Gossypium arboreum 7-10 dpa 33 9.2
40	Query= AtACS6_at 12891_at /id_source genbank /description gb aac63850.1  (u73786) acc synthase [arabidopsis thaliana] /blast_score 0 /ec_number /family synthase /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb atacs6  /ncgi http://www.ncgr.org/cgi-bin/ff?atacs6 (1567 letters)
45	Database: plantfungal 661,018 sequences; 426,114,510 total letters
50	Searchingdone
	Score E Sequences producing significant alignments: (bits) Value
55	emb X82273 BOACCS B.oleracea mRNA for ACC synthase. 944 0.0 emb X72676 BJMACC B. juncea mRNA for 1-Aminocyclopropane-1-carbo 833 0.0 emb AB034992 AB034992 Malus domestica MdACS-5A mRNA for 1-aminoc 725 0.0 emb AF057563 AF057563 Nicotiana glutinosa 1-aminocyclopropane-1 729 0.0
60	emb AJ005002 NTAJ5002 Nicotiana tabacum mRNA for 1-aminocyclopro 726 0.0 emb AB034993 AB034993 Malus domestica MdACS-5B mRNA for 1-aminoc 723 0.0 gb U72389 LEU72389 Lycopersicon esculentum 1-aminocyclopropane-1 724 0.0 gb U72390 LEU72390 Lycopersicon esculentum 1-aminocyclopropane-1 720 0.0

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Database: plantfungal
661,018 sequences; 426,114,510 total letters

Searching.....done

45 Score E

Sequences producing significant alignments: (bits) Value

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emblAB013346lAB013346 Lycopersicon esculentum mRNA for 1-aminocy... 704 0.0 gb|U88971|PHU88971 Pelargonium hortorum 1-aminocyclopropane-1-ca... 423 0.0 emb|Z11613|VRACCSYNM V.radiata mRNA for ACC synthase. emb|X98492|NTACCS Nicotiana tabacum mRNA for ACC-synthase (clone... 425 0.0 5 emb|X67100|GMCACCS1 G.max mRNA for ACC synthase. emb|AJ012696|CSI012696 Citrus sinensis mRNA for ACC synthase (AC... 422 0.0 emb|Z18953|PHAMCRBSY P.hybrida mRNA for 1-aminocyclopropane 1-ca... 424 0.0 emb|AJ011095|CSI011095 Citrus sinensis mRNA for ACC synthase (ac... 424 0.0 emb|Z18952|DCAMCRBSY D.caryophyllus mRNA for 1-aminocyclopropane... 686 0.0 10 emb|AB006804|AB006804 Cucumis sativus CS-ACS2 mRNA for ACC synth... 423 0.0 emb|AF080258|AF080258 Musa acuminata 1-aminocyclopropane-1-carbo... 680 0.0 emb|AF016459|AF016459 Pisum sativum 1-aminocyclopropane-1-carbox... 415 0.0 dbi|D30805|CUS1A1CS Melon mRNA for 1-aminocyclopropane-1-carboxy... 678 0.0 emb|X65982|NTXACCSYN N.tabacum mRNA for 1-aminocyclopropane-1-ca... 417 0.0 15 emb|AF109927|AF109927 Musa acuminata 1-aminocyclopropane-1-carbo... 676 0.0 emb|AB006803|AB006803 Cucumis sativus CS-ACS1 mRNA for ACC synth... 676 0.0 gb|U17231|PHU17231 Pelargonium hortorum clone pGAC-2 1-aminocycl... 423 0.0 emb|AB021906|AB021906 Musa acuminata MA-ACS1 mRNA for ACC syntha... 671 0.0 emb|AF129508|AF129508 Musa acuminata 1-aminocyclopropane-1-carbo... 671 0.0 20 emb|AB015625|AB015625 Pyrus pyrifolia pPPACS3 mRNA for 1-aminocy... 393 0.0 emb|AB031026|AB031026 Prunus mume PM-ACS1 mRNA for ACC synthase,... 405 0.0 emb[Y11357]CP1A1CS C.papaya mRNA for 1-aminocyclopropane-1-carbo... 404 0.0 dbj|D01032|CUCACCW Cucurbita maxima mRNA for 1-aminocyclopropane... 666 0.0 dbj|E03724|E03724 cDNA encoding detriment induced type 1-aminocy... 666 0.0 - 25 emb|X62536|LEACC L.esculentum mRNA for ACC synthase. 327 0.0 gb|L20634|POTACCSYN Solanum tuberosum 1-aminocyclopropane-1-carb... 403 0.0 emb|X59145|LEACC2MR Lycopersicon esculentum LE-ASCC2 mRNA (ptACC... 326 0.0 gb|M58323|CUCACCSYN Cucurbita pepo 1-aminocyclopropane-1-carboxy... 663 0.0 emb|AF057562|AF057562 Nicotiana glutinosa 1-aminocyclopropane-1-... 412 0.0 30 emb|AF239987|AF239987 Prunus persica ACC synthase ACS1 mRNA, par... 662 0.0 gb|M34289|TOMACS Tomato 1-aminocyclopropane-1-carboxylate syntha... 327 0.0 gb|M63490|TOMACCS Tomato 1-aminocyclopropane-1-carboxylate synth... 657 0.0 emb|X59146|LEACC4MR Lycopersicon esculentum LE-ACC4 mRNA (ptACC4... 656 0.0 emb|Y15739|MAACSYNTH Musa acuminata mRNA for 1-aminocyclopropane... 655 0.0 35 gb|U79999|MAU79999 Musa acuminata ACC synthase (acs3) mRNA, comp... 655 0.0 emb|AF178076|AF178076 Carica papaya 1-aminocyclopropane-1-carbox... 395 0.0 gb|U17229|PHU17229 Pelargonium hortorum clone pGAC-1 1-aminocycl... 407 0.0 emb|X82265|CAACC1 C.anuum mRNA for 1-aminocylopropane-1-carboxyl... 639 0.0 gb|U70842|STU70842 Solanum tuberosum 1-aminocyclopropane-1-carbo... 637 0.0 40 emb|AF144746|AF144746 Solanum melongena 1-aminocyclopropane-1-ca... 636 0.0 emb|AJ276295|CSI276295 Citrus sinensis partial mRNA for ACC synt... 418 0.0 emb|AJ012577|CPA012577 Carica papaya mRNA for 1-aminocyclopropan... 387 e-180 emb|AF038945|AF038945 Rumex palustris 1-aminocyclopropane-1-carb... 372 e-179 gb[U73815]MDU73815 Malus domestica ACC synthase (MdACS-2) mRNA, ... 623 e-178 45 dbj|D37937|D37937 Cucumis melo mRNA for 1-aminocyclopropane-1-ca... 380 e-177 emb|Z11562|VRACCSYN V.radiata mRNA for 1-aminocyclopropane-1-car... 375 e-176 emb|AF177769|AF177769 Carica papaya 1-aminocyclopropane-1-carbox... 369 e-175 emb|AF083814|AF083814 Antirrhinum majus ACC synthase 1 (ACS1) mR... 605 e-172 emb|AF178077|AF178077 Carica papaya 1-aminocyclopropane-1-carbox... 316 e-171 50 emb|AF239989|AF239989 Prunus persica ACC synthase ACS25 mRNA, pa... 597 e-170 gb|U22523|MIU22523 Mangifera indica 1-aminocyclopropane-carboxyl... 359 e-169 emb|X66605|DCACCS D.caryophyllus mRNA for 1-aminocyclopropane-1-... 396 e-168 emb|AF049137|AF049137 Dianthus caryophyllus 1-aminocyclopropane-... 338 e-167 emb|AF170705|AF170705 Mangifera indica 1-aminocyclopropane-1-car... 290 e-167 emb|AJ277160|CPA277160 Carica papaya partial paccs1A gene for 1-... 266 e-165 emb|AB015624|AB015624 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 359 e-163 gb|U03294|MSU03294 Malus sylvestris 1-aminocyclopropane-1-carbox... 359 e-163 gb|L31347|MAUACCSYN Malus domestica 1-aminocyclopropane-1-carbox... 357 e-162 emb|AB015495|AB015495 Passiflora edulis PE-ACS2 mRNA for ACC syn... 368 e-162 60 emb|Z77854|PSPACS1 Phalaenopsis species mRNA for 1-aminocyclopro... 514 e-161 dbj|D01033|CUCACCA Cucurbita maxima mRNA for 1-aminocyclopropane... 349 e-159

emblX87112IPCPCACS1G P.communis mRNA for 1-aminocyclopropane-1-c... 346 e-159 emb|AB007449|AB007449 Actinidia deliciosa mRNA for 1-aminocyclop... 336 e-159 gb|U73816|MDU73816 Malus domestica ACC synthase (MdACS-3) mRNA, ... 504 e-158 gb|U17972|LEU17972 Lycopersicon esculentum 1-aminocyclopropane-1... 343 e-157 5 emb|AB007639|AB007639 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 502 e-157 gb|M66619|DINCARACC D.caryophyllus 1-aminocyclopropane-1-carboxy... 495 e-156 emb|AB006805|AB006805 Cucumis sativus CS-ACS3 mRNA for ACC synth... 342 e-156 gb|U34987|VRU34987 Vigna radiata 1-aminocyclopropane-1-carboxyli... 305 e-151 emb|AB000679|AB000679 Vigna radiata mRNA for 1-aminocyclopropane... 340 e-151 10 gb|U34986|VRU34986 Vigna radiata 1-aminocyclopropane-1-carboxyli... 340 e-149 gb|U64031|DCU64031 Dendrobium crumenatum ACC synthase gene, comp... 259 e-149 emb|AF016458|AF016458 Pisum sativum 1-aminocyclopropane-1-carbox... 336 e-148 emb|Z27233|STACCAS1 S.tuberosum (STAC1) gene for amino cycloprop... 238 e-147 emb|Z27234|STACCAS2 S.tuberosum STACS2 gene for 1-Aminocycloprop... 234 e-146 15 gb|U37774|CMU37774 Cucurbita maxima 1-aminocyclopropane-1-carbox... 235 e-145 emb|AF043122|AF043122 Lycopersicon esculentum ACC synthase (LE-A... 237 e-145 emb|Z12135|VRACCSYN4 V.radiata gene for ACC synthase (pMAC-4). gb|L34171|TOMACS3A Lycopersicon esculentum 1-aminocyclopropane-1... 236 e-143 gb|U18055|LEU18055 Lycopersicon esculentum 1-aminocyclopropane-1... 236 e-143 20 emb|AF074927|AF074927 Sinapis arvensis 1-aminocyclopropane-1-car... 330 e-142 emb|AF083815|AF083815 Antirrhinum majus ACC synthase 2 (ACS2) mR... 482 e-141 emb|AF151961|AF151961 Vigna radiata 1-aminocyclopropane-1-carbox... 234 e-141 emb|AB021907|AB021907 Musa acuminata MA-ACS2 mRNA for ACC syntha... 310 e-140 gb|L07883|DORAMICARB Moth orchid 1-aminocyclopropane-1-carboxyla... 243 e-139 25 emb|AB021908|AB021908 Musa acuminata MA-ACS3 mRNA for ACC syntha... 298 e-138

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

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Score E
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emb|AW267820|AW267820 EST305948 DSIR Medicago truncatula cDNA cl... 160 1e-38
emb|AB035270|AB035270 Matricaria chamomilla McEREBP1 mRNA for et... 155 5e-37
gb|U89255|LEU89255 Lycopersicon esculentum DNA-binding protein P... 138 3e-32
emb|AI771213|AI771213 EST252409 tomato ovary, TAMU Lycopersicon ... 135 1e-31

emb|AW034216|AW034216 EST277787 tomato callus, TAMU Lycopersicon... 134 1e-30 emb|AW0729466|AW729466 GA\_Ea0025B11 Gossypium arboreum 7-10 dpa ... 132 4e-30 emb|AF057373|AF057373 Nicotiana tabacum ethylene response elemen... 125 4e-28 emb|AW981151|AW981151 EST392345 DSIL Medicago truncatula cDNA cl... 124 8e-28

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	emb[AW759236]AW759236 sl38f08.yl Gm-c1027 Glycine max cDNA clone 80 2e-14
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10	gb[T14923[T14923 crs406 lambdaZAPST Ricinus communis cDNA clone 56 9e-14
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	emb AW033743 AW033743 EST277314 tomato callus, TAMU Lycopersicon 52 le-13
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	emb AI771755 AI771755 EST252855 tomato ovary, TAMU Lycopersicon 52 1e-13
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	emb AI485460 AI485460 EST243781 tomato ovary, TAMU Lycopersicon 52 1e-13
	emb AI771795 AI771795 EST252895 tomato ovary, TAMU Lycopersicon 52 1e-13
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	5 [arabidopsis thaliana] /blast score 1.00e-117 /ec number /family
	/chip nova /gb_link /ncgi
20	(1059 letters)
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	Database: plantfungal
	661,018 sequences; 426,114,510 total letters
	Searchingdone
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	Sequences producing significant alignments: (bits) Value
	(cit), the
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	emb AI897834 AI897834 EST267277 tomato ovary, TAMU Lycopersicon 132 2e-30
	emb AW349516 AW349516 GM210007B10A12R Gm-r1021 Glycine max cDNA 130 2e
45	29
	emb AW981151 AW981151 EST392345 DSIL Medicago truncatula cDNA cl 125 2e-29
	emb AI899000 AI899000 EST268443 tomato ovary, TAMU Lycopersicon 128 3e-29
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	dbj D38125 TOBBY4C Tobacco mRNA for EREBP-4, complete cds. 127 1e-28
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55 -	emb AW775819 AW775819 EST334884 DSIL Medicago truncatula cDNA cl 95 Se-27
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	emb AW573782 AW573782 EST316373 GVN Medicago truncatula cDNA clo 93 9e-27
	emb AW620490 AW620490 sj05h02.yl Gm-c1032 Glycine max cDNA clone 121 1e-26
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UU	emb AI778693 AI778693 EST259572 tomato susceptible, Cornell Lyco 120 2e-26
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emb|AW776668|AW776668 EST335733 DSIL Medicago truncatula cDNA cl... 118 6e-26 emb|AW574222|AW574222 EST316813 GVN Medicago truncatula cDNA clo... 118 6e-26 emb|AI899889|AI899889 sb94g05.y1 Gm-c1017 Glycine max cDNA clone... 117 1e-25 emb[AW185128]AW185128 se87b10.y1 Gm-c1023 Glycine max cDNA clone... 116 2e-25 emb|AW688546|AW688546 NF008H02ST1F1000 Developing stem Medicago ... 89 2e-25 emblAV425560|AV425560 AV425560 Lotus japonicus young plants (two... 115 5e-25 emb|AV423260|AV423260 AV423260 Lotus japonicus young plants (two... 115 5e-25 emb|AI782381|AI782381 EST263260 tomato susceptible, Cornell Lyco... 115 8e-25 emb|AF245119|AF245119 Mesembryanthemum crystallinum AP2-related ... 114 1e-24 10 emb|AW781602|AW781602 s182d06.y1 Gm-c1037 Glycine max cDNA clone... 114 1e-24 emb[AW267820]AW267820 EST305948 DSIR Medicago truncatula cDNA cl... 113 2e-24 emb|AI965917|AI965917 sc79f12.y1 Gm-c1018 Glycine max cDNA clone... 113 3e-24 emb|AI484961|AI484961 EST243224 tomato ovary, TAMU Lycopersicon ... 112 5e-24 emb|AB035270|AB035270 Matricaria chamomilla McEREBP1 mRNA for et... 111 le-23 15 emb|AW220395|AW220395 EST302878 tomato root during/after fruit s... 110 2e-23 emb|AV421566|AV421566 AV421566 Lotus japonicus young plants (two... 109 3e-23 dbj|D38126|TOBBY4D Tobacco mRNA for EREBP-2, complete cds. 109 3e-23 emb|AW101306|AW101306 sd77c11.yl Gm-c1009 Glycine max cDNA clone... 89 5e-23 emb|AW349638|AW349638 GM210005B21A4R Gm-r1021 Glycine max cDNA 3... 107 2e-20 emb|AI771213|AI771213 EST252409 tomato ovary, TAMU Lycopersicon ... 107 2e-22 gb|U89255|LEU89255 Lycopersicon esculentum DNA-binding protein P... 107 2e-22 emb|AW596384|AW596384 sj02f12.y1 Gm-c1032 Glycine max cDNA clone... 107 2e-22 dbj|D38123|TOBBY4A Nicotiana tabacum mRNA for ERF1, complete cds. 105 8e-22 25 emb|AV417552|AV417552 AV417552 Lotus japonicus young plants (two... 103 2e-21 emb|AW034216|AW034216 EST277787 tomato callus, TAMU Lycopersicon... 103 2e-21 emb|AW278190|AW278190 sf40g11.yl Gm-c1009 Glycine max cDNA clone... 103 2e-21 emb|AJ238740|CRO238740 Catharanthus roseus mRNA for AP2-domain D... 102 4e-21 emb|AI973653|AI973653 sd07h05.y1 Gm-c1020 Glycine max cDNA clone... 102 4e-21 30 emb|AW690929|AW690929 NF034G08ST1F1000 Developing stem Medicago ... 73 9e-21 emb|AW233956|AW233956 sf32e02.yl Gm-c1028 Glycine max cDNA clone... 101 1e-20 emb|AW736415|AW736415 EST332429 KV3 Medicago truncatula cDNA clo... 73 2e-20 emb|AI440657|AI440657 sa63d09.yl Gm-c1004 Glycine max cDNA clone... 100 2e-20 35 emb|AV422393|AV422393 AV422393 Lotus japonicus young plants (two... 100 2e-20 emb|AI967551|AI967551 Ljirnpest05-400-d11 Ljirnp Lambda HybriZap... 100 2e-20 emb|AI731242|AI731242 BNLGHi9002 Six-day Cotton fiber Gossypium ... 100 3e-20 gb|U89256|LEU89256 Lycopersicon esculentum DNA-binding protein P... 99 5e-20 40 emb|AW030009|AW030009 EST273264 tomato callus, TAMU Lycopersicon... 99 5e-20 emb|AI894873|AI894873 EST264316 tomato callus, TAMU Lycopersicon... 99 5e-20 emb|AW030386|AW030386 EST273641 tomato callus, TAMU Lycopersicon... 99 5e-20 emb|AW729466|AW729466 GA\_Ea0025B11 Gossypium arboreum 7-10 dpa ... 98 1e-19 emb|AW507898|AW507898 si46f03.yl Gm-r1030 Glycine max cDNA clone... 98 1e-19 45 emb|AW618245|AW618245 EST314295 L. pennellii trichome, Cornell U... 98 1e-19 emb|AW840611|AW840611 00057 leafy spurge Lambda HybriZAP 2.1 two... 97 2e-19 emb|AI736796|AI736796 sb33f06.v1 Gm-c1012 Glycine max cDNA clone... 97 2e-19 emblAI966559lAI966559 sc52a04.yl Gm-c1015 Glycine max cDNA clone... 97 2e-19 emb|AF057373|AF057373 Nicotiana tabacum ethylene response elemen... 96 3e-19 50 emb|AI896308|AI896308 EST265751 tomato callus, TAMU Lycopersicon... 96 5e-19 emb|AI898992|AI898992 EST268435 tomato ovary, TAMU Lycopersicon ... 95 6e-19 emb|AW980654|AW980654 EST391807 GVN Medicago truncatula cDNA clo... 95 6e-19 emb|AI495036|AI495036 sa90a09.y1 Gm-c1004 Glycine max cDNA clone... 94 2e-18 gb|U81157|NTU81157 Nicotiana tabacum S25-XP1 DNA binding protein... 94 2e-18 emb|AW706628|AW706628 sj62g05.y1 Gm-c1033 Glycine max cDNA clone... 70 3e-18 emb|AI442716|AI442716 sa85d10.yl Gm-c1004 Glycine max cDNA clone... 93 4e-18 emb|AW559315|AW559315 EST306358 DSIR Medicago truncatula cDNA cl... 93 4e-18 emb[AW035119]AW035119 EST280381 tomato callus, TAMU Lycopersicon... 91 1e-17 emb|AI490591|AI490591 EST249145 tomato ovary, TAMU Lycopersicon ... 91 1e-17 60 emb|AI489147|AI489147 EST247486 tomato ovary, TAMU Lycopersicon ... 91 1e-17 emb|AW782252|AW782252 sm03d11.yl Gm-c1027 Glycine max cDNA clone... 63 1e-17

emb|AW348322|AW348322 GM210001B23F6R Gm-r1021 Glycine max cDNA 3... 90 2e-17 emb|AW759236|AW759236 sl38f08.yl Gm-c1027 Glycine max cDNA clone... 90 2e-17 emb|AW686013|AW686013 NF033D04NR1F1000 Nodulated root Medicago t... 90 2e-17 emb|AI776626|AI776626 EST257726 tomato resistant, Cornell Lycope... 89 4e-17 5 emb|AW443265|AW443265 EST308195 tomato mixed elicitor, BTI Lycop... 89 5e-17 emb|AW308784|AW308784 sf71h01.y1 Gm-c1013 Glycine max cDNA clone... 89 5e-17 emb|AF211541|AF211541 AF211541 34.1B Nicotiana tabacum cDNA clon... 87 2e-16 emb|AW621700|AW621700 EST312498 tomato root during/after fruit s... 87 2e-16 emb|AI489709|AI489709 EST248048 tomato ovary, TAMU Lycopersicon ... 62 2e-16 10 emb|AW560968|AW560968 EST316016 DSIR Medicago truncatula cDNA cl... 60 4e-16 emb|AW560196|AW560196 EST315244 DSIR Medicago truncatula cDNA cl... 60 4e-16 emb[AW185802]AW185802 se59h01.yl Gm-c1019 Glycine max cDNA clone... 71 5e-16 emb|AW734757|AW734757 sk88d02.yl Gm-c1035 Glycine max cDNA clone... 85 1e-15 emblAV417624|AV417624 AV417624 Lotus japonicus young plants (two... 58 1e-15 15 emb|AV407462|AV407462 AV407462 Lotus japonicus young plants (two... 58 1e-15 emb|AW256448|AW256448 EST304585 KV2 Medicago truncatula cDNA clo... 59 1e-15' emb|AW560135|AW560135 EST315183 DSIR Medicago truncatula cDNA cl... 59 2e-15 emb|AW267756|AW267756 EST305884 DSIR Medicago truncatula cDNA cl... 59 2e-15 emb|AW560134|AW560134 EST315182 DSIR Medicago truncatula cDNA cl... 59 2e-15 20 emb|AW574073|AW574073 EST316664 GVN Medicago truncatula cDNA clo... 61 2e-15 Query= AthCOR1\_s\_at 12916\_s\_at /id\_source genbank /description gb|aac13947.1| (af021244) coronatine-induced protein 1 [arabidopsis 25 thaliana] /blast score 0 /ec number /family /chip nova /gb link . (1188 letters) Database: plantfungal 30 661,018 sequences; 426,114,510 total letters Searching......done Score 35 Sequences producing significant alignments: (bits) Value emb|AF134301|AF134301 Chenopodium album CaCLH (CaCLH) mRNA, comp... 89 4e-57 emb|AB025025|AB025025 Chenopodium album mRNA for chlorophyllase.... 89 4e-57 emb]AF160869]AF160869 Citrus sinensis chlorophyllase mRNA, compl... 84 4e-54 40 emb|AW039135|AW039135 EST281108 tomato mixed elicitor, BTI Lycop... 127 2e-38 gb|BE037470|BE037470 MP21E05 MP Mesembryanthemum crystallinum cD... 81 4e-35 emb|AW093776|AW093776 EST286956 tomato mixed elicitor, BTI Lycop... 59 4e-34 emb|AW616099|AW616099 EST296862 L. hirsutum trichome, Cornell Un... 67 1e-29 emb|AW266073|AW266073 L30-2749T3 Ice plant Lambda Uni-Zap XR exp... 79 3e-29 45 emb|AA887366|AA887366 L30-338T3 Ice plant Lambda Uni-Zap XR expr... 79 2e-26 emb|AA556157|AA556157 12 Loblolly pine N Pinus taeda cDNA clone ... 120 3e-26 emb|AW042994|AW042994 ST27H11 Pine TriplEx shoot tip library Pin... 106 7e-26 emb|AW265807|AW265807 L30-2522 T3 Ice plant Lambda Uni-Zap XR ex... 84 2e-21 emb|AW678075|AW678075 WS1\_13\_D09.b1\_A002 Water-stressed I (WS1) ... 81\_1e-19 50 emb|AW133452|AW133452 se19b10.yl Gm-c1015 Glycine max cDNA clone... 62 1e-17 emb|AI777679|AI777679 EST258474 tomato susceptible, Cornell Lyco... 53 7e-17 emb|AW596770|AW596770 sj16e05.yl Gm-c1032 Glycine max cDNA clone... 89 9e-17 emb|AW596245|AW596245 sj01a09.y1 Gm-c1032 Glycine max cDNA clone... 87 3e-16 emb|AI771244|AI771244 EST252260 tomato ovary, TAMU Lycopersicon ... 67 3e-16 55 emb|AW831810|AW831810 sm36f12.yl Gm-c1028 Glycine max cDNA clone... 85 8e-16

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gb|BE058853|BE058853 sn21g06.y1 Gm-c1016 Glycine max cDNA clone ... 41 0.019 emb|AW773701|AW773701 EST332687 KV3 Medicago truncatula cDNA clo... 39 0.094

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	emb AV428585 AV428585 AV428585 Lotus japonicus young plants (two 37 0.33
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	gb L33647 L33647 BNAESTF649 Mustard flower buds Brassica rapa cD 35 0.86
	emb AW455247 AW455247 EST311907 tomato root during/after fruit s 35 0.86
15	emb AQ909224 AQ909224 GSSTc07661 Trypanosome cruzi random genomi 35 0.86
15	embly 00002221A 0000222 GSST-07660 Trypanosome cruzi random genomi 35 0.80
	emb AQ909223 AQ909223 GSSTc07660 Trypanosome cruzi random genomi 35 0.86
	emb AV421772 AV421772 AV421772 Lotus japonicus young plants (two 35 0.86
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	emb AW625056 AW625056 EST313873 tomato radicle, 5 d post-imbibit 35 0.86
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	emb AW615881 AW615881 EST325379 tomato flower buds 0-3 mm, Corne 35 1.2
30	emb AV410945 AV410945 AV410945 Lotus japonicus young plants (two 35 1.2
30	emb AW616956 AW616956 EST323367 L. hirsutum trichome, Cornell Un 35 1.6
	emb AW626060 AW626060 EST319967 tomato radicle, 5 d post-imbibit 35 1.6
	emb AI055068 AI055068 coau0002P04 Cotton Boll Abscission Zone cD 35 1.6
	emb AJ242531 TAE242531 Triticum aestivum rht-D1a gene for gibber 35 1.6
~ -	emb AW926582 AW926582 HVSMEg0007J16 Hordeum vulgare pre-anthesis 27 1.9
35	emb AC067936 AC067936 Neurospora crassa chromosome 7, clone X3-B 34 2.2
	emb AW649885 AW649885 EST328339 tomato germinating seedlings, TA 34 2.2
	emb AW092862 AW092862 EST286042 tomato mixed elicitor, BTI Lycop 34 2.2
	emb AW037417 AW037417 EST278919 tomato mixed elicitor, BTI Lycop 34 2.2
	emb AI771746 AI771746 EST252846 tomato ovary, TAMU Lycopersicon 34 2.2
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post/entrez/query?db=n&form=6&dopt=g&uid=gb|atthired4|/ncgi http://www.ncgr.org/cgi-bin/ff?atthired4 (528 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

Searching......done

Score E

25 Sequences producing significant alignments:

(bits) Value

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      gb|M16936|COTSPB G.hirsutum (cotton) storage protein (beta-globu... 92 2e-18
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      emb|Y16976|CAR16976 Coffea arabica mRNA for 11S storage protein.
      gb|M28832|HNNHAG3D1S Sunflower 11S storage protein (G3-D1) DNA, ... 91 4e-18
      emb|AW036288|AW036288 EST278367 tomato seed, TAMU Lycopersicon e... 90 7e-18
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      gb|M36407|CUC11SGB Pumpkin 11-S globulin beta-subunit mRNA, comp... 89 2e-17
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      emb|Y09116|SSPPLEGUM S.sagittifolia mRNA for pre-pro-legumin.
      emb|X82465|MSLEGB14 M.salicifolia mRNA for legumin precursor (B14). 89 2e-17
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	emb AF216801 AF216801 Fagopyrum esculentum allergenic protein ge 87 6e-17
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	gb M69188 COTDGALA Cotton legumin A D-genome alloallele gene, co 86 1e-16
	gb M16905 COTSPD G.hirsutum (cotton) storage protein (beta-globu 86 1e-16
	emb[X74741]ASGLAV3G A.sativa GLAV 3 gene for 11S globulin. 86 1e-16
	emb AF091842 AF091842 Sesamum indicum strain Tainan 1 11S globul 86 2e-16
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-	emb X17637 AS12SSP Oat gene for 12S seed storage protein. 85 2e-16
	gb T24158 T24158 crs141 lambdaZAPST Ricinus communis cDNA clone 85 3e-16
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	emb AW318315 AW318315 sg63f06.yl Gm-c1007 Glycine max cDNA clone 84 4e-16
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55	emb Y09117 SSPPLTAND S.sagittifolia mRNA for pre-pro-legumin and 84 8e-16
	citio 1 0 11 7 3 5 F F L 1 AND 3. sagituio ila micha for pre-pro-legumin and 84 8e-10
	emb AI941031 AI941031 sb83e04.yl Gm-c1010 Glycine max cDNA clone 83 1e-15
	emb AI748075 AI748075 sb47h08.yl Gm-c1011 Glycine max cDNA clone 83 1e-15
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emb|AW625527|AW625527 EST319434 tomato radicle, 5 d post-imbibit... 133 8e-53 emb|AW218547|AW218547 EST303730 tomato radicle, 5 d post-imbibit... 133 8e-53 emb|AW733300|AW733300 sk71f10.yl Gm-c1016 Glycine max cDNA clone... 134 1e-52 emb|AW442463|AW442463 EST307393 tomato mixed elicitor, BTI Lycop... 133 1e-52 5 emb|AW094365|AW094365 EST287545 tomato mixed elicitor, BTI Lycop... 133 1e-52 gb|BE021597|BE021597 sm60a04.yl Gm-c1028 Glycine max cDNA clone ... 135 1e-52 emb|AW034347|AW034347 EST277918 tomato callus, TAMU Lycopersicon... 133 2e-52 emb|AI443130|AI443130 sa84g01.y1 Gm-c1004 Glycine max cDNA clone... 131 2e-51 emb|AI898952|AI898952 EST268395 tomato ovary, TAMU Lycopersicon ... 139 2e-51 10 emb|AW424002|AW424002 sh59c10.yl Gm-c1015 Glycine max cDNA clone... 115 2e-51 emb|AW707192|AW707192 sk22a02.y1 Gm-c1028 Glycine max cDNA clone... 132 3e-50 emb|AI899227|AI899227 EST268670 tomato ovary, TAMU Lycopersicon ... 139 3e-50 emb|AW980337|AW980337 EST391490 GVN Medicago truncatula cDNA clo... 127 3e-50 emb|AW648499|AW648499 EST326953 tomato germinating seedlings, TA... 99 7e-50 15 emb|AW690185|AW690185 NF029D09ST1F1000 Developing stem Medicago ... 142 1e-49 emb|AI895091|AI895091 EST264534 tomato callus, TAMU Lycopersicon... 99 3e-49 emb|AW306980|AW306980 sf51c05.yl Gm-c1009 Glycine max cDNA clone... 135 8e-48 emb|AW695176|AW695176 NF092D12ST1F1101 Developing stem Medicago ... 142 1e-47 emb[AW684885]AW684885 NF022F07NR1F1000 Nodulated root Medicago t... 142 2e-47 20 emb|AA660318|AA660318 00189 MtRHE Medicago truncatula cDNA 5' si... 144 2e-47 emb|AW703717|AW703717 sk23e07.yl Gm-c1028 Glycine max cDNA clone... 132 4e-47 emb|AW776435|AW776435 EST335500 DSIL Medicago truncatula cDNA cl... 142 5e-47 emb|AI486948|AI486948 EST245270 tomato ovary, TAMU Lycopersicon ... 100 1e-46 emb|AW620537|AW620537 sj06d09.yl Gm-c1032 Glycine max cDNA clone... 133 1e-46 25 emb|AI775483|AI775483 EST256583 tomato resistant, Cornell Lycope... 139 2e-46 gb|BE124333|BE124333 EST393368 GVN Medicago truncatula cDNA clon... 125 3e-46 emb|AW704578|AW704578 sk38h08.y1 Gm-c1028 Glycine max cDNA clone... 140 3e-46 emb|AW350997|AW350997 GM210010A10C11R Gm-r1021 Glycine max cDNA ... 97 6e-46 emb|AW707224|AW707224 sk22c12.yl Gm-c1028 Glycine max cDNA clone... 140 9e-46 30 emb|AW625126|AW625126 EST313943 tomato radicle, 5 d post-imbibit... 133 6e-45 emb|AI974778|AI974778 T113251e KV2 Medicago truncatula cDNA clon... 142 6e-45 emb|AI898101|AI898101 EST267544 tomato ovary, TAMU Lycopersicon ... 98 8e-45 gb|BE124175|BE124175 EST394300 DSIL Medicago truncatula cDNA clo... 127 1e-44 emb]AW686370]AW686370 NF040G10NR1F1000 Nodulated root Medicago t... 127 4e-44 35 emb|AI166314|AI166314 xylem.est.159 Poplar xylem Lambda ZAPII li... 136 4e-44 emb|AW256836|AW256836 EST304973 KV2 Medicago truncatula cDNA clo... 144 5e-44 emb|AW559274|AW559274 EST306110 DSIR Medicago truncatula cDNA cl... 142 5e-44 emb|AW278618|AW278618 sf46h04.y1 Gm-c1009 Glycine max cDNA clone... 139 1e-43 gb|BE059325|BE059325 sn31c09.yl Gm-c1016 Glycine max cDNA clone ... 140 3e-43 40 emb|AW831448|AW831448 sm23c03.y1 Gm-c1028 Glycine max cDNA clone... 140 6e-43 emb|AI938893|AI938893 sc62d08.y1 Gm-c1016.Glycine max cDNA clone... 134 2e-42 emb|AW257069|AW257069 EST305206 KV2 Medicago truncatula cDNA clo... 142 2e-42 emb|AW695899|AW695899 NF100E02ST1F1017 Developing stem Medicago ... 125 5e-42 emb|AW774697|AW774697 EST333848 KV3 Medicago truncatula cDNA clo... 84 8e-42 45 emb|AW683492|AW683492 NF012H09LF1F1079 Developing leaf Medicago ... 134 2e-41 emb|AW152892|AW152892 se32d04.y1 Gm-c1015 Glycine max cDNA clone... 82 2e-41 gb|BE125749|BE125749 DG1 55 H11.b1 A002 Dark Grown 1 (DG1) Sorgh... 127 2e-41 gb|BE022002|BE022002 sm65c10.yl Gm-c1028 Glycine max cDNA clone ... 137 3e-41 emb|AI488060|AI488060 EST246382 tomato ovary, TAMU Lycopersicon ... 100 7e-41 50 emb|AW622558|AW622558 EST313358 tomato root during/after fruit s... 139 1e-40 emb|AW625461|AW625461 EST319368 tomato radicle, 5 d post-imbibit... 99 1e-40 emb|AW625476|AW625476 EST319383 tomato radicle, 5 d post-imbibit... 139 3e-40 emb|Al960196|Al960196 sc80a08.yl Gm-c1018 Glycine max cDNA clone... 132 5e-40 emb|Z82982|NTZ82982 N.tabacum mRNA for caffeoyl-CoA O-methyltran... 136 1e-39 55 ... emb|AW218486|AW218486 EST303669 tomato radicle, 5 d post-imbibit... 136 1e-39

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(827 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters 5 Searching.....done E Score Sequences producing significant alignments: (bits) Value 10 emb|AI489346|AI489346 EST247685 tomato ovary, TAMU Lycopersicon ... 79 2e-28 emb|AW350323|AW350323 GM210007B20E12R Gm-r1021 Glycine max cDNA ... 80 2e-27 emb|AW509006|AW509006 si38h07.yl Gm-r1030 Glycine max cDNA clone... 79 2e-26 emb|AW685404|AW685404 NF028H10NR1F1000 Nodulated root Medicago t... 74 2e-24 15 emb|AW781320|AW781320 sk68d07.y1 Gm-c1016 Glycine max cDNA clone... 86 2e-24 emb|AI897832|AI897832 EST267275 tomato ovary, TAMU Lycopersicon ... 78 1e-23 emb|AW102460|AW102460 sd88d10.y1 Gm-c1009 Glycine max cDNA clone... 70 2e-22 emb|AI490284|AI490284 EST248610 tomato ovary, TAMU Lycopersicon ... 103 2e-22 emb|AI487362|AI487362 EST245684 tomato ovary, TAMU Lycopersicon ... 72 3e-20 20 gb|BE125690|BE125690 DG1\_54\_A02.g1\_A002 Dark Grown 1 (DG1) Sorgh... 96 3e-19 emb|X80231|LTCAMA L.tarentolae CAM A gene for calmodulin. 73 le-18 emb|AL115248|CNS01C9K Botrytis cinerea strain T4 cDNA library un... 65 4e-18 dbilD10521|TETCALW T.pyriformis mRNA for calmodulin. 73 7e-18 gb|M76407|SLECALMODU Stylonychia lemnae calmodulin gene, complet... 74 9e-18 25 emb|AF007889|AF007889 Symbiodinium microadriaticum calmodulin (S... 74 1e-17 emb|AF078679|AF078679 Olea europaea calcium-binding protein (PCA... 57 2e-17 emb|X56511|TBCALUBG T. brucei genes for calmodulin, EFH5 and ubi... 77 2e-17 emb|X52096|TCCALB2 Trypanosoma cruzi CalA2 calmodulin gene. emb|AF030033|AF030033 Phaseolus vulgaris calmodulin (CaM) mRNA, ... 75 2e-17 30 emb|X90560|PPCAMPROT Physcomitrella patens mRNA for calmodulin. gb|U91642|POU91642 Pleurotus ostreatus calmodulin mRNA, complete... 75 2e-17 emblAF078680|AF078680 Olea europaea calcium-binding protein (PCA... 57 2e-17 emb|X85091|MPCAM M.pyrifera mRNA for calmodulin. 74 2e-17 gb|M83535|PHTCALPIA P.infestans calmodulin (calA) gene, complete... 75 2e-17 35 emb|Y08373|TGCM T.gondii mRNA for calmodulin. 74 2e-17 gb|C96396|C96396 C96396 Marchantia polymorpha immature sex organ... 73 3e-17 emb|AL113315|CNS01ARV Botrytis cinerea strain T4 cDNA library un... 72 3e-17 emb|AB044286|AB044286 Chara corallina ccam mRNA for calmodulin, ... 75 4e-17 emb|AB041712|AB041712 Chara corallina cccam2 mRNA for calmodulin... 75 4e-17 40 emb|AB041711|AB041711 Chara corallina cccam1 mRNA for calmodulin... 75 4e-17 gblJ05116|ACKCAL A.klebsiana calmodulin gene, complete cds. 75 4e-17 emb|X70923|NCCALMOE N.crassa mRNA for calmodulin. 74 6e-17 emb|AI328739|AI328739 a6d02ne.fl Neurospora crassa evening cDNA ... 74 6e-17 emb|AF034964|AF034964 Glomerella cingulata calmodulin (cam) mRNA... 74 6e-17 45 gb|U12505|HCCMD Histoplasma capsulatum 186AS calmodulin mRNA, co... 74 8e-17 gb[M34540]PARCAM P. tetraurelia calmodulin gene, complete cds. gb|S68025|S68025 CAM=calmodulin [Paramecium tetraurelia, Genomic... 74 8e-17 emb|AW164773|AW164773 se77e12.yl Gm-c1023 Glycine max cDNA clone... 64 8e-17 emb|AW719875|AW719875 LjNEST11d4r Lotus japonicus nodule library... 57 8e-17 50 emb|AW830090|AW830090 sm22a12.yl Gm-c1028 Glycine max cDNA clone... 64 8e-17 emb|AW126204|AW126204 N100049e rootphos(-) Medicago truncatula c... 57 8e-17 emb|X52242|TTCALM T.thermophila mRNA for calmodulin. gb|L02963|NEUCLMDLN Neurospora crassa calmodulin mRNA, complete ... 74 1e-16 emb|AW625406|AW625406 EST319229 tomato radicle, 5 d post-imbibit... 62 1e-16 55 gb[K02944]TRBCMRSG Trypanosoma brucei gambiense calmodulin genes... 73 2e-16 gb|M88307|BNACALM Brassica juncea calmodulin mRNA, complete cds. 73 2e-16 gb|U10150|BNU10150 Brassica napus Naehan calmodulin (bcm1) mRNA,... 73 2e-16 emb|X89890|BPCALMGEN B.pilosa mRNA for calmodulin. 73 2e-16 emb|AW099396|AW099396 sd39h01.yl Gm-c1016 Glycine max cDNA clone... 76 2e-16 60 emb|AW101324|AW101324 sd77e09.y1 Gm-c1009 Glycine max cDNA clone... 59 2e-16 emb|AL112713|CNS01AB5 Botrytis cinerea strain T4 cDNA library un... 73 2e-16

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       emblAW573768|AW573768 EST316359 GVN Medicago truncatula cDNA clo... 72 2e-16
       emb|Y13784|MSCALMODU Mougeotia scalaris mRNA for calmodulin.
      emb|AI727960|AI727960 BNLGHi9833 Six-day Cotton fiber Gossypium ...
                                                                         73 3e-16
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      emb|AW164628|AW164628 se74e01.y1 Gm-c1023 Glycine max cDNA clone... 60 3e-16
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      emblAW738989|AW738989 gb23c06.yl Moss EST library PPN Physcomitr... 73 3e-16
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      gb|S81594|S81594 auxin-regulated calmodulin [Vigna radiata=mung ... 73 4e-16
      gb|L20691|VIRCALMOD Vigna radiata calmodulin mRNA, complete cds.
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      gb|BE036340|BE036340 MO23E04 MO Mesembryanthemum crystallinum cD... 73 4e-16
      emb|AF030032|AF030032 Phaseolus vulgaris calmodulin (CaM) mRNA, ... 73 4e-16
      gb|L01431|SOYSCAM2X Soybean calmodulin (SCaM-2) mRNA, complete cds. 73 4e-16
      emb|Y09853|CACAM Cicer arietinum mRNA for CaM protein.
      gb|L01432|SOYSCAM3X Soybean calmodulin (SCaM-3) mRNA, complete cds. 73 4e-16
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      gb|U49104|TAU49104 Triticum aestivum calmodulin TaCaM3-3 mRNA, c... 73 4e-16
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Score E

Sequences producing significant alignments:

(bits) Value

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5	emb AW289567 AW289567 NXNV002G05F Nsf Xylem Normal wood Vertical 98 5e-20
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	emb X70688 NTHSP18P N.tabacum hsp18p mRNA for heat shock protein 61 8e-20
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•	emb[X53852]DCHSP179 Carrot gene for a heat shock protein. 61 1e-18
	emb AW223810 AW223810 EST300621 tomato fruit red ripe, TAMU Lyco 57 1e-18
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55	gb U55859 TAU55859 Triticum aestivum heat shock protein 80 mRNA, 575 0.0
	emb X98582 TAX98582 T.aestivum mRNA for HSP80-2 protein. 572 0.0
	A 100 a a a a a 10 annua a a a a a a a a a a a a a a a a a
	emb X63195 NTHSP82 N.tabacum mRNA for heat shock protein 82. 601 0.0
	emb AF123259 AF123259 Lycopersicon esculentum heat shock protein 483 0.0
60	emb AF165818 AF165818 Guillardia theta nucleomorph 5S ribosomal 505 0.0
60	emb AF042329 AF042329 Eimeria tenella heat shock protein 90 (hsp 514 0.0
	gb M57386 THEHSP90 T.parva heat shock protein 90 (hsp90) mRNA, c 500 0.0

emb|Z29667|PFHESHPR P.falciparum (7) mRNA for heat-shock protein. gb|U45449|EBU45449 Eimeria bovis heat shock protein 90 (hsp90) m... 495 0.0 emb|AF151114|AF151114 Tetrahymena thermophila strain B2086 hsp82... 486 0.0 emb|AF136649|AF136649 Babesia bovis heat shock protein 90 (HSP90... 487 0.0 gb]M15346|TRBHSC T.cruzi tandemly repeated gene encoding an 85 k... 457 0.0 5 emb[X14176]TBHSP83 Trypanosoma brucei HSP83 gene. gb|M73492|LEIHSP90 Leishmania donovani heat shock protein 90 mRN... 456 0.0 emb|X87770|LIHSP83GN L.infantum hsp83 gene. 454 0.0 gblM92926|LEIHSP01 Leishmania amazonensis heat shock protein 83 ... 449 0.0 10 gb|U92465|AFU92465 Aspergillus fumigatus heat shock protein (Hsp... 458 e-165 emb|AF212996|AF212996 Neurospora crassa heat shock protein 80 ge... 473 e-165 emb|AL110469|SPAC926 S.pombe chromosome I cosmid c926. gb|L35550|YSPHSP90X Schizosaccharomyces pombe heat shock protein... 460 e-162 gb|U81165|PAU81165 Podospora anserina suppressor of vegetative i... 292 e-161 15 emb|Z67751|SC38KCXVI S.cerevisiae DNA (chromosome XVI; 38 kb). emb|Z73596|SCYPL240C S.cerevisiae chromosome XVI reading frame O... 448 e-159 gb[K01387|YSCHSP82A Yeast (S.cerevisiae) HSP82 (heat shock-induc... 448 e-159 emb|X81025|CAHSP90 C.albicans hsp90 gene. 451 e-159 emb|AF251005|AF251005 Candida tropicalis heat shock protein 90 g... 444 e-158 emb|Z49808|SC8010 S.cerevisiae chromosome XIII cosmid 8010. emb|AF221856|AF221856 Euphorbia esula heat-shock protein 80 mRNA... 292 e-158 gb[M26044]YSCHSC82 S. cerevisiae HSC82 gene encoding hsc82, the ... 447 e-157 25 emb|AI489832|AI489832 EST248171 tomato ovary, TAMU Lycopersicon ... 471 e-132 emb|AW982575|AW982575 HVSMEg0003K07f Hordeum vulgare pre-anthesi... 471 e-131 emb|AW774793|AW774793 EST333944 KV3 Medicago truncatula cDNA clo... 459 e-128 emb|AW221746|AW221746 EST298557 tomato fruit red ripe, TAMU Lyco... 350 e-128 30 gb|BE035845|BE035845 MO11A12 MO Mesembryanthemum crystallinum cD... 455 e-127 emb|AW650299|AW650299 EST328753 tomato germinating seedlings, TA... 455 e-127 emb|AW982497|AW982497 HVSMEg0003G20f Hordeum vulgare pre-anthesi... 439 e-126 emb|AF078070|AF078070 Griffithsia japonica heat-shock protein 90... 245 e-126 emb|AW218188|AW218188 EST303369 tomato radicle, 5 d post-imbibit... 452 e-126 emb|AW731352|AW731352 GA\_\_Ea0030E22 Gossypium arboreum 7-10 dpa ... 449 e-125 35 emb|AW621485|AW621485 EST312283 tomato root during/after fruit s... 445 e-124 emb|AW905909|AW905909 EST342082 potato stolon, Cornell Universit... 331 e-124 emb|AW618647|AW618647 EST320633 L. pennellii trichome, Cornell U... 439 e-122 emb|AI730385|AI730385 BNLGHi6770 Six-day Cotton fiber Gossypium ... 439 e-122 40 emb|AW931002|AW931002 EST356845 tomato fruit mature green, TAMU ... 436 e-121 emb|AW186517|AW186517 se68g12.y1 Gm-c1019 Glycine max cDNA clone... 432 e-120 gb|BE055051|BE055051 GA\_Ea0031H10f Gossypium arboreum 7-10 dpa ... 381 e-118 emb|AW217359|AW217359 EST296010 tomato flower buds 0-3 mm, Corne... 427 e-118 emb|AW216642|AW216642 EST295356 tomato callus, TAMU Lycopersicon... 423 e-117 45 emb|AW217696|AW217696 EST296410 tomato flower buds 8 mm to pre-a... 422 e-117 emb|AW011081|AW011081 ST16E03 Pine TriplEx shoot tip library Pin... 421 e-116 emb|AI730419|AI730419 BNLGHi7100 Six-day Cotton fiber Gossypium ... 417 e-115 emb|AW907064|AW907064 EST343096 potato stolon, Cornell Universit... 413 e-114 emb|AW696590|AW696590 NF108G11ST1F1087 Developing stem Medicago ... 413 e-114 50 emb|AW738699|AW738699 EST340126 tomato flower buds, anthesis, Co... 412 e-114 emb|AI442468|AI442468 sa27d08.yl Gm-c1004 Glycine max cDNA clone... 412 e-114 emb|AW684853|AW684853 NF022C08NR1F1000 Nodulated root Medicago t... 403 e-114 emb|AW693965|AW693965 NF071A06ST1F1040 Developing stem Medicago ... 408 e-113 emb|AW690869|AW690869 NF036A07ST1F1000 Developing stem Medicago ... 390 e-113 emb|AW030608|AW030608 EST273863 tomato callus, TAMU Lycopersicon... 407 e-112 emb|AW983321|AW983321 HVSMEg0010D10f Hordeum vulgare pre-anthesi... 354 e-112 emb|AI726426|AI726426 BNLGHi5602 Six-day Cotton fiber Gossypium ... 407 e-112 emb|AW930252|AW930252 EST340709 tomato fruit mature green, TAMU ... 406 e-112 emb|AW979717|AW979717 EST341337 tomato root deficiency, Cornell ... 404 e-111 60 emb|AW695561|AW695561 NF096B09ST1F1076 Developing stem Medicago ... 404 e-111 emb|AW691505|AW691505 NF045G02ST1F1000 Developing stem Medicago ... 361 e-111

emb|AW735801|AW735801 EST336569 tomato flower buds 0-3 mm, Corne... 402 e-111 emb|AW737406|AW737406 EST338749 tomato flower buds, anthesis, Co... 402 e-111 emb|AL112936|CNS01AHC Botrytis cinerea strain T4 cDNA library un... 400 e-110 emb|AW217697|AW217697 EST296411 tomato flower buds 8 mm to pre-a... 400 e-110 5 emb|AW218948|AW218948 EST301430 tomato root during/after fruit s... 396 e-110 emb|AW220869|AW220869 EST297338 tomato fruit mature green, TAMU ... 398 e-109 emb|AW616417|AW616417 EST322828 L. hirsutum trichome, Cornell Un... 397 e-109 emb|AW688548|AW688548 NF008H04ST1F1000 Developing stem Medicago ... 396 e-109 emb|AW738535|AW738535 EST339962 tomato flower buds, anthesis, Co... 242 e-109 10 emb|AW649397|AW649397 EST327851 tomato germinating seedlings, TA... 394 e-108 emblAW693115|AW693115 NF060C10ST1F1081 Developing stem Medicago ... 226 e-107 emb|AI488065|AI488065 EST246387 tomato ovary, TAMU Lycopersicon ... 390 e-107 emb|AI726157|AI726157 BNLGHi5098 Six-day Cotton fiber Gossypium ... 226 e-107 gb|BE060856|BE060856 HVSMEg0013J20f Hordeum vulgare pre-anthesis... 388 e-107 emb|AI781620|AI781620 EST262499 tomato susceptible, Cornell Lyco... 389 e-107 emb|AL111751|CNS019KF Botrytis cinerea strain T4 cDNA library un... 389 e-107 emb|AW092845|AW092845 EST286025 tomato mixed elicitor, BTI Lycop... 386 e-106 emb|AW983154|AW983154 HVSMEg0008G17f Hordeum vulgare pre-anthesi... 224 e-106 emb|AI780272|AI780272 EST261151 tomato susceptible, Cornell Lyco... 215 e-106 20 emb|AW651194|AW651194 EST329648 tomato germinating seedlings, TA... 266 e-106 emb|AW690804|AW690804 NF035B11ST1F1000 Developing stem Medicago ... 375 e-106 emb|AW154860|AW154860 EST290253 tomato root deficiency, Cornell ... 385 e-105 emb|AI780075|AI780075 EST260954 tomato susceptible, Cornell Lyco... 383 e-105 emb|AW928471|AW928471 EST337259 tomato flower buds 8 mm to pre-a... 382 e-105 25 gb|L34028|PFAHSP86B Plasmodium falciparum (clone HB3) heat shock... 382 e-105 emblAF030694|AF030694 Plasmodium falciparum strain Dd2 heat shoc... 382 e-105 gb|L34027|PFAHSP86A Plasmodium falciparum (clone Dd2) heat shock... 382 e-105 emb|AW737397|AW737397 EST338740 tomato flower buds, anthesis, Co... 381 e-104 emb|AW334905|AW334905 S40F8 AGS-1 Pneumocystis carinii f. sp. ca... 381 e-104 30 emb|AW035244|AW035244 EST280506 tomato callus, TAMU Lycopersicon... 380 e-104 Query= Mlolike2\_s\_at 13685\_s\_at /id\_source genbank /description gb|aad25552.1|ac005850\_9 (ac005850) highly similar to mlo proteins 35 [arabidopsis thaliana] /blast\_score 0 /ec number /family /chip nova /gb\_link /ncgi (1752 letters) Database: plantfungal 40 661,018 sequences; 426,114,510 total letters Searching.....done Score 45 Sequences producing significant alignments: (bits) Value emb|A92838|A92838 Sequence 12 from Patent WO9804586. 258 e-123 emb|Z83834|HVMLO H.vulgare mRNA for Mlo protein. 261 e-116 emb|A92833|A92833 Sequence 7 from Patent WO9804586. 261 e-116 50 emb|A92828|A92828 Sequence 2 from Patent WO9804586. 261 e-116 emb|AW216578|AW216578 EST295292 tomato callus, TAMU Lycopersicon... 165 6e-78 emb|AJ005341|LUAJ5341 Linum usitatissimum mRNA for MLO-like prot... 287 2e-76 emblAW934153|AW934153 EST359996 tomato fruit mature green, TAMU ... 235 4e-74 emb|AW132264|AW132264 sd98f11.yl Gm-c1013 Glycine max cDNA clone... 128 2e-60 55 emb|AI779924|AI779924 EST260803 tomato susceptible, Cornell Lyco... 228 8e-59 gb|BE020055|BE020055 sm38e01.y1 Gm-c1028 Glycine max cDNA clone ... 199 2e-58 emb|AW132268|AW132268 sd98g11.yl Gm-c1013 Glycine max cDNA clone... 124 5e-58 emb|AI729603|AI729603 BNLGHi13760 Six-day Cotton fiber Gossypium... 202 8e-51 emb|AI054629|AI054629 coau0001J02 Cotton Boll Abscission Zone cD... 133 2e-44

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	emb AJ242498 CCL242498 Candida cloacae mRNA for long chain fatty 35 1.8
<b>55</b> '	emb AQ949106 AQ949106 Sheared DNA-49L9.TR Sheared DNA Trypanosom 35 2.5
	emb AQ650344 AQ650344 Sheared DNA-28J12.TF Sheared DNA Trypanoso 35 2.5
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	embla 0445280 0445280 GSSTc01558 Transposoms crizi random genomi 34.3.5

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10	emb AC013353 AC013353 Trypanosoma brucei chromosome VI clone RPC 34 4.7 emb AQ948416 AQ948416 Sheared DNA-38M22.TR Sheared DNA Trypanoso 34 4.7
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••	emb AZ2123/3 AZ2123/3 Sheared DNA-101E1.TF Sheared DNA Trypanoso 34 4.7 emb AJ250726 TBR250726 Trypanosoma brucei HSP100 gene, GPI-PLC g 34 4.7 gb M17420 YSTHEP Saccharomyces cerevisiae heptapeptide repeat re 33 6.5 emb AI730517 AI730517 BNLGHi6942 Six-day Cotton fiber Gossypium 33 6.5
20	emb Z49511 SCYJR011C S.cerevisiae chromosome X reading frame ORF 33 6.5 gb U59312 HVU59312 Hordeum vulgare (1,4)-beta-xylan endohydrolas 33 6.5 emb Z74188 SCYDL140C S.cerevisiae chromosome IV reading frame OR 33 6.5
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	emb AI563168 AI563168 EST00292 watermelon lambda zap library Cit 47 le-04 emb AI491014 AI491014 EST241723 tomato shoot, Cornell Lycopersic 44 0.001 emb AW034769 AW034769 EST278805 tomato callus, TAMU Lycopersicon 44 0.001
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60	emb AL109815 PFMAL13PA Plasmodium falciparum chromosome 13 strai 33 2.0 emb AW257048 AW257048 EST305185 KV2 Medicago truncatula cDNA clo 32 3.8 gb U52151 APU52151 Aspergillus parasiticus polyketide synthase P 32 3.8 emb AC644937 AC644937 PROCESS

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5	gb M60208 YSPHSP70 S.pombe mitochondrial heat shock protein (HSP 32 5.2 emb AF083468 AF083468 Emericella nidulans putative zinc finger p 32 5.2 emb AU011047 AU011047 AU011047 Schizosaccharomyces pombe late lo 32 5.2
	emb AL136235 SPAC664 S.pombe chromosome I cosmid c664. 32 5.2 emb AL160493 LMFLCHR26 Leishmania major Friedlin assembled chrom 31 7.1
10	emb X70810 CLEGCGA Euglena gracilis chloroplast complete genome. 31 7.1 emb AW923451 AW923451 DG1_52_H09.b1_A002 Dark Grown 1 (DG1) Sorg 31 7. emb AQ845394 AQ845394 LMAJFV1_lm24h05.y1 Leishmania major FV1 ra 31 7.1
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	Database: plantfungal 661,018 sequences; 426,114,510 total letters
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•	Score E Sequences producing significant alignments: (bits) Value
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	emb AL115822 CNS01CPI Botrytis cinerea strain T4 cDNA library un 33 1.5 emb AL031744 PFMAL1P1 Plasmodium falciparum chromosome 1 strain 33 2.0 emb AW033792 AW033792 EST277363 tomato callus, TAMU Lycopersicon 33 2.5
50	emb Z49705 SC8520X S.cerevisiae chromosome XIII cosmid 8520. 32 3.8 emb AB001684 AB001684 Chlorella vulgaris C-27 chloroplast DNA, c 32 3.8
	emb AL049185 PFMAL13P2 Plasmodium falciparum chromosome 13 strai 32 3.8 emb AW101318 AW101318 sd77e03.yl Gm-c1009 Glycine max cDNA clone 32 3.8 emb AQ909718 AQ909718 GSSTc09185 Trypanosome cruzi random genomi 31 5.3
55	emb AW991095 AW991095 SsS0129 Suaeda salsa ZAP cDNA library Suae 28 5.9 emb AJ273065 AJ273065 AJ273065 Metarhizium anisopliae ARSEF 2575 31 6.5
	emb AF057379 AF057379 Nicotiana tabacum clone G8-1 unknown mRNA. 31 6.5 emb AB005455 AB005455 Antirrhinum majus DNA, genomic survey sequ 31 7.3 gb BE036743 BE036743 MP04G02 MP Mesembryanthemum crystallinum cD 31 7.3
50	gb BE036743 BE036743 MP04G02 MP Mesembryanthemum crystallinum cD 31 7.3 emb AI460510 AI460510 sa80h11.yl Gm-c1004 Glycine max cDNA clone 31 7.3 emb AI026500 AI026500 TENU0712 T. cruzi epimastigote normalized 31 7.3

emb|AW651081|AW651081 EST329535 tomato germinating seedlings, TA... 30 10.0 emb|Z50113|SPAC31A2 S.pombe chromosome I cosmid c31A2. emb|AW728950|AW728950 GA\_ Ea0018L17 Gossypium arboreum 7-10 dpa ... 30 10.0 gb|L38011|L38011 BNAF0228E Mustard flower buds Brassica rapa cDN... 30 10.0

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Database: plantfungal

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15 661,018 sequences; 426,114,510 total letters

Searching.....done

Score E

20 Sequences producing significant alignments:

(bits) Value emb|AV428133|AV428133 AV428133 Lotus japonicus young plants (two... 53 9e-07 emb|AV412192|AV412192 AV412192 Lotus japonicus young plants (two... 53 9e-07 emb|AW350135|AW350135 GM210007B10E8R Gm-r1021 Glycine max cDNA 3... 51 2e-06 25 emb|AW349971|AW349971 GM210006B10B12R Gm-r1021 Glycine max cDNA ... 51 2e-06 emb|AW032575|AW032575 EST276134 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AW030108|AW030108 EST273363 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AI895822|AI895822 EST265265 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AI759855|AI759855 sb65c01.yl Gm-c1017 Glycine max cDNA clone... 51 2e-06 30 emb|AI777275|AI777275 EST258240 tomato resistant, Cornell Lycope... 51 2e-06 emb|AW625260|AW625260 EST319263 tomato radicle, 5 d post-imbibit... 51 2e-06 emb|AW441936|AW441936 EST311332 tomato fruit red ripe, TAMU Lyco... 51 2e-06 emb|AI441970|AI441970 sa82e02.yl Gm-c1004 Glycine max cDNA clone... 51 2e-06 emb|AW705827|AW705827 sk63d05.yl Gm-c1016 Glycine max cDNA clone... 51 3e-06 35 emb|AV428923|AV428923 AV428923 Lotus japonicus young plants (two... 51 3e-06 gb|BE124339|BE124339 EST393374 GVN Medicago truncatula cDNA clon... 48 2e-05 emb|AW442099|AW442099 EST311495 tomato fruit red ripe, TAMU Lyco... 48 3e-05 emb|AW625258|AW625258 EST319261 tomato radicle, 5 d post-imbibit... 47 4e-05 emb|AI779024|AI779024 EST259903 tomato susceptible, Cornell Lyco... 46 1e-04 40 emb|AW775746|AW775746 EST334811 DSIL Medicago truncatula cDNA cl... 46 1e-04 emb|AI960660|AI960660 sc87f03.yl Gm-c1018 Glycine max cDNA clone... 46 1e-04 emb|AW278224|AW278224 sf41c11.yl Gm-c1009 Glycine max cDNA clone... 45 3e-04 gb|BE059759|BE059759 sn36g02.yl Gm-c1016 Glycine max cDNA clone ... 45 3e-04 emb|AW348296|AW348296 GM210001B23C7R Gm-r1021 Glycine max cDNA 3... 44 4e-04 45 emb|AW685445|AW685445 NF029E11NR1F1000 Nodulated root Medicago t... 44 4e-04 emb|AI442537|AI442537 sa32g06.yl Gm-c1004 Glycine max cDNA clone... 41 0.004 emb|Z99164|SPAC29B12 S.pombe chromosome I cosmid c29B12. 41 0.005 emb|AW099733|AW099733 sd30f03.y2 Gm-c1012 Glycine max cDNA clone... 40 0.005 emb|AU011832|AU011832 AU011832 Schizosaccharomyces pombe late lo... 40 0.006 50 emb|AW622091|AW622091 EST312889 tomato root during/after fruit s... 39 0.017 emb|AW152886|AW152886 se32c07.yl Gm-c1015 Glycine max cDNA clone... 38 0.024 emb|AW661378|AW661378 833004H08.yl C. reinhardtii CC-125 -S, Lam... 38 0.033 emb|AW625785|AW625785 EST319692 tomato radicle, 5 d post-imbibit... 37 0.045

55 emb|X02873|DCEXTG Carrot gene for extensin. 37 0.062 emb|AW926428|AW926428 HVSMEg0007D04 Hordeum vulgare pre-anthesis... 35 0.063 emb|AI988320|AI988320 sc99b12.yl Gm-c1020 Glycine max cDNA clone... 36 0.086 emb|X55685|LEEXTEN5 Tomato extensin mRNA (clone uG-18). 36 0.12 emb|X55682|LEEXTEN2 Tomato extensin mRNA (clone wY). 35 0.16 60 emb|X55681|LEEXTEN1 Tomato extensin mRNA (clone w17-1). 35 0.16

emb|AQ639212|AQ639212 927P1-10E7.TP 927P1 Trypanosoma brucei gen... 30 0.055

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	111715005017745050
	emb AF163959 AF163959 Saccharomyces cerevisiae isolate wt-20 tel 29 0.23
10	gb[M34311]YSCTSDNAB S.cerevisiae telomeric sequence DNA, clone Y 27 0.25
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	emb X86030 VURNEXT26 V.unguiculata mRNA for extensine-like prote 34 0.58
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	emb AI895893 AI895893 EST265336 tomato callus, TAMU Lycopersicon 33 0.79
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35	emb AI066197 AI066197 TENU2588 T. cruzi epimastigote normalized 33 0.79
	gb L16776 LEIGP63Z Leishmania guyanensis (clone Lg63c7) major su 29 1.0
	gb L07282 CREPETE Chlamydomonas reinhardtii plastocyanin (petE) 33 1.1
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50	emb AI496548 AI496548 sb13b10.y1 Gm-c1004 Glycine max cDNA clone 32 1.5
30	emb AW686104 AW686104 NF034D07NR1F1000 Nodulated root Medicago t 32 1.5
	gb M76670 TOMEXTENA L.esculentum extensin (class I) gene, comple 32 1.5
	gb BE034655 BE034655 ML01E08 ML Mesembryanthemum crystallinum cD 32 1.5
	emb AL161025 L3320Y Leishmania major Friedlin cosmid L3320 t7 en 32 1.5
<i>5                                    </i>	emb AW329194 AW329194 N200406e rootphos(-) Medicago truncatula c 32 1.5
55 .	emb AF163952 AF163952 Saccharomyces cerevisiae isolate wt-1 telo 27 1.5
	emb Z98532 SPAC1B1 S.pombe chromosome I cosmid c1B1. 28 1.6
	emb X01469 PLHRP1 Plasmodium gene fragment for histidine-rich pr 32 2.0
	gb U66179 SCU66179 Skeletonema costatum fucoxanthin-chlorophyll 32 2.0
60	emb AC006280 AC006280 Plasmodium falciparum chromosome 12 clone 32 2.0
111.7	embla W277058 A W277058 a Real 1 v1 Cm al 010 Chroine may a DNA alama 22 2 0

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dbj|D85900|ASNPDIA Aspergillus oryzae DNA for protein disulfide ... 33 4.6

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5	emb AF019613 AF019613 Solanum tuberosum lipoxygenase (plox1) mRN 33 4.6
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	emb X95512 STLIPOXT6 S.tuberosum mRNA for lipoxygenase (clone T6). 33 4.6
	emb AW278834 AW278834 sf99c02.yl Gm-c1019 Glycine max cDNA clone 32 6.3
	emb AL160371 LMFLCHR15 Leishmania major Friedlin assembled chrom 32 6.3
	emb AB005555 AB005555 Triticum aestivum mRNA for wga20, complete 32 6.3
10	emb AA786508 AA786508 m3a04a1.rl Aspergillus nidulans 24hr asexu 32 6.3
	emb AJ274333 AJ274333 AJ274333 Metarhizium anisopliae ARSEF 2575 32 6.3
	emb Y14007 TAY14007 Triticum aestivum mRNA for gibberellin 20-ox 32 6.3
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25	gb U10247 TGU10247 Toxoplasma gondii RH hypoxanthine-xanthine-gu 32 8.6
	emb AB012774 AB012774 AB012774 Toxoplasma gondii mRNA (T.Koyama) 32 8.6
	gb S73865 S73865 linoleate:oxygen oxidoreductase [Solanum tubero 32 8.6
	emb AW761275 AW761275 sl65d12.yl Gm-c1027 Glycine max cDNA clone 32 8.6
	emb AL112245 CNS019Y5 Botrytis cinerea strain T4 cDNA library un 32 8.6
30	emb X56260 HVGLUEND Barley DNA for (1,3;1,4)-beta-glucanase.(EC 32 8.6
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	emb AI162017 AI162017 A011P07U Hybrid aspen plasmid library Popu 30 9.4
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40 45 50	Query= Novartis31_at 14141_at /id_source /description no hits found less than or equal to 1e-15. /blast_score /ec_number /family /chip nova /gb_link /ncgi (309 letters)  Database: plantfungal 661,018 sequences; 426,114,510 total letters  Searching
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Sequences producing significant alignments: (bits) Value emb|AQ952678|AQ952678 Sheared DNA-48H20.TF Sheared DNA Trypanoso... 35 0.32 emb|AW698685|AW698685 R125 non-glandular-haired subtracted cDNA ... 32 1.4 emb|AF061282|AF061282 Sorghum bicolor 22 kDa kafirin cluster. emb|AL031854|SPBC337 S.pombe chromosome II cosmid c337. emb|AC004709|AC004709 Plasmodium falciparum chromosome 12, \*\*\* S... 31 5.6 emb|AC006280|AC006280 Plasmodium falciparum chromosome 12 clone ... 31 5.6 gb]M74062|YSPCDC27A Schizosaccharomyces pombe cdc27+ gene, compl... 31 5.6 emb|AQ947500|AQ947500 Sheared DNA-39L12.TF Sheared DNA Trypanoso... 31 5.6 10 gb|M83307|YSPCDC27B Schizosaccharomyces pombe cdc27+ mRNA, compl... 31 5.6 emb|AL031856|SPBC1734 S.pombe chromosome II cosmid c1734. emb|AC073246|AC073246 Trypanosoma brucei chromosome II clone RPC... 30 7.7 emb|AL031535|SPCC16C4 S.pombe chromosome III cosmid c16C4. 15 emb|AC012647|AC012647 Trypanosoma brucei chromosome II clone RPC... 30 7.7 gb|L28038|TRBKINCSBA Trypanosoma rangeli kinetoplast DNA sequenc... 30 7.7 emb|AB004322|AB004322 Nicotiana sylvestris NsPMT1 gene for putre... 30 7.7 emb|AQ941496|AQ941496 Sheared DNA-44O18.TF Sheared DNA Trypanoso... 30 7.7 emb|AQ946940|AQ946940 Sheared DNA-36C7.TF Sheared DNA Trypanosom... 30 7.7 emb|AQ942433|AQ942433 Sheared DNA-45G7.TR Sheared DNA Trypanosom... 30 7.7 20 emb|AF133529|AF133529 Candida albicans mRNA cap methyltransferas... 30 7.7 Query= Novartis73 RC at 14202 at /id source 25 /description no hits found less than or equal to 1e-15. /blast\_score /ec number /family /chip nova /gb link (430 letters) Database: plantfungal 30 661,018 sequences; 426,114,510 total letters Searching.....done Score E 35 Sequences producing significant alignments: (bits) Value emb|AQ952678|AQ952678 Sheared DNA-48H20.TF Sheared DNA Trypanoso... 35 0.32 emb|AW698685|AW698685 R125 non-glandular-haired subtracted cDNA ... 32 1.4 emb|AF061282|AF061282 Sorghum bicolor 22 kDa kafirin cluster. 40 emb|AL031854|SPBC337 S.pombe chromosome II cosmid c337. emb|AC004709|AC004709 Plasmodium falciparum chromosome 12, \*\*\* S... 31 5.6 emb|AC006280|AC006280 Plasmodium falciparum chromosome 12 clone ... 31 5.6 gb[M74062|YSPCDC27A Schizosaccharomyces pombe cdc27+ gene, compl... 31 5.6 emb|AQ947500|AQ947500 Sheared DNA-39L12.TF Sheared DNA Trypanoso... 31 5.6 45 gb]M83307|YSPCDC27B Schizosaccharomyces pombe cdc27+ mRNA, compl... 31 5.6 emb|AL031856|SPBC1734 S.pombe chromosome II cosmid c1734. emb|AC073246|AC073246 Trypanosoma brucei chromosome II clone RPC... 30 7.7 emb|AL031535|SPCC16C4 S.pombe chromosome III cosmid c16C4. emb|AC012647|AC012647 Trypanosoma brucei chromosome II clone RPC... 30 7.7 50 gb|L28038|TRBKINCSBA Trypanosoma rangeli kinetoplast DNA sequenc... 30 7.7 emb|AB004322|AB004322 Nicotiana sylvestris NsPMT1 gene for putre... 30 7.7 emb|AQ941496|AQ941496 Sheared DNA-44O18.TF Sheared DNA Trypanoso... 30 7.7 emb|AQ946940|AQ946940 Sheared DNA-36C7.TF Sheared DNA Trypanosom... 30 7.7 emb|AQ942433|AQ942433 Sheared DNA-45G7.TR Sheared DNA Trypanosom... 30 7.7 55 emb|AF133529|AF133529 Candida albicans mRNA cap methyltransferas... 30 7.7 Query= Novartis9\_at 14223\_at /id\_source /description emb|caa19683.1| (al024486) putative protein [arabidopsis 60 thaliana] /blast\_score 0 /ec number /family /chip nova /gb\_link (1761 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters 5 Searching......done Score Sequences producing significant alignments: (bits) Value 10 emb|AW745943|AW745943 WS1\_38 H06.b1\_A002 Water-stressed I (WS1) ... 240 3e-62 gb|BE023215|BE023215 sm70a04.yl Gm-c1028 Glycine max cDNA clone ... 236 5e-61 emb|AW399343|AW399343 EST309843 L. pennellii trichome, Cornell U... 234 2e-60 emb|AW618318|AW618318 EST320304 L. pennellii trichome, Cornell U... 177 1e-53 emb|AW310982|AW310982 sg31b02.x1 Gm-c1024 Glycine max cDNA clone... 97 1e-51 15 emb|AI442277|AI442277 sa66a04.yl Gm-c1004 Glycine max cDNA clone... 200 2e-50 emb|AW687759|AW687759 NF013B05RT1F1044 Developing root Medicago ... 160 1e-46 emb|AW684122|AW684122 NF012F06NR1F1000 Nodulated root Medicago t... 83 2e-44 emb|AW309867|AW309867 sf25g10.x1 Gm-c1028 Glycine max cDNA clone... 89 5e-43 gb|BE020423|BE020423 sm40e08.yl Gm-c1028 Glycine max cDNA clone ... 88 2e-39 20 emb|AW691118|AW691118 NF041D08ST1F1000 Developing stem Medicago ... 78 2e-35 emb|AW160235|AW160235 EST290093 L. pennellii trichome, Cornell U... 120 6e-32 emb|AI960862|AI960862 sc91f07.yl Gm-c1019 Glycine max cDNA clone... 74 5e-30 emb|AW349633|AW349633 GM210006A11B3R Gm-r1021 Glycine max cDNA 3... 89 8e-29 emb|AW775076|AW775076 EST334227 KV3 Medicago truncatula cDNA clo... 81 6e-26 25 gb|BE020331|BE020331 sm40d08.yl Gm-c1028 Glycine max cDNA clone ... 69 4e-23 emb|AW746031|AW746031 WS1\_39 A03.b1\_A002 Water-stressed 1 (WS1) ... 74 8e-23 emb|AW621546|AW621546 EST312344 tomato root during/after fruit s... 64 le-13 emb|AW622846|AW622846 EST306916 tomato flower buds 3-8 mm, Corne... 54 2e-12 gb|L43984|BLYBD Hordeum vulgare (clone ABG377) chromosome 3H STS... 69 8e-11 30 emb|AW774216|AW774216 EST333367 KV3 Medicago truncatula cDNA clo... 69 1e-10 emb|AW774324|AW774324 EST333475 KV3 Medicago truncatula cDNA clo... 69 1e-10 emb|AW686169|AW686169 NF038H07NR1F1000 Nodulated root Medicago t... 68 2e-10 emb|AW160234|AW160234 EST290092 L. pennellii trichome, Cornell U... 46 2e-08 emb|AW685496|AW685496 NF030G02NR1F1000 Nodulated root Medicago t... 45 2e-07 35 emb|AW684082|AW684082 NF011H08NR1F1000 Nodulated root Medicago t... 55 2e-06 emb|AW693839|AW693839 NF069F08ST1F1074 Developing stem Medicago ... 53 6e-06 emb|AI163659|AI163659 A046p04u Hybrid aspen plasmid library Popu... 53 6e-06 emb|AW033404|AW033404 EST276975 tomato callus, TAMU Lycopersicon... 49 le-04 emb|AI054662|AI054662 coau0001K11 Cotton Boll Abscission Zone cD... 37 2e-04 40 emb|AW758414|AW758414 874008A01.y1 C. reinhardtii CC-1690, Lambd... 41 0.022 emb|AW746188|AW746188 WS1\_39\_A03.g1\_A002 Water-stressed 1 (WS1) ... 41 0.022 emb|Z99126|SPAC26H5 S.pombe chromosome I cosmid c26H5. emb|AJ225805|EDKCHALPH Egeria densa mRNA for inward potassium ch... 39 0.14 emb|AI488725|AI488725 EST247064 tomato ovary, TAMU Lycopersicon ... 38 0.20 45 emb|Z69731|SPAC6C3 S.pombe chromosome I cosmid c6C3. 36 0.71 emb|Z38061|SC9168 S.cerevisiae chromosome IX cosmid 9168. emb|AW925349|AW925349 HVSMEg0001B12 Hordeum vulgare pre-anthesis... 36 0.71 emb|AW671585|AW671585 LG1\_348\_H11.b1\_A002 Light Grown 1 (LG1) So... 36 0.97 gb|BE024935|BE024935 894007A03.x1 C. reinhardtii CC-1690, normal... 36 0.97 50 emb|AW216501|AW216501 EST295215 tomato callus, TAMU Lycopersicon... 36 0.97 emb|AW680841|AW680841 WS1\_7\_E11.g1\_A002 Water-stressed 1 (WS1) S... 35 1.3 emb|AQ911629|AQ911629 LMAJFV1\_lm96c04.yl Leishmania major FV1 ra... 35 1.3 emb|AW100578|AW100578 sd57b08.y1 Gm-c1008 Glycine max cDNA clone... 35 1.3 emb|AW010724|AW010724 ST11E02 Pine TriplEx shoot tip library Pin... 35 1.8 emb|AL138854|SPAC23G3 S.pombe chromosome I cosmid c23G3. emb|AW934681|AW934681 EST353573 tomato flower buds 0-3 mm, Corne... 34 3.5 emb|Z98975|SPAC19E9 S.pombe chromosome I cosmid c19E9. emb|AW690479|AW690479 NF035A03ST1F1000 Developing stem Medicago ... 34 3.5 emb|Z50142|SPAC2F7 S.pombe chromosome I cosmid c2F7. 60 emblAA728334|AA728334 LmLv39p7/70M Leishmania major promastigote... 34 4.8

emb|AI779448|AI779448 EST260327 tomato susceptible, Cornell Lyco... 34 4.8

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15	emb AI778201 AI778201 EST259080 tomato susceptible, Cornell Lyco 33 9.0 emb AZ215585 AZ215585 Sheared DNA-74E7.TR Sheared DNA Trypanosom 33 9.0 emb AF216960 AF216960 Pichia pastoris strain GS115 Sec12 (SEC12) 33 9.0 emb AI896488 AI896488 EST265931 tomato callus, TAMU Lycopersicon 33 9.0 emb AW670919 AW670919 LG1_266_B11.b1_A002 Light Grown 1 (LG1) So 33 9.0
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30	emb AZ215288 AZ215288 Sheared DNA-83F7.TR Sheared DNA Trypanosom 33 9.0 emb AW282605 AW282605 LG1_310_E08.g1_A002 Light Grown 1 (LG1) So 29 9.8
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40	Database: plantfungal 661,018 sequences; 426,114,510 total letters
	Searchingdone
45	Score E Sequences producing significant alignments: (bits) Value
50	emb AV417327 AV417327 AV417327 Lotus japonicus young plants (two       28 0.003         emb AV411846 AV411846 AV411846 Lotus japonicus young plants (two       28 0.005         emb AV423458 AV423458 AV423458 Lotus japonicus young plants (two       28 0.005         emb AV422547 AV422547 AV422547 Lotus japonicus young plants (two       28 0.006         emb AW706554 AW706554 sj58h12.yl Gm-c1033 Glycine max cDNA clone       28 0.010         emb AW508852 AW508852 si41b07.yl Gm-r1030 Glycine max cDNA clone       28 0.015
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60	gb BE055645 GA_Ea0021E15f Gossypium arboreum 7-10 dpa 33 0.19 gb U91857 SHU91857 Stylosanthes hamata ethylene-responsive eleme 28 0.29 emb AW329270 AW329270 N200490e rootphos(-) Medicago truncatula c 29 0.33 emb X02873 DCEXTG Carrot gene for extensin. 35 0.34

	gb M11221 DAREXTA Carrot (D.carota) extensin mRNA, partial cds. 35 0.47
	emb AW306694 AW306694 sf47b08.yl Gm-c1009 Glycine max cDNA clone 27 0.5
	emb AW035648 AW035648 EST281480 tomato callus, TAMU Lycopersicon 28 0.5
	emb AW559374 AW559374 EST314422 DSIR Medicago truncatula cDNA cl 27 0.5
5	emb AV411578 AV411578 AV411578 Lotus japonicus young plants (two 26 0.68
	emb AW618112 AW618112 EST314162 L. pennellii trichome, Cornell U 28 0.72
,	emb AW164515 AW164515 se74g06.yl Gm-c1023 Glycine max cDNA clone 27 0.7
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	emb AW328989 AW328989 N200181e rootphos(-) Medicago truncatula c 27 0.81
10	emb AV425804 AV425804 AV425804 Lotus japonicus young plants (two 34 0.89
	emb AV427232 AV427232 AV427232 Lotus japonicus young plants (two 34 0.89
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	emb AI486798 AI486798 EST245120 tomato ovary, TAMU Lycopersicon 28 1.3
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	gb[M12865]YSCRSCATC Yeast (S. cerevisiae) CAT repetitive element, 27 2.7
	emb AW040234 AW040234 EST282740 tomato mixed elicitor, BTI Lycop 28 3.0
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	emb AI900301 AI900301 sc03g01.yl Gm-c1012 Glycine max cDNA clone 31 6.0
60	emb AI487770 AI487770 EST246092 tomato ovary, TAMU Lycopersicon 28 6.6
	emb AA840807 AA840807 CFB53 Floral bud cDNA library of Hot penne 23 68

emb|AF056182|AF056182 Emericella nidulans G-protein beta subunit... 30 8.2 emb|AC007862|AC007862 Trypanosoma brucei chromosome II clone RPC... 30 8.2 emb|AC010851|AC010851 Leishmania major chromosome 22 clone L4134... 30 8.2 emb|AW707173|AW707173 sk10f12.yl Gm-c1023 Glycine max cDNA clone... 30 8.2 5 emb|AW686992|AW686992 NF004G07RT1F1055 Developing root Medicago ... 30 8.2 emb|AW618891|AW618891 EST320877 L. pennellii trichome, Cornell U... 30 8.2 emb|AW507786|AW507786 si45a10.yl Gm-r1030 Glycine max cDNA clone... 30 8.2 emb|AQ910557|AQ910557 GSSTc05793 Trypanosome cruzi random genomi... 28 8.5 emb|AW032140|AW032140 EST275594 tomato callus, TAMU Lycopersicon... 26 8.7 10 emb|AW932239|AW932239 EST358082 tomato fruit mature green, TAMU ... 26 8.7 emb|AW933044|AW933044 EST358887 tomato fruit mature green, TAMU ... 26 8.7 emb|AW030545|AW030545 EST273800 tomato callus, TAMU Lycopersicon... 26 8.8 emb|AW932961|AW932961 EST358804 tomato fruit mature green, TAMU ... 26 8.8 15. Query= PAD3 at 14248 at /id source genbank /description "gb|aad31062.1|ac007357 11 (ac007357) strong similarity to gb|x97864 cytochrome p450 from arabidopsis thaliana and is a member of the pf|00067 cytochrome p450 family. ests gb|n65665, gb|t14112, 20 gb|t76255, gb|t20906 and gb|ai100027 come from this gene." /blast\_score 0 /ec\_number /family /chip nova /gb\_link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|pad3| /ncgi http://www.ncgr.org/cgi-bin/ff?pad3 25 (1635 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 30 Searching......done Score Sequences producing significant alignments: (bits) Value 35 emblAF029858|AF029858 Sorghum bicolor cytochrome P450 CYP71E1 (C... 220 e-104 gb[M32885]AVOCYP Avocado cytochrome P-450LXXIA1 (cyp7JA1) mRNA.... 173 3e-89 gb|BE054146|BE054146 GA\_Ea0034H12f Gossypium arboreum 7-10 dpa ... 167 7e-50 gb|L24438|THLCYP450A Thlaspi arvense germline cytochrome P450 mR... 144 2e-48 emb|Z22545|PHFLAHYDB P.hybrida flavonoid 3',5'-hydroxylase mRNA. 40 dbj|D14588|PETHF1 Petunia hybrida Hf1 mRNA for flavonoid-3',5'-h... 77 9e-47 emb|Z22544|PHFLAHYDA P.hybrida flavonoid 3',5'-hydroxylase mRNA. 71 2e-44 dbi|D85184|D85184 Gentiana triflora mRNA for flavonoid 3',5'-hyd... 85 8e-44 emb|AW234222|AW234222 sf22f08.yl Gm-c1028 Glycine max cDNA clone... 147 5e-43 emb|Y09423|NRCYP71A5 Nepeta racemosa mRNA for cytochrome P450, C... 126 8e-40 45 emb|AF122821|AF122821 Capsicum annuum cytochrome P450 (PepCYP) m... 127 2e-39 gb|U48435|SCU48435 Solanum chacoense putative cytochrome P450 ge... 136 3e-39 emb|AJ238612|CRO238612 Catharanthus roseus mRNA for cytochrome P... 135 3e-39 emb|AF124815|AF124815 Mentha spicata cytochrome p450 mRNA, compl... 134 4e-39 emb|AW053855|AW053855 L30-2274T3 Ice plant Lambda Uni-Zap XR exp... 130 9e-39 50 gb|U48434|SCU48434 Solanum chacoense cytochrome P450 mRNA, compl... 124 2e-38 emb|AW255799|AW255799 ML868 peppermint glandular trichome Mentha... 116 2e-38 emb|AF124816|AF124816 Mentha x piperita cytochrome p450 isoform ... 123 1e-37 emb|AF022460|AF022460 Glycine max cytochrome P450 monooxygenase ... 116 3e-37 emb|AW349428|AW349428 GM210007A20E11R Gm-r1021 Glycine max cDNA ... emb|X71658|SMCYPEG8 S.melongena CYP76A1 mRNA. emb|AW830233|AW830233 sm24f03.y1 Gm-c1028 Glycine max cDNA clone... 110 2e-36 emb|AW668053|AW668053 GA\_Ea0012G23 Gossypium arboreum 7-10 dpa ... 129 3e-36 emb|Z33875|CYMPCP450 M.piperita gene for cytochrome P-450 oxidase. 126 3e-36 emb|AW690420|AW690420 NF033B04ST1F1000 Developing stem Medicago ... 79 5e-36 60 emb|AF134590|AF134590 Papaver somniferum (S)-N-methylcoclaurine ... 140 2e-35 emb|AW774664|AW774664 EST333815 KV3 Medicago truncatula cDNA clo... 144 6e-35

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55	emb AW687408 AW687408 NF009C08RT1F1065 Developing root Medicago 49 2e-10 emb AW870069 AW870069 NXNV_123_G03_F Nsf Xylem Normal wood Verti 58 2e-10 emb AI855891 AI855891 sc30h08.y1 Gm-c1014 Glycine max cDNA clone 58 1e-09 emb AW349142 AW349142 GM210003B22H5R Gm-r1021 Glycine max cDNA 3 57 3e-09 emb AW065112 AW065112 ST39G09 Pine TriplEx shoot tip library Pin 62 1e-08
60	emb AW736866 AW736866 NXNV_083_H05_F Nsf Xylem Normal wood Verti 52 2e-08 emb AW685287 AW685287 NF027B11NR1F1000 Nodulated root Medicago t 46 4e-08

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10
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                                                                     34 3.4
      emb|AA741585|AA741585 LmLv39p3/132B Leishmania major promastigot... 34 3.4
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      emb|X81715|SCMDYGENE S.cerevisiae partial MDY gene.
                                                                    34 3.4
      emb|X73956|TBMAP292 T.brucei Tb-292 mRNA for membrane associated... 34 3.4
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emblAZ212769lAZ212769 Sheared DNA-66C3.TR Sheared DNA Trypanosom... 34 3.4 emb|X82612|SCATM1 S.cerevisiae ATM1 gene. 34 3.4 emb|AC005139|AC005139 Plasmodium falciparum chromosome 12, \*\*\* S... 34 4.7 emb|AA649446|AA649446 T4369 MVAT4 bloodstream form of serodeme W... 34 4.7 5 Query= PAL1-mRNA s at 14254 s at /id source genbank /description gb|aad18156.2| (ac006260) phenylalanine ammonia lyase (pal1) [arabidopsis /blast\_score 1.00e-134 /ec\_number /family /chip nova 10 /gb link /ncgi (848 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 15 Searching.....done Score E Sequences producing significant alignments: (bits) Value 20 emb|AF036948|AF036948 Prunus avium phenylalanine ammonia-lyase (... 419 e-115 emb|AJ002221|DLJ002221 Digitalis lanata mRNA for phenylalanine a... 412 e-113 dbj|D83076|D83076 Lithospermum erythrorhizon mRNA for phenylalan... 411 e-113 dbj|D83075|D83075 Lithospermum erythrorhizon mRNA for phenylalan... 408 e-112 25 emb[Y12461|HAPHAMLY Helianthus annuus mRNA for phenylalanine amm... 331 e-112 emb|AF237955|AF237955 Rubus idaeus phenylalanine ammonia-lyase 2... 407 e-112 gb|M29232|IPBPAL Sweet potato phenylalanine ammonia-lyase mRNA (... 407 e-112 emb|AB042520|AB042520 Catharanthus roseus mRNA for phenylalanine... 328 e-111 gb|L36822|SSNPAL Stylosanthes humilis phenylalanine ammonia-lyas... 403 e-111 30 dbj D26596 CMEPAL Camellia sinensis mRNA for phenylalanine ammon... 403 e-111 emb[X58180]MSPAL M.sativa PAL mRNA for phenylalanine ammonia-lyase. 398 e-109 gb|L11747|POPPALGA Populus tricocarpa X Populus deltoides (hybri... 318 e-109 gb|U43338|CLU43338 Citrus limon phenylalanine ammonia-lyase (pal... 396 e-109 emb|AF237954|AF237954 Rubus idaeus phenylalanine ammonia-lyase 1... 395 e-108 35 emb|X17462|PCPAL4 P.crispum RNA for PAL4, phenylalanine ammonia-... 319 e-108 emb[X78269[NTPHEAL N.tabacum (Samsun NN) mRNA for phenylalanine ... 312 e-106 dbj|D17467|TOBPAL1 Tobacco mRNA for phenylalanine ammonia-lyase,... 312 e-105 emb|AJ250836|CAR250836 Cicer arietinum mRNA for phenylalanine am... 373 e-102 emb|X81159|PCPAL3 P.crispum mRNA for phenylalanine ammonia-lyase 3. 174 e-102 40 emb|X81158|PCPAL2 P.crispum mRNA for phenylalanine ammonia-lyase 2. 173 e-101 emb|X99997|BFPAL B.finlaysoniana mRNA for phenylalanine ammonia-... 214 2e-95 emb|AW667320|AW667320 GA\_Ea0008P06 Gossypium arboreum 7-10 dpa ... 349 8e-95 dbj|E04042|E04042 cDNA sequence coding for pea phenylalanine amm... 347 3e-94 dbi|D10001|PEAPAL P.sativum mRNA for phenylalanine ammonia-lyase... 347 3e-94. 45 dbj|D78640|IPBPALA Ipomoea batatas mRNA for phenylalanine ammoni... 291 3e-94 emb|AJ238754|CCL238754 Citrus clementina X Citrus reticulata mRN... 314 3e-91 emb|AW220322|AW220322 EST302805 tomato root during/after fruit s... 326 6e-88 emb|AW725857|AW725857 GA\_Ea0019O22 Gossypium arboreum 7-10 dpa ... 319 9e-86 emb|AJ238753|CCL238753 Citrus clementina X Citrus reticulata mRN... 319 1e-85 50 dbj|D10003|PEAPAL2 P.sativum gene for phenylalanine ammonia-lyas... 220 2e-85 emb|Z49147|HVPAL7RM H.vulgare partial PAL:mRNA for phenylalanine... 154 5e-85 gb|U16130|PAU16130 Persea americana phenylalanine ammonia lyase ... 296 9e-81 emb|AW774460|AW774460 EST333611 KV3 Medicago truncatula cDNA clo... 302 1e-80 emb|X63103|STPAL1 S.tuberosum PAL-1 gene for phenylalanine ammon... 196 6e-79 55 emb|AW685111|AW685111 NF026A05NR1F1000 Nodulated root Medicago t... 292 8e-78 emb|AF165998|AF165998 Vigna unguiculata phenylalanine ammonia-ly... 292 1e-77 emb|AI899906|AI899906 sb96a02.yl Gm-c1012 Glycine max cDNA clone... 287 3e-76 emb|AW691458|AW691458 NF045B11ST1F1000 Developing stem Medicago ... 286 6e-76 emb|AW696215|AW696215 NF103G12ST1F1099 Developing stem Medicago ... 283 7e-76 60 emb|AW688208|AW688208 NF004F12ST1F1000 Developing stem Medicago ... 278 1e-73 emb|AF218453|AF218453 Coffea arabica clone 369.1.6r phenylalanin... 275 9e-73

emb|AF218454|AF218454 Coffea arabica clone 430.4 phenylalanine a... 275 2e-72 emblAW560722|AW560722 EST315770 DSIR Medicago truncatula cDNA cl... 269 7e-71 emb|X76130|CMPAL C.melo (cantaloupe) pal mRNA. 262 le-68 emb|AV407891|AV407891 AV407891 Lotus japonicus young plants (two... 260 3e-68 5 emb|AI729040|AI729040 BNLGHi12413 Six-day Cotton fiber Gossypium... 258 1e-67 emb|AI780119|AI780119 EST260998 tomato susceptible, Cornell Lyco... 237 3e-66 emb[X99705]TAPALGEN1 T.aestivum PAL gene. emb|AW694081|AW694081 NF072E06ST1F1050 Developing stem Medicago ... 225 8e-65 emb|AW688761|AW688761 NF011C12ST1F1000 Developing stem Medicago ... 246 7e-64 10 emb|AW684895|AW684895 NF022G07NR1F1000 Nodulated root Medicago t... 237 3e-63 emb|AW267882|AW267882 EST306160 DSIR Medicago truncatula cDNA cl... 240 4e-62 emb|AW685144|AW685144 NF025D11NR1F1000 Nodulated root Medicago t... 232 5e-61 emb|AW686498|AW686498 NF042A10NR1F1000 Nodulated root Medicago t... 168 2e-60 gb|U39792|PTU39792 Pinus taeda phenylalanine ammonia-lyase (lpPA... 129 5e-60 15 dbj|D10002|PEAPAL1 P.sativum gene for phenylalanine ammonia-lyas... 231 2e-59 emblAW720528|AW720528 LjNEST18f8r Lotus japonicus nodule library... 146 7e-58 emb|AI563248|AI563248 EST00372 watermelon lambda zap library Cit... 222 7e-57 emb|AJ289609|BPE289609 Betula pendula partial pal gene for pheny... 133 3e-56 emb|AW685192|AW685192 NF027D11NR1F1000 Nodulated root Medicago t... 219 6e-56 20 emb|AJ278116|BPE278116 Betula pendula partial pal1 gene for phen... 131 9e-56 emb|AW033848|AW033848 EST277419 tomato callus, TAMU Lycopersicon... 218 1e-55 emb|AW039368|AW039368 EST281625 tomato mixed elicitor, BTI Lycop... 218 1e-55 emb|AB015871|AB015871 Vitis vinifera gene for phenylalanine ammo... 215 8e-55 emb|AW031670|AW031670 EST275124 tomato callus, TAMU Lycopersicon... 215 8e-55 25 emb|AW650661|AW650661 EST329115 tomato germinating seedlings, TA... 213 4e-54 emb|AB015870|AB015870 Vitis vinifera gene for phenylalanine ammo... 213 4e-54 emb[X52953|DMPAL1 Glycine max PAL1 gene for phenylalanine ammoni... 213 5e-54 emb|AW219754|AW219754 EST302236 tomato root during/after fruit s... 211 2e-53 emb|AW690758|AW690758 NF033B06ST1F1000 Developing stem Medicago ... 210 3e-53 30 dbj|D30656|POPPALA Populus kitakamiensis gene for phenylalanine ... 208 1e-52 dbi|D43802|POPPALG2BA Populus kitakamiensis gene for phenylalani... 208 2e-52 emb|AW689402|AW689402 NF018G12ST1F1000 Developing stem Medicago ... 207 2e-52 gb|M91192|TFRPAL1X Trifolium subterraneum phenylalanine ammonia-... 197 3e-52 emblAW034358|AW034358 EST277929 tomato callus, TAMU Lycopersicon... 190 1e-51 35 emb|AB008200|AB008200 Nicotiana tabacum palB gene for phenylalan... 204 2e-51 gb|M84466|TOBTPA1A Tobacco phenylalanine ammonialyase (tpa1) gen... 204 2e-51 gbM90692 TOMPAL5A Lycopersicon esculentum phenylalanine ammonia... 203 5e-51 emb[Y07654]PCPAL1 P.crispum pal1 gene. dbi|D43803|POPPALG4B Populus kitakamiensis gene for phenylalanin... 201 2e-50 40 emb|AI495627|AI495627 sb11c09.yl Gm-c1004 Glycine max cDNA clone... 140 4e-50 gb|M83314|TOMPHEAMLY Lycopersicon esculentum phenylalanine ammon... 199 7e-50 dbj|D85850|D85850 Daucus carota gDcPAL1 gene for phenylalanine a... 198 1e-49 emb]AW034782|AW034782 EST278818 tomato callus, TAMU Lycopersicon... 141 1e-49 emb|AW689099|AW689099 NF015D05ST1F1000 Developing stem Medicago ... 197 3e-49 45 emb|AB008199|AB008199 Nicotiana tabacum palA gene for phenylalan... 195 6e-49 emb|X15473|PCPAL1GN P.crispum PAL-1 gene for phenylalanine ammon... 120 6e-49 emb[X63104|STPAL2 S.tuberosum PAL-2 gene for phenylalanine ammon... 193 5e-48 emb|AI974372|AI974372 T110225e KV0 Medicago truncatula cDNA clon... 191 2e-47 dbj|D30657|POPPALB Populus kitakamiensis gene for phenylalanine ... 185 9e-46 50 emb|AF019965|AF019965 Pinus monticola phenylalanine ammonia lyas... 130 4e-45 emb|AI778115|AI778115 EST258994 tomato susceptible, Cornell Lyco... 166 le-44 emb|AW031859|AW031859 EST275313 tomato callus, TAMU Lycopersicon... 165 4e-44 emb[X16772]PCPAL1EX2 P.crispum PAL-1 gene for phenylalanine ammo... 173 1e-42 emb|AW757191|AW757191 sl30c12.yl Gm-c1027 Glycine max cDNA clone... 174 2e-42 55 emb|AI484637|AI484637 EST242898 tomato ovary, TAMU Lycopersicon ... 174 2e-42 emb|AW683457|AW683457 NF012E02LF1F1017 Developing leaf Medicago ... 173 3e-42 emb|AI495119|AI495119 sb03f10.yl Gm-c1004 Glycine max cDNA clone... 169 7e-41 emb|AW922611|AW922611 DG1\_46\_C11.b1\_A002 Dark Grown 1 (DG1) Sorg... 102 1e-40 emb|AV409143|AV409143 AV409143 Lotus japonicus young plants (two... 166 3e-40

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Query= pall-intron\_f\_at 14256\_f\_at /id\_source genbank /description gb[aad18156.2] (ac006260) phenylalanine ammonia lyase (pal1) [arabidopsis thaliana] /blast score 7.00e-77 /ec number /family /chip nova /gb link /ncgi (3115 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

10 Searching......done

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Score Sequences producing significant alignments: (bits) Value 15 dbi|E04042|E04042 cDNA sequence coding for pea phenylalanine amm... 232 9e-60 dbj|D10001|PEAPAL P.sativum mRNA for phenylalanine ammonia-lyase... 232 9e-60 dbj|D10002|PEAPAL1 P.sativum gene for phenylalanine ammonia-lyas... 231 2e-59 emb|AW774460|AW774460 EST333611 KV3 Medicago truncatula cDNA clo... 228 2e-58 emb|AW688761|AW688761 NF011C12ST1F1000 Developing stem Medicago ... 226 5e-58 20 emb|AW694081|AW694081 NF072E06ST1F1050 Developing stem Medicago ... 225 1e-57 emblAW684895|AW684895 NF022G07NR1F1000 Nodulated root Medicago t... 222 9e-57 emb|AI899906|AI899906 sb96a02.yl Gm-c1012 Glycine max cDNA clone... 220 5e-56 emb|AF036948|AF036948 Prunus avium phenylalanine ammonia-lyase (... 220 6e-56 emb|AF237955|AF237955 Rubus idaeus phenylalanine ammonia-lyase 2... 220 6e-56 25 dbj|D10003|PEAPAL2 P.sativum gene for phenylalanine ammonia-lyas... 219 9e-56 emb|X58180|MSPAL M.sativa PAL mRNA for phenylalanine ammonia-lyase. 219 9e-56 emb|AB015871|AB015871 Vitis vinifera gene for phenylalanine ammo... 215 1e-54 dbj|D26596|CMEPAL Camellia sinensis mRNA for phenylalanine ammon... 215 2e-54 emb|AJ002221|DLJ002221 Digitalis lanata mRNA for phenylalanine a... 214 4e-54 30 emb|AB015870|AB015870 Vitis vinifera gene for phenylalanine ammo... 213 5e-54 emb|AW725857|AW725857 GA\_Ea0019O22 Gossypium arboreum 7-10 dpa ... 212 1e-53 emb|AW696215|AW696215 NF103G12ST1F1099 Developing stem Medicago ... 211 2e-53 emb|AW691458|AW691458 NF045B11ST1F1000 Developing stem Medicago ... 211 2e-53 35 dbj|D83076|D83076 Lithospermum erythrorhizon mRNA for phenylalan... 211 2e-53 gb|M29232|IPBPAL Sweet potato phenylalanine ammonia-lyase mRNA (... 211 3e-53 emb|X52953|DMPAL1 Glycine max PAL1 gene for phenylalanine ammoni... 210 5e-53 emb[Y12461|HAPHAMLY Helianthus annuus mRNA for phenylalanine amm... 205 7e-53 emb|AW267882|AW267882 EST306160 DSIR Medicago truncatula cDNA cl... 209 9e-53 40 gb|L36822|SSNPAL Stylosanthes humilis phenylalanine ammonia-lyas... 209 9e-53 dbj|D83075|D83075 Lithospermum erythrorhizon mRNA for phenylalan... 209 1e-52 emb[AW685192]AW685192 NF027D11NR1F1000 Nodulated root Medicago t... 209 1e-52 emb|AW690758|AW690758 NF033B06ST1F1000 Developing stem Medicago ... 209 1e-52 emb|AW688208|AW688208 NF004F12ST1F1000 Developing stem Medicago ... 209 1e-52 45 emb|AF237954|AF237954 Rubus idaeus phenylalanine ammonia-lyase 1... 208 2e-52 emb|AW689402|AW689402 NF018G12ST1F1000 Developing stem Medicago ... 207 3e-52 dbj|D43802|POPPALG2BA Populus kitakamiensis gene for phenylalani... 207 3e-52 gb[M91192|TFRPAL1X Trifolium subterraneum phenylalanine ammonia-... 197 3e-52 dbj D30656 POPPALA Populus kitakamiensis gene for phenylalanine ... 207 5e-52 50 emb|AB042520|AB042520 Catharanthus roseus mRNA for phenylalanine... 207 5e-52 gb|M90692|TOMPAL5A Lycopersicon esculentum phenylalanine ammonia... 207 5e-52 emb|AI729040|AI729040 BNLGHi12413 Six-day Cotton fiber Gossypium... 206 6e-52 emb|AW686498|AW686498 NF042A10NR1F1000 Nodulated root Medicago t... 153 9e-52 gb|L11747|POPPALGA Populus tricocarpa X Populus deltoides (hybri... 205 2e-51 55 emb|AB008200|AB008200 Nicotiana tabacum palB gene for phenylalan... 204 2e-51 gb|M84466|TOBTPA1A Tobacco phenylalanine ammonialyase (tpa1) gen... 204 2e-51 gb|U43338|CLU43338 Citrus limon phenylalanine ammonia-lyase (pal... 204 3e-51 emb|AW033848|AW033848 EST277419 tomato callus, TAMU Lycopersicon... 203 8e-51 emb|AW650661|AW650661 EST329115 tomato germinating seedlings, TA... 203 8e-51 60 emb|AW219754|AW219754 EST302236 tomato root during/after fruit s... 203 8e-51 emb|AW220322|AW220322 EST302805 tomato root during/after fruit s... 203 8e-51

	emb AW039368 AW039368 EST281625 tomato mixed elicitor, BTI Lycop 203 8e-51 dbj D78640 IPBPALA Ipomoea batatas mRNA for phenylalanine ammoni 202 1e-50
	emb AI563248 AI563248 EST00372 watermelon lambda zap library Cit 202 2e-50 emb AW031670 AW031670 EST275124 tomato callus, TAMU Lycopersicon 202 2e-50
5	
_	dbj D43803 POPPALG4B Populus kitakamiensis gene for phenylalanin 201 3e-50
	embly 21150 PCPA12 Projection and PNA from the model to the control of the contro
	emb X81159 PCPAL3 P.crispum mRNA for phenylalanine ammonia-lyase 3. 174 4e-50
	emb X17462 PCPAL4 P.crispum RNA for PAL4, phenylalanine ammonia 199 4e-50
10	emb X81158 PCPAL2 P.crispum mRNA for phenylalanine ammonia-lyase 2. 173 7e-50
10	emb AJ238754 CCL238754 Citrus clementina X Citrus reticulata mRN 199 7e-50
	emb X78269 NTPHEAL N.tabacum (Samsun NN) mRNA for phenylalanine 195 le-49
	gb M83314 TOMPHEAMLY Lycopersicon esculentum phenylalanine ammon 199 1e-49
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15	dbj D85850 D85850 Daucus carota gDcPAL1 gene for phenylalanine a 198 2e-49
13	dbj D17467 TOBPAL1 Tobacco mRNA for phenylalanine ammonia-lyase, 195 3e-49
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	emb AW720528 AW720528 LjNEST18f8r Lotus japonicus nodule library 116 5e-49
	emb AW689099 AW689099 NF015D05ST1F1000 Developing stem Medicago 197 5e-49
20	emb X63103 STPAL1 S.tuberosum PAL-1 gene for phenylalanine ammon 197 5e-49
20	emb AF218453 AF218453 Coffea arabica clone 369.1.6r phenylalanin 196 7e-49
	emb AB008199 AB008199 Nicotiana tabacum palA gene for phenylalan 195 9e-49
	emb X15473 PCPAL1GN P.crispum PAL-1 gene for phenylalanine ammon 120 9e-49
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25	emb AF165998 AF165998 Vigna unguiculata phenylalanine ammonia-ly 194 2e-48
	emb AW560722 AW560722 EST315770 DSIR Medicago truncatula cDNA cl 194 2e-48
	emb AV407891 AV407891 AV407891 Lotus japonicus young plants (two 193 9e-48 emb X63104 STPAL2 S.tuberosum PAL-2 gene for phenylalanine ammon 193 9e-48
	emb AJ250836 CAR250836 Cicer arietinum mRNA for phenylalanine am 191 3e-47
30	dbj D30657 POPPALB Populus kitakamiensis gene for phenylalanine 185 1e-45
	emb AI974372 AI974372 T110225e KV0 Medicago truncatula cDNA clon 185 2e-45
	emb AW034782 AW034782 EST278818 tomato callus, TAMU Lycopersicon 141 3e-45
	emb AW685144 AW685144 NF025D11NR1F1000 Nodulated root Medicago t 179 5e-45
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      emb[X05959]NTPR1AG Tobacco PR-1a gene for pathogenesis-related p... 104 4e-50
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       emb[X06361]NTPR1AG1 Nicotiana tabacum gene for pathogenesis-rela... 104 4e-50
       emb|X12485|NTPR1A Tobacco mRNA fragment for pathogenesis-related... 104 4e-50
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       emb|X06930|NTPR1AG2 Tobacco PR-1a gene for pathogenesis-related ... 104 7e-50
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       emb|AI782416|AI782416 EST263295 tomato susceptible, Cornell Lyco... 95 7e-50
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       emb|AW034167|AW034167 EST277738 tomato callus, TAMU Lycopersicon... 94 2e-49
      emb|X17680|NTPR1BA Tobacco gene for pathogenesis-related protein... 103 3e-49
       emb|X03465|NTPRPR Nicotiana tabacum mRNA for pathogenesis-relate... 103 3e-49
      dbj|D90197|TOBPR1B1 Nicotiana tabacum mRNA for PR1b protein.
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      emb[Y08804|LEPR1B1 L.esculentum mRNA for PR protein.
                                                                      92 6e-49
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      emb|X68738|LEP1P14A L.esculentum mRNA for ethylene-induced P1(p1... 92 6e-49
      emb|AI781499|AI781499 EST262378 tomato susceptible, Cornell Lyco... 92 6e-49
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      emblAW031093|AW031093 EST274400 tomato callus, TAMU Lycopersicon... 92 6e-49
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      emb|AI782545|AI782545 EST263424 tomato susceptible, Cornell Lyco...
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      emb|AI778686|AI778686 EST259565 tomato susceptible, Cornell Lyco... 92 6e-49
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      emb|AI778680|AI778680 EST259559 tomato susceptible, Cornell Lyco... 92 6e-49
      emb|AI899514|AI899514 EST268957 tomato susceptible, Cornell Lyco... 92 6e-49
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5	emb AI779287 AI779287 EST260166 tomato susceptible, Cornell Lyco 92 6e-49 emb AI773130 AI773130 EST254230 tomato resistant, Cornell Lycope 92 6e-49 emb AI778790 AI778790 EST259669 tomato susceptible, Cornell Lyco 92 6e-49 emb AI778791 AI778791 EST259670 tomato susceptible, Cornell Lyco 92 6e-49 emb AI778985 AI778985 EST259864 tomato susceptible, Cornell Lyco 92 6e-49 emb AW032026 AW032026 EST275480 tomato callus, TAMU Lycopersicon 92 6e-49 emb AI782288 AI782288 EST263167 tomato susceptible, Cornell Lyco 92 6e-49 emb AI779013 AI779013 EST259892 tomato susceptible, Cornell Lyco 92 6e-49
10	emb AI781431 AI781431 EST262298 tomato susceptible, Cornell Lyco 92 6e-49 emb AI780973 AI780973 EST261852 tomato susceptible, Cornell Lyco 92 6e-49 emb AI895236 AI895236 EST264679 tomato callus, TAMU Lycopersicon 92 6e-49
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20	Database: plantfungal 661,018 sequences; 426,114,510 total letters
	Searchingdone
25	Score E Sequences producing significant alignments: (bits) Value
30	gb M60729 HRAHRPCC A.rusticana peroxidase isoenzyme C (HPR C) ge 744 0.0 emb A00741 A00741 A.rusticana synthetic gene (reverse complement 692 0.0 emb A00740 A00740 A.rusticana synthetic gene for peroxidase. 692 0.0 dbj E01651 E01651 cDNA encoding horseradish peroxidase. 692 0.0 gb M37157 HRAHRPCB A.rusticana peroxidase isoenzyme C (HPR C) ge 371 e-133 emb X97349 PTPXP2PER P.trichocarpa mRNA for anionic peroxidase P 473 e-132
35	emb X97350 PTPXP3PER P.trichocarpa mRNA for anionic peroxidase P 469 e-131 dbj D83224 POPP01 Populus nigra mRNA for peroxidase, complete cds. 468 e-131 gb M37156 HRAHRPCA A.rusticana peroxidase isoenzyme C (HPR C) ge 363 e-130 emb X97348 PTPXP1PER P.trichocarpa mRNA for anionic peroxidase P 463 e-129
40	dbj D30652 POPPA Populus kitakamiensis mRNA for peroxidase, part 441 e-123 emb X97351 PTPXP4PER P.trichocarpa mRNA for anionic peroxidase P 383 e-120 dbj D30653 POPPB Populus kitakamiensis mRNA for peroxidase, part 356 e-112 emb AF149277 AF149277 Phaseolus vulgaris peroxidase 1 precursor 227 e-107 gb L36157 ALFPXDC Medicago sativa peroxidase (pxdC) mRNA, comple 229 e-106 emb AF007211 AF007211 Glycine max peroxidase precursor (GMIPER1) 229 e-106
45	gb L07554 LINPEROX Linum usitatissimum peroxidase (FLXPER1) mRNA 381 e-105 emb X90692 MSRNAPE1A M.sativa mRNA for peroxidase 1A. 211 e-105 dbj D90115 HRAPRXC2 Horseradish prxC2 gene encoding peroxidase i 296 e-103 emb AJ242742 IBA242742 Ipomoea batatas mRNA for peroxidase (pod 138 2e-97 emb AF149280 AF149280 Phaseolus vulgaris peroxidase 5 precursor 142 4e-97
50	emb X90693 MSRNAPE1B M.sativa mRNA for peroxidase 1B. 231 1e-95 emb X90694 MSRNAPE1C M.sativa mRNA for peroxidase 1C. 226 7e-94 gb L36111 SSNPEROXIB Stylosanthes humilis peroxidase mRNA. 210 2e-89 emb AI959837 AI959837 sc94h07.y1 Gm-c1019 Glycine max cDNA clone 228 6e-89 emb AF244924 AF244924 Spinacia oleracea peroxidase prx15 precurs 208 9e-88
55	emb AW559660 AW559660 EST314772 DSIR Medicago truncatula cDNA cl 236 1e-8' emb AW774581 AW774581 EST333732 KV3 Medicago truncatula cDNA clo 236 7e-8 emb AF155124 AF155124 Gossypium hirsutum bacterial-induced perox 215 2e-86 emb AW981426 AW981426 EST392579 DSIL Medicago truncatula cDNA cl 236 2e-85 emb AB042103 AB042103 Asparagus officinalis AspPOX1 mRNA for per 209 2e-85
60	emb AF244923 AF244923 Spinacia oleracea peroxidase prx14 precurs 205 9e-85 emb AJ250121 PAB250121 Picea abies mRNA for SPI2 protein (spi2 g 253 1e-83 emb AW775762 AW775762 EST334827 DSIL Medicago truncatula cDNA cl 236 6e-83 emb Y10466 SOPRXR5 S.oleracea mRNA for peroxidase, clone PC18. 218 3e-82

emb|AW775425|AW775425 EST334490 DSIL Medicago truncatula cDNA cl... 208 1e-81 emb|AW685437|AW685437 NF029D09NR1F1000 Nodulated root Medicago t... 230 9e-80 emb|AB024439|AB024439 Scutellaria baicalensis mRNA for peroxidas... 209 2e-79 emb|AF244922|AF244922 Spinacia oleracea peroxidase prx13 precurs... 212 3e-79 emb|X57564|ARNEUPERO A.rusticana mRNA for neutral peroxidase. emb|AW776273|AW776273 EST335338 DSIL Medicago truncatula cDNA cl... 222 1e-77 emb|AW267813|AW267813 EST305941 DSIR Medicago truncatula cDNA cl... 200 1e-76 emb|X91232|MARNAPRX M.annua mRNA for peroxidase. 196 5e-76 emb|AF049881|AF049881 Linum usitatissimum peroxidase FLXPER4 (PE... 207 7e-76 10. emb|AW256487|AW256487 EST304624 KV2 Medicago truncatula cDNA clo... 200 9e-76 emb|AW775890|AW775890 EST334955 DSIL Medicago truncatula cDNA cl... 200 3e-74 emb|AW257195|AW257195 EST305332 KV2 Medicago truncatula cDNA clo... 214 5e-74 gb[M91373]CUSPREPER Cucumis sativus peroxidase mRNA, complete cds. 86 2e-73 gb|U41657|GMU41657 Glycine max seed coat peroxidase isozyme (SPO... 159 3e-73 15 emb|AB027752|AB027752 Nicotiana tabacum mRNA for peroxidase, com... 197 5e-73 gb|L36110|SSNPEROXIA Stylosanthes humilis peroxidase mRNA. 199 5e-73 gb|M37636|ARCPNC1 Arachis hypogaea cationic peroxidase (PNC1) mR... 204 7e-73 emb|AW685235|AW685235 NF027H10NR1F1000 Nodulated root Medicago t... 236 '9e-73 emb|AW278775|AW278775 sf97d02.yl Gm-c1019 Glycine max cDNA clone... 155 1e-72 20. emb|X71593|LECEVI1A L.esculentum CEVI-1 mRNA. 99 2e-72 gb|J02979|TOBPXDLF Nicotiana tabacum lignin-forming peroxidase m... 100 2e-72 emb|AW559945|AW559945 EST314993 DSIR Medicago truncatula cDNA cl... 233 2e-72 emb|AW980744|AW980744 EST391897 GVN Medicago truncatula cDNA clo... 235 5e-70 emb|Y10467|SOPRXR6 S.oleracea mRNA for peroxidase, clone PC23. 105 1e-69 25 dbj|D83225|POPP02 Populus nigra peroxidase gene, complete cds. 196 2e-69 gb|M74103|TOBANPER Nicotiana sylvestris anionic peroxidase mRNA,... 205 6e-69 emb|AF043234|AF043234 Striga asiatica ferriprotein porphyrin-con... 211 1e-68 gb|BE034991|BE034991 MM01A12 MM Mesembryanthemum crystallinum cD... 202 6e-68 emb|AW574244|AW574244 EST316835 GVN Medicago truncatula cDNA clo... 202 1e-67 30 emb[Y10465]SOPRXR4 S.oleracea mRNA for peroxidase, clone PC44. emb|AW686084|AW686084 NF038B07NR1F1000 Nodulated root Medicago t... 236 5e-67 emb|AW561032|AW561032 EST316080 DSIR Medicago truncatula cDNA cl... 203 7e-67 emb|X56011|TAPERO Wheat mRNA for peroxidase. 111 2e-66 emb|AW185769|AW185769 se59d08.y1 Gm-c1019 Glycine max cDNA clone... 219 4e-65 35 emb[Y17192|CPY17192 Cucurbita pepo mRNA for peroxidase. 96 2e-64 gb|BE033422|BE033422 ME01E09 ME Mesembryanthemum crystallinum cD... 184 4e-64 emb|AB024438|AB024438 Scutellaria baicalensis mRNA for peroxidas... 205 7e-64 gblM91374|CUSPREPERA Cucumis sativus peroxidase mRNA, complete cds. 76 2e-63 gb|L24120|LINFLXP Linum usitatissimum peroxidase precursor (FLXP... 167 2e-63 40 gb|M91372|CUSPREPERB Cucumis sativus peroxidase mRNA, complete cds. 134 2e-63 emb|AF043235|AF043235 Striga asiatica ferriprotein porphyrin-con... 199 3e-63 emb|AW288002|AW288002 N100846e rootphos(-) Medicago truncatula c... 182 7e-63 emb|AW126121|AW126121 N100318e rootphos(-) Medicago truncatula c... 216 1e-62 gb|L36093|BLYPRX Barley peroxidase mRNA, complete cds. 133 5e-62 45 emb|AW687443|AW687443 NF009F07RT1F1062 Developing root Medicago ... 235 1e-61 emb|AW687957|AW687957 NF001D11ST1F1000 Developing stem Medicago ... 236 3e-61 emb|AB024437|AB024437 Scutellaria baicalensis mRNA for peroxidas... 163 3e-61 emb|X58396|HVPEROXI Barley mRNA for peroxidase (EC=1.11.1.7). emb|AI496388|AI496388 sb04a11.y1 Gm-c1004 Glycine max cDNA clone... 224 6e-61 50 emb|AW686765|AW686765 NF042E07NR1F1000 Nodulated root Medicago t... 222 8e-61 dbj|D38050|POPP1 Aspen prxA3a gene for peroxidase, complete cds. 121 9e-61 emb|AW704659|AW704659 sk54h10.y1 Gm-c1019 Glycine max cDNA clone... 149 9e-61 emb|Y10464|SOPRXR3 S.oleracea mRNA for peroxidase, clone PC42. 80 3e-60 emb|AW705730|AW705730 sk51b02.y1 Gm-c1019 Glycine max cDNA clone... 149 8e-60 55 gb|U51191|GMU51191 Glycine max peroxidase precursor (sEPa1) mRNA... 100 9e-60 gb|U12314|CCU12314 Cenchrus ciliaris clone PX7 peroxidase mRNA, ... 105 9e-60 emb|AI938533|AI938533 sb46h09.y1 Gm-c1015 Glycine max cDNA clone... 145 1e-59 emb|AW705617|AW705617 sk50d03.y1 Gm-c1019 Glycine max cDNA clone... 149 2e-59 emb|AI781859|AI781859 EST262738 tomato susceptible, Cornell Lyco... 102 2e-59 60 emb|AF149278|AF149278 Phaseolus vulgaris peroxidase 3 precursor ... 90 3e-59 gb|U51192|GMU51192 Glycine max peroxidase precursor (sEPa2) mRNA... 102 4e-59

emb|AW441632|AW441632 EST311028 tomato fruit red ripe, TAMU Lyco... 105 7e-59 emb|AF244921|AF244921 Spinacia oleracea peroxidase prx12 precurs... 156 8e-59 gb|M32742|CUSCUPER C.sativus peroxidase (put.) (CuPer2) mRNA, 3'... 107 2e-58 emblAW705946|AW705946 sk52h07.yl Gm-c1019 Glycine max cDNA clone... 224 6e-58 5. Query= TSA1\_s\_at 14672\_s\_at /id\_source genbank /description gb|aac49117.1| (u18993) tryptophan synthase alpha chain [arabidopsis thaliana] /blast score 1.00e-158 /ec number /family /chip nova 10 /gb link /ncgi (939 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 15 Score E Sequences producing significant alignments: (bits) Value 20 emb|AW730233|AW730233 GA\_Ea0014B16 Gossypium arboreum 7-10 dpa ... 352 5e-98 emb|AW649960|AW649960 EST328414 tomato germinating seedlings, TA... 308 4e-83 emb|AW621664|AW621664 EST312462 tomato root during/after fruit s... 290 1e-77 emb|AW031327|AW031327 EST274781 tomato callus, TAMU Lycopersicon... 282 3e-75 25 gb|U38804|PPU38804 Porphyra purpurea chloroplast, complete genome. 271 4e-72 emb|AW775853|AW775853 EST334918 DSIL Medicago truncatula cDNA cl... 264 9e-70 emb|Z21642|CHASTRNAA Antithamnion sp. Chloroplast trnK,trnE,trpA... 255 4e-67 gb|BE124387|BE124387 EST393422 GVN Medicago truncatula cDNA clon... 227 9e-59 gb|BE121873|BE121873 894015F07.yl C. reinhardtii CC-1690, normal... 227 1e-58 30 emb|AF022186|AF022186 Cyanidium caldarium strain RK1 chloroplast... 220 1e-56 emb|AI782364|AI782364 EST263243 tomato susceptible, Cornell Lyco... 210 1e-53 emb|AW691298|AW691298 NF040B01ST1F1000 Developing stem Medicago ... 193 1e-49 dbj|D63675|D63675 Cyanidioschyzon merolae trnK, trpA, trnT, rps4... 130 le-49 emb|AV392084|AV392084 AV392084 Chlamydomonas reinhardtii C9 Chla... 195 5e-49 35 emb|AA660642|AA660642 00530 MtRHE Medicago truncatula cDNA 5' si... 190 1e-47 dbj|D17791|CYNPLTRNK C.caldarium chloroplast gene for trnK and g... 121 1e-46 emb|AW034248|AW034248 EST277819 tomato callus, TAMU Lycopersicon... 184 9e-46 dbi|D63676|D63676 Cyanidium caldarium trnK, trpA, trnT, rps4, tr... 122 1e-45 gb|L38526|L38526 BNAF0168E Mustard flower buds Brassica rapa cDN... 177 1e-43 40 emb[V01342]SCTRP5A Yeast gene (trp5) for tryptophan synthetase. 86 9e-42 emb|Z72548|SCYGL026C S.cerevisiae chromosome VII reading frame O... 86 9e-42 emb|AW398861|AW398861 EST309361 L. pennellii trichome, Cornell U... 158 5e-38 emb|AU090244|AU090244 AU090244 Hordeum vulgare subsp. vulgare Up... 149 7e-38 emb|AW650911|AW650911 EST329365 tomato germinating seedlings, TA... 141 1e-32 45 emb|AI773494|AI773494 EST254594 tomato resistant, Cornell Lycope... 138 5e-32 emb|Z98974|SPAC19A8 S.pombe chromosome I cosmid c19A8. emb|V01343|SCTRP5B Part of the yeast gene for tryptophan synthet... 86 2e-26 emb|AI165371|AI165371 A082p39u Hybrid aspen plasmid library Popu... 81 le-21 emb|AU090028|AU090028 AU090028 Hordeum vulgare subsp. vulgare Up... 97 2e-19 50 gb|J04594|NEUTRP3A N.crassa tryptophan synthetase (trp3) alpha-2... 78 1e-17 emb|AF084886|AF084886 Neurospora crassa 314-448A mutant tryptoph... 68 9e-11 emb|AF084890|AF084890 Neurospora crassa TD 554-6A mutant tryptop... 53 4e-10 emb|AF084880|AF084880 Neurospora crassa EMSG9-9A mutant tryptoph... 66 5e-10 emb|AF084885|AF084885 Neurospora crassa NG61-6A mutant tryptopha... 64 1e-09 55 emb|AF084884|AF084884 Neurospora crassa 314-722A mutant tryptoph... 64 le-09 emb|AF084883|AF084883 Neurospora crassa 314-709A mutant tryptoph... 64 le-09 emb|AF084882|AF084882 Neurospora crassa 314-682A mutant tryptoph... 64 le-09 emb|AF084881|AF084881 Neurospora crassa 314-693A mutant tryptoph... 64 2e-09 emb|AF084889|AF084889 Neurospora crassa 314-492-2A mutant trypto... 53 3e-09 60 gb|M29094|CCITRP01 Mushroom (C.cinereus) tryptophan synthetase (... 59 4e-08 emb|AU011013|AU011013 AU011013 Schizosaccharomyces pombe late lo... 54 2e-06

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	emb AV390244 AV390244 AV390244 Chlamydomonas reinhardtii C9 Chla 49 4e-05
	emb AW616467 AW616467 EST322878 L. hirsutum trichome, Cornell Un 48 7e-05
	emb AL354022 P761R Leishmania major Friedlin PAC P761 right end 44 0.002
5	gb M91656 CCITRP05 Coprinus cinereus tryptophan synthetase (TRP1 43 0.005
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	emb AF262997 AF262997 Ricinus communis NADP-dependent malic prot 34 1.6
	gb[M19025]CFUCPOR C.fumago cpo gene encoding chloroperoxidase, c 34 1.6
20	emb AF084888 AF084888 Neurospora crassa 656-2A mutant tryptophan 31 1.7
20	emb AF084887 AF084887 Neurospora crassa TDA78(1-A)9A(TD201) muta 31 1.7
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25	emb AW187498 AW187498 BNLGHi6414 Six-day Cotton fiber Gossypium 33 3.1
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	emb AF129874 AF129874 Pichia angusta peroxin-6 (PEX6) gene, comp 33 4.3
	emb AL112679 CNS01AA7 Botrytis cinerea strain T4 cDNA library un 33 4.7
35	gb BE028433 BE028433 EtESTea78d07.yl Eimeria M5-6 Merozoite stag 32 5.9
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gb|aab80922.1| (af022658) putative c2h2 zinc finger transcription 5 factor [arabidopsis thaliana] /blast\_score 3.00e-94 (938 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Searching......done Score Sequences producing significant alignments: (bits) Value 15 gb|U68763|GMU68763 Glycine max putative transcription factor SCO... 71 1e-35 emb|Y18788|MSY18788 Medicago sativa mRNA for putative TFIIIA (or... 71 3e-35 emb|AF053077|AF053077 Nicotiana tabacum osmotic stress-induced z... 70 3e-33 emb|AW729218|AW729218 GA Ea0024G18 Gossypium arboreum 7-10 dpa ... 71 5e-33 20 emb|AW781249|AW781249 sk67b08.y1 Gm-c1016 Glycine max cDNA clone... 70 7e-31 dbj|D26086|PETZFP4 Petunia zinc-finger protein gene. 68 3e-30 emb|AW560934|AW560934 EST315982 DSIR Medicago truncatula cDNA cl... 68 8e-30 emb|AW775559|AW775559 EST334624 DSIL Medicago truncatula cDNA cl... 70 1e-29 gb|BE123920|BE123920 EST394045 DSIL Medicago truncatula cDNA clo... 70 1e-29 25 emb|AI988657|AI988657 sd06b03.yl Gm-c1020 Glycine max cDNA clone... 70 9e-29 emb|AW102472|AW102472 sd88f02.yl Gm-c1009 Glycine max cDNA clone... 64 1e-28 dbj|D26084|PETZFDB2 Petunia mRNA for zinc-finger DNA binding pro... 69 1e-28 dbj|D26083|PETZFDB1 Petunia hybrida gene for zinc-finger DNA bin... 68 4e-28 emb|AI988290|AI988290 sc98f10.yl Gm-c1020 Glycine max cDNA clone... 64 3e-27 30 emb|AW706944|AW706944 sk08e10.y1 Gm-c1023 Glycine max cDNA clone... 69 2e-26 emb|AW153229|AW153229 se37f05.y1 Gm-c1015 Glycine max cDNA clone... 69 3e-26 emb|AW616587|AW616587 EST322998 L. hirsutum trichome, Cornell Un... 68 5e-26 gb|BE095284|BE095284 00345 leafy spurge Lambda HybriZAP 2.1 two-... 69 5e-26 emb|AW278572|AW278572 sf46c03.y1 Gm-c1009 Glycine max cDNA clone... 65 8e-26 35 emb|AI487287|AI487287 EST245609 tomato ovary, TAMU Lycopersicon ... 68 9e-26 dbj|D26085|PETZFDB3 Petunia zinc-finger DNA binding protein gene. 60 1e-25 emb|AI778714|AI778714 EST259593 tomato susceptible, Cornell Lyco... 68 1e-25 emb|AW279005|AW279005 sg04d04.y1 Gm-c1019 Glycine max cDNA clone... 67 2e-25 emb|AW164639|AW164639 se74f02.y1 Gm-c1023 Glycine max cDNA clone... 67 2e-25 40 emb|AF119050|AF119050 Datisca glomerata zinc-finger protein 1 (z... 71 2e-25 gb|BE059872|BE059872 sn38c04.y1 Gm-c1016 Glycine max cDNA clone ... 71 4e-25 gb|U76554|BRU76554 Brassica rapa zinc-finger protein-1 (BR42) mR... 63 9e-25 gb|U76555|BRU76555 Brassica rapa zinc-finger protein BcZFP1 (BcA... 62 3e-24 emb|AI966679|AI966679 sc55a11.yl Gm-c1015 Glycine max cDNA clone... 64 4e-24 45 emb|AW034622|AW034622 EST278306 tomato callus, TAMU Lycopersicon... 62 6e-24 emb|AI488218|AI488218 EST246540 tomato ovary, TAMU Lycopersicon ... 62 9e-24 emb|AW037956|AW037956 EST279600 tomato mixed elicitor, BTI Lycop... 59 4e-23 emb[Y16131|Y16131 Y16131 young root nodules Medicago sativa subs... 67 8e-23 emb|AW030858|AW030858 EST274148 tomato callus, TAMU Lycopersicon... 66 9e-23 50 emb|AW032112|AW032112 EST275566 tomato callus, TAMU Lycopersicon... 68 5e-22 emb|AW625323|AW625323 EST319146 tomato radicle, 5 d post-imbibit... 59 8e-21 emb|AW033257|AW033257 EST276828 tomato callus, TAMU Lycopersicon... 66 2e-20 emb|AI485651|AI485651 EST243972 tomato ovary, TAMU Lycopersicon ... 62 2e-20 emb|AW033574|AW033574 EST277145 tomato callus, TAMU Lycopersicon... 66 2e-20 55 emb|AI896031|AI896031 EST265474 tomato callus, TAMU Lycopersicon... 66 3e-20 emb|AI771191|AI771191 EST252387 tomato ovary, TAMU Lycopersicon ... 66 3e-20 emb|AW032357|AW032357 EST275811 tomato callus, TAMU Lycopersicon... 66 3e-20 emb|AW219736|AW219736 EST302218 tomato root during/after fruit s... 59 1e-19 emb|AW219517|AW219517 EST301915 tomato root during/after fruit s... 59 2e-19 60 emb|AW706014|AW706014 sk64g01.y1 Gm-c1016 Glycine max cDNA clone... 69 1e-18

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WO 02/22675

Database: plantfungal

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יי ענ	Databases miantifum col
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Score E

Sequences producing significant alignments:

(bits) Value

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	ALIMONA CAIDITE TITUDO CA DIL A L'AL MILLANA ANA ANTA DE L'AL CONTROL CON COMPANION DE L'AL MANAGEMENT DE L'AL CONTROL CON COMPANION DE L'AL MANAGEMENT DE L'AL CONTRO
	gb M99431 PHNHSP83A Pharbitis nil heat shock protein 83 (Hsp83) 597 0.0
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5	emb AV413314 AV413314 AV413314 Lotus japonicus young plants (two 102 8e-21 emb AW693546 AW693546 NF067D12ST1F1101 Developing stem Medicago 101 1e-20 emb AI895294 AI895294 EST264737 tomato callus, TAMU Lycopersicon 101 1e-20 emb AJ010644 PSA010643 Pisum sativum hsfA gene, exons 1 to 2, pa 99 7e-20 emb AW924303 AW924303 WS1_52_H11.b1_A002 Water-stressed 1 (WS1) 99 7e-20 emb AI895834 AI895834 EST265277 tomato callus, TAMU Lycopersicon 95 9e-19 emb AW756148 AW756148 sl16e07.yl Gm-c1036 Glycine max cDNA clone 93 4e-18 gb M94683 YSPHSF Schizosaccharomyces pombe heat shock transcript 93 6e-18
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25	emb AW933529 AW933529 EST359288 tomato fruit mature green, TAMU 72 1e-11 emb AW307441 AW307441 sf57a06.yl Gm-c1009 Glycine max cDNA clone 70 5e-11 emb AW620962 AW620962 sj98b03.yl Gm-c1023 Glycine max cDNA clone 70 5e-11
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	Searchingdone
40	Score E Sequences producing significant alignments: (bits) Value
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50	emb AI352937 AI352937 MB74-5H PZ204.BNlib Brassica napus cDNA cl 115 5e-40 emb AW621250 AW621250 EST312048 tomato root during/after fruit s 162 6e-40 emb AW626221 AW626221 EST320128 tomato radicle, 5 d post-imbibit 162 6e-40 emb AI779211 AI779211 EST260090 tomato susceptible, Cornell Lyco 162 6e-40 emb AW443140 AW443140 EST308070 tomato mixed elicitor, BTI Lycop 162 6e-40
55	emb AI776056 AI776056 EST257156 tomato resistant, Cornell Lycope 162 6e-40 emb AW032764 AW032764 EST276323 tomato callus, TAMU Lycopersicon 162 6e-40 emb AW217082 AW217082 EST295796 tomato callus, TAMU Lycopersicon 162 6e-40 emb AW035571 AW035571 EST281309 tomato callus, TAMU Lycopersicon 162 6e-40 emb AV413531 AV413531 AV413531 Lotus japonicus young plants (two 163 1e-39
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      emblAW649092|AW649092 EST327546 tomato germinating seedlings, TA... 123 3e-32
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	emb AF211541 AF211541 AF211541 34.1B Nicotiana tabacum cDNA clon 87 2e-16 emb AW621700 AW621700 EST312498 tomato root during/after fruit s 87 2e-16
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30	Sequences producing significant alignments: (bits) Value
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35	gb M23120 TOBGLUCB2 Tobacco (N.plumbaginifolia) beta-glucanase m 265 e-105 emb AJ277900 VVI277900 Vitis vinifera mRNA for beta 1-3 glucanas 284 e-104
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	dbj E02108 E02108 cDNA sequence coding for beta-1,3-endoglucanase. 226 e-100
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	gb M80608 TOMB13GLUB Lycopersicon esculentum beta-1,3-glucanase 210 1e-98
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	emb AF067863 AF067863 Solanum tuberosum 1,3-beta-glucan glucanoh 210 6e-95
	gb U01900 U01900 Solanum tuberosum Datura endo-1,3-beta-D-glucan 210 8e-95
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	emb A26449 A26449 Soya mutant beta-1,3-glucanase cDNA. 244 3e-93
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5	a learn and a summary and a su
J	emb X74906 LEQB L.esculentum TomQ'b mRNA for beta(1,3)glucanase. 110 1e-86 emb AF227953 AF227953 Capsicum annuum basic beta-1,3-glucanase ( 252 3e-86
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,	emb AF001523 AF001523 Musa acuminata beta-1, 3-glucananse mRNA, 148 5e-86
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	emb AF186083 AF186083 AF186083 Populus alba x Populus tremula ba 218 5e-85
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	gb M59443 TOBGLUCB N.tabacum acidic beta-1,3-glucanase gene, com 100 1e-75
	gopus 3-4-3 TODOLOGO 14. dozeni actuic beta-1,3-giucanase gene, com 100 1e-75
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20	gb M63634 TOBGCBREG Nicotiana plumbaginifolia beta(1,3)-glucanas 268 2e-75
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	emb AB028153 AB028153 Prunus avium mRNA for S1-RNase, complete cds. 56 3e-15
10	emb AW223027 AW223027 EST299838 tomato fruit red ripe, TAMU Lyco 43 4e-15
10	emb AW224120 AW224120 EST300931 tomato fruit red ripe, TAMU Lyco 43 5e-15
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	emb AB011470 AB011470 Prunus dulcis mRNA for Sc-RNase, complete 60 3e-13
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35	dbj E01267 E01267 cDNA encoding S2-protein linked to its signal 56 1e-11
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	emb AB026981 AB026981 Prunus salicina mRNA for Sa-RNase, partial 56 3e-11
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55	gb L40542 POTDSCS Solanum carolinense self-incompatibility ribon 65 7e-10
	Bola . 12 . 12 . 00 /010

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60

Database: plantfungal

661,018 sequences; 426,114,510 total letters 5 Score E Sequences producing significant alignments: (bits) Value emb|AI782831|AI782831 EST263710 tomato susceptible, Cornell Lyco... 270 1e-71 10 emb|AW093706|AW093706 EST286886 tomato mixed elicitor, BTI Lycop... 256 4e-67 emb|AW279194|AW279194 sf67g10.yl Gm-c1013 Glycine max cDNA clone... 205 2e-53 emb|AI776928|AI776928 EST258028 tomato resistant, Cornell Lycope... 174 1e-42 emb|AW441294|AW441294 EST310690 tomato fruit red ripe, TAMU Lyco... 88 9e-42 emb|AW035369|AW035369 EST280931 tomato callus, TAMU Lycopersicon... 88 3e-35 15 gb]BE037451|BE037451 MP21C02 MP Mesembryanthemum crystallinum cD... 88 2e-34 gb|BE058288|BE058288 sn14b07.yl Gm-c1016 Glycine max cDNA clone ... 88 2e-34 emb|AI895992|AI895992 EST265435 tomato callus, TAMU Lycopersicon... 80 3e-34 emb|AW441248|AW441248 EST310644 tomato fruit red ripe, TAMU Lyco... 88 2e-33 emb|AW223990|AW223990 EST300801 tomato fruit red ripe, TAMU Lyco... 88 2e-33 20 emb|AW761120|AW761120 sl63e05.yl Gm-c1027 Glycine max cDNA clone... 127 9e-32 gb|BE037096|BE037096 MP14H01 MP Mesembryanthemum crystallinum cD... 86 2e-30 emb|AW038060|AW038060 EST279717 tomato mixed elicitor, BTI Lycop... 84 1e-29 emb|AW030010|AW030010 EST273265 tomato callus, TAMU Lycopersicon... 84 1e-29 emb|AW035432|AW035432 EST281170 tomato callus, TAMU Lycopersicon... 83 1e-29 25 emb|AW221846|AW221846 EST298657 tomato fruit red ripe, TAMU Lyco... 80 2e-28 emb|AW907125|AW907125 EST343157 potato stolon, Cornell Universit... 127 3e-28 emb[AW306189]AW306189 se47b02.y1 Gm-c1017 Glycine max cDNA clone... 112 4e-27 emb|AW686451|AW686451 NF038B03NR1F1000 Nodulated root Medicago t... 57 1e-24 emb|AW934306|AW934306 EST360149 tomato fruit mature green, TAMU ... 80 4e-22 30 emb|AI896882|AI896882 EST266325 tomato callus, TAMU Lycopersicon... 84 6e-22 emb|AW030032|AW030032 EST273287 tomato callus, TAMU Lycopersicon... 80 8e-22 emb|AW216521|AW216521 EST295235 tomato callus, TAMU Lycopersicon... 80 8e-22 emb|AW216877|AW216877 EST295591 tomato callus, TAMU Lycopersicon... 84 2e-21 emb|AI895032|AI895032 EST264475 tomato callus, TAMU Lycopersicon... 98 2e-19 35 emb|AI054656|AI054656 coau0001K05 Cotton Boll Abscission Zone cD... 95 2e-18 emb|AW350604|AW350604 GM210008B20C3R Gm-r1021 Glycine max cDNA 3... 89 1e-16 emb|AW985224|AW985224 NXNV\_132\_F08\_F Nsf Xylem Normal wood Verti... 84 2e-15 emb|AW782240|AW782240 sm03c06.yl Gm-c1027 Glycine max cDNA clone... 80 2e-15 emb|AI894672|AI894672 EST264115 tomato callus, TAMU Lycopersicon... 80 3e-14 40 emb|AW034079|AW034079 EST277574 tomato callus, TAMU Lycopersicon... 77 3e-13 emb|AW991119|AW991119 SsS0174 Suaeda salsa ZAP cDNA library Suae... 58 7e-10 emb|X95732|NPZEAXANT N.plumbaginifolia mRNA for zeaxanthin epoxi... 63 6e-09 emb|AV418279|AV418279 AV418279 Lotus japonicus young plants (two... 48 8e-09 emb|X91491|CAXANEPOX C.annuum mRNA for xanthophyll epoxidase. 61 2e-08 45 emb|Z83835|LEZEAXAN L.esculentum mRNA for zeaxanthin epoxidase. 59 1e-07 emb|AI966377|AI966377 sc38a10.yl Gm-c1014 Glycine max cDNA clone... 49 6e-07 emb|AF159948|AF159948 Prunus armeniaca zeaxanthin epoxidase (ZEA... 54 3e-06 emb|AF071888|AF071888 Prunus armeniaca zeaxanthin epoxidase (ZEA... 54 3e-06 dbj|D38415|AED4ABH Agaricus bisporus DNA for 4-aminobenzoate hyd... 47 3e-04 50 emb|AQ274524|AQ274524 mgxb0022E01r CUGI Rice Blast BAC Library P... 47 3e-04 emb|AW982239|AW982239 HVSMEg0002G20f Hordeum vulgare pre-anthesi... 36 5e-04 emb|AJ274152|AJ274152 AJ274152 Metarhizium anisopliae ARSEF 2575... 41 0.018 emb|Z35859|SCYBL098W S.cerevisiae chromosome II reading frame OR... 39 0.12 emb|X79489|SCDNCH2 S.cerevisiae genomic DNA, chromosome II from ... 39 0.12 emb|AW265977|AW265977 L30-2632T3 Ice plant Lambda Uni-Zap XR exp... 39 0.12 emb|AQ936092|AQ936092 Gm UMb001 014 G18F UMN Soybean BAC Library... emb|AA786833|AA786833 m6d11a1.rl Aspergillus nidulans 24hr asexu... 38 0.23 emb|AV425826|AV425826 AV425826 Lotus japonicus young plants (two... 37 0.42 emb|AB017765|AB017765 Trypanosoma cruzi genes for GATase-CPSase,... 36 0.80 60 emb|AL115728|CNS01CMW Botrytis cinerea strain T4 cDNA library un... 36 0.80 gb|U35892|NHU35892 Nectria haematococca maackiain detoxification... 36 0.80

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•	gb aaa32857.1  (m84658) receptor-like protein kinase [arabidopsis
	thaliana] /blast_score 0
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E E	
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	661,018 sequences; 426,114,510 total letters
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	Sequences producing significant alignments: (bits) Value

emb|AI896318|AI896318 EST265761 tomato callus, TAMU Lycopersicon... 182 5e-65 emb|A67797|A67797 Sequence 2 from Patent WO9743427. gb|U93048|DCU93048 Daucus carota somatic embryogenesis receptor-... 128 5e-51 5 emb|AW432288|AW432288 sh71g05.y1 Gm-c1015 Glycine max cDNA clone... 200 5e-50 emblAI730535lAI730535 BNLGHi7007 Six-day Cotton fiber Gossypium ... 152 1e-49 emb|AF085168|AF085168 Triticum aestivum receptor-like protein ki... 77 2e-46 emb|AF085166|AF085166 Hordeum vulgare receptor-like kinase gene,... 86 2e-45 emb|AF085167|AF085167 Hordeum vulgare receptor-like kinase ARK1A... 85 3e-45 10 gb|U51330|TAU51330 Triticum aestivum leaf rust resistance kinase... 82 8e-45 emblAF100771|AF100771 Hordeum vulgare receptor-like kinase (Hv3A... 100 2e-44 gb|U78762|TAU78762 Triticum aestivum receptor-like kinase ARK1AS... 86 2e-44 emb|AA738545|AA738545 SbRLK3 Sorghum bicolor cv. TX430 leaf Sorg... 129 7e-44 emb|AA738544|AA738544 SbRLK2 Sorghum bicolor cv. TX430 leaf Sorg... 129 1e-43 15 emb|AF085164|AF085164 Hordeum vulgare receptor-like kinase LRK10... 82 2e-43 emb|AW776492|AW776492 EST335557 DSIL Medicago truncatula cDNA cl... 80 7e-43 dbi|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 77 3e-42 emblAB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 86 4e-42 emb|AI771630|AI771630 EST252730 tomato ovary, TAMU Lycopersicon ... 86 5e-42 20 emb|AW621923|AW621923 EST312721 tomato root during/after fruit s... 123 8e-42 emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 87 2e-41 dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 84 2e-41 emb|AI484701|AI484701 EST242962 tomato ovary, TAMU Lycopersicon ... 126 2e-41 gb|U51741|ITU51741 Ipomoea trifida receptor protein kinase 2 (IR... 75 3e-41 25 dbj|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 82 9e-41 gb|U20948|ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 76 3e-40 emb|AB041503|AB041503 Populus nigra PnPK1 mRNA for protein kinas... 72 5e-40 emb|AF142596|AF142596 Nicotiana tabacum LRR receptor-like protei... 108 2e-39 emb|AF220602|AF220602 Lycopersicon pimpinellifolium Rio Grande 7... 91 3e-39 30 emb|Y14285|BOY14285 Brassica oleracea mRNA for SFR1 protein. emb|AB024420|AB024419S2 Brassica oleracea SRK13 gene, exon 2, 3,... 86 5e-39 emb|AW930866|AW930866 EST356709 tomato fruit mature green, TAMU ... 126 1e-38 emb|AW621706|AW621706 EST312504 tomato root during/after fruit s... 122 1e-38 emb|AB012106|AB012106 Brassica rapa mRNA for SRK45, complete cds. 35 emb|AB024422|AB024421S2 Brassica oleracea SRK13-b gene, exon 2, ... 84 2e-38 emb|AB013718|AB013717S2 Brassica rapa gene for SRK46Bra, exon 2.... 84 2e-38 emb|AB041504|AB041504 Populus nigra PnPK2 mRNA for protein kinas... 68 2e-38 gb|BE057261|BE057261 sm99f12.yl Gm-c1015 Glycine max cDNA clone ... 86 3e-38 emb[Y14286]BOY14286 Brassica oleracea SFR3 gene, partial. 40 gb|M97667|BNASTKR Brassica napus ssp. oleifera serine/threonine ... 82 3e-38 emblZ18921BOSRKL B.oleracea gene for S-receptor kinase-like pro... 83 4e-38 emb|AQ989326|AQ989326 Gm ISb001 058 O23R ISU Soybean BAC Library... 94 5e-38 dbj|D31737|TOBPSTK Tobacco mRNA for protein-serine/threonine kin... 75 5e-38 dbj E05046 E05046 DNA encoding ZmPK1 homologue protein in tobacco. 75 5e-38 45 gb|U00443|BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 81 6e-38 emb|AF220603|AF220603 Lycopersicon esculentum VFNT Cherry Pto lo... 91 1e-37 emb|X98520|BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. gb|BE060160|BE060160 HVSMEg0011B12f Hordeum vulgare pre-anthesis... 111 2e-37 emb|AF131222|AF131222 Lophopyrum elongatum protein serine/threon... 110 3e-37 50 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 78 5e-37 emb|AI486547|AI486547 EST244868 tomato ovary, TAMU Lycopersicon ... 100 6e-37 emb|AW687233|AW687233 NF007D09RT1F1077 Developing root Medicago ... 100 7e-37 emb|AW617954|AW617954 EST314028 L. pennellii trichome, Cornell U... 100 9e-37 gb|M76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 78 1e-36 55 emb|AW030223|AW030223 EST273478 tomato callus, TAMU Lycopersicon... 92 2e-36 emblAW929662|AW929662 EST338450 tomato flower buds 8 mm to pre-a... 118 2e-36 emb|AW704997|AW704997 sk41c03.yl Gm-c1019 Glycine max cDNA clone... 89 2e-36 emb|AI967314|AI967314 Ljirnpest00-017 Ljirnp Lambda HybriZap two... 69 2e-36 gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 78 2e-36 60 emb|AJ245479|BNA245479 Brassica napus Sl13, slk, srk, CePP, Fmt.... 82 4e-36 emb|Y14600|SBRLK1 Sorghum bicolor mRNA for protein serine/threon... 122 8e-36

emb|AW687349|AW687349 NF008F08RT1F1074 Developing root Medicago ... 80 9e-36 emblAI487456|AI487456 EST245778 tomato ovary, TAMU Lycopersicon ... 97 1e-35 emb|AW668493|AW668493 GA\_Ea0014C20 Gossypium arboreum 7-10 dpa ... 73 2e-35 emb|AI731504|AI731504 BNLGHi9991 Six-day Cotton fiber Gossypium ... 64 2e-35 5 emb|AW565461|AW565461 LG1\_344\_F11.g1\_A002 Light Grown 1 (LG1) So... .106 4e-35 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 76 4e-35 gb|BE059471|BE059471 sn32d11.yl Gm-c1016 Glycine max cDNA clone ... 115 5e-35 emblAA738546|AA738546 SbRLK4 Sorghum bicolor cv. TX430 leaf Sorg... 120 1e-34 emb|AW760240|AW760240 s159g07.yl Gm-c1027 Glycine max cDNA clone... 76 1e-34 10 emb|AW597214|AW597214 si71g06.yl Gm-c1031 Glycine max cDNA clone... 126 2e-34 emb|AW092144|AW092144 EST285240 tomato mixed elicitor, BTI Lycop... 122 2e-34 emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 74 2e-34 emb[Y18260]BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 74 3e-34 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 74 3e-34 emblAW934655|AW934655 EST353547 tomato flower buds 0-3 mm, Corne... 93 3e-34 15 emb|AI729170|AI729170 BNLGHi12834 Six-day Cotton fiber Gossypium... 67 3e-34 emb|AI896277|AI896277 EST265720 tomato callus, TAMU Lycopersicon... 111 3e-34 emb|AW666082|AW666082 sk31h04.yl Gm-c1028 Glycine max cDNA clone... 116 3e-34 emb|AW233982|AW233982 sf32g05.y1 Gm-c1028 Glycine max cDNA clone... 130 3e-34 20 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. emb|AF197946|AF197946 Glycine max receptor protein kinase-like p... 77 6e-34 emb|AB030083|AB030083 Populus nigra PnLPK mRNA for lectin-like p... 71 6e-34 emb|AW329840|AW329840 N201116e rootphos(-) Medicago truncatula c... 119 9e-34 emb|AW679172|AW679172 WS1\_23\_D02.b1\_A002 Water-stressed 1 (WS1) ... 145\_1e-33 25 emb|AA738547|AA738547 SbRLK5 Sorghum bicolor cv. TX430 leaf Sorg... 115 2e-33 emb|AW267961|AW267961 EST306183 DSIR Medicago truncatula cDNA cl... 76 2e-33 emb|AI736063|AI736063 sb22d04.y1 Gm-c1007 Glycine max cDNA clone... 68 8e-33 emb|AI967315|AI967315 Ljirnpest00-018 Ljirnp Lambda HybriZap two... 75 2e-32 emb|AI823209|AI823209 L30-1092T3 Ice plant Lambda Uni-Zap XR exp... 85 3e-32 30 emb|AF197947|AF197947 Glycine max receptor protein kinase-like p... 72 4e-32 emb|AW684339|AW684339 NF015G04NR1F1000 Nodulated root Medicago t... 79 4e-32 emb|AW329560|AW329560 N200811e rootphos(-) Medicago truncatula c... 95 4e-32 emb|AW031255|AW031255 EST274630 tomato callus, TAMU Lycopersicon... 78 5e-32 gb|BE058691|BE058691 sn19e05.yl Gm-c1016 Glycine max cDNA clone ... 108 5e-32 35 emb|AW031816|AW031816 EST275270 tomato callus, TAMU Lycopersicon... 76 7e-32 emb|AW694732|AW694732 NF079D07ST1F1061 Developing stem Medicago ... 118 7e-32 emb|AI727531|AI727531 BNLGHi8333 Six-day Cotton fiber Gossypium ... 125 7e-32 emb|AW649958|AW649958 EST328412 tomato germinating seedlings, TA... 79 2e-31 emb|AW279355|AW279355 sf65g10.yl Gm-c1013 Glycine max cDNA clone... 118 3e-31 40 emb|AW040672|AW040672 EST283536 tomato mixed elicitor, BTI Lycop... 111 3e-31 Query= atu18770 s at 17083 s at /id source

Query= atu18770\_s\_at 17083\_s\_at /id\_source /description gb|aad25838.1|ac006951\_17 (ac006951) putative indole-3-glycerol phosphate synthase [arabidopsis thaliana] /blast\_score 0 (1362 letters)

Database: plantfungal

50

60

661,018 sequences; 426,114,510 total letters

Searching......done

Score E

55 Sequences producing significant alignments:

(bits) Value

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emb|AI731648|AI731648 BNLGHi10360 Six-day Cotton fiber Gossypium... 182 2e-61 emb|AW458591|AW458591 shl 1b01.yl Gm-c1016 Glycine max cDNA clone... 138 1e-59 emb|AW033644|AW033644 EST277215 tomato callus, TAMU Lycopersicon... 144 1e-51 emb[AW704608]AW704608 sk54c06.yl Gm-c1019 Glycine max cDNA clone... 153 5e-51 emblAW458875|AW458875 sh16b06.yl Gm-c1016 Glycine max cDNA clone... 113 3e-49 5 emb|AW648549|AW648549 EST327003 tomato germinating seedlings, TA... 116 3e-48 gb|BE125163|BE125163 DG1\_13\_H04.b1\_A002 Dark Grown 1 (DG1) Sorgh... 180 2e-44 emb|AW648007|AW648007 EST326461 tomato germinating seedlings, TA... 100 2e-38 10 emb|AI773256|AI773256 EST254356 tomato resistant, Cornell Lycope... 99 5e-38 gb|BE052703|BE052703 GA\_Ea0031N16f Gossypium arboreum 7-10 dpa ... 86 1e-36 emb|AW676692|AW676692 DG1\_13\_H04.g1\_A002 Dark Grown 1 (DG1) Sorg... 105 4e-31 emb|AW758260|AW758260 874006H11.y1 C. reinhardtii CC-1690, Lambd... 132 6e-30 emb|AW564868|AW564868 LG1\_310\_H09.b1\_A002 Light Grown 1 (LG1) So... 104 3e-29 15 emb|AL096874|SPBC1539 S.pombe chromosome II cosmid c1539. emb|AW696155|AW696155 NF103A11ST1F1084 Developing stem Medicago ... 76 6e-29 emb|Y09137|SPTRP1 S.pombe trp-1 mRNA. 57 1e-28 emb|AW618611|AW618611 EST320597 L. pennellii trichome, Cornell U... 82 4e-24 emb|AV420994|AV420994 AV420994 Lotus japonicus young plants (two... 106 3e-22 20 emb|AI490823|AI490823 EST241532 tomato shoot, Cornell Lycopersic... 104 2e-21 emb|AW432225|AW432225 sh70g02.y1 Gm-c1015 Glycine max cDNA clone... 95 2e-18 gb|BE059030|BE059030 sn24c03.y1 Gm-c1016 Glycine max cDNA clone ... 78 1e-13 emb|AU012336|AU012336 AU012336 Schizosaccharomyces pombe late lo... 48 1e-10 emb|X53576|ANTRPCA Aspergillus niger trpC gene for glutamine ami... 64 3e-09 25 emb[X07071]ANTRPC Aspergillus niger trpC gene. 64 3e-09 emb|X05033|PCTRPC Penicillium chrysogenum trpC gene for put. tri... 62 7e-09 emb|X02390|ANTRPC1 Aspergillus nidulans trpC gene. 62 7e-09 gb|M23177|PBLTRPX1A Phycomyces blakesleeanus (TRP1) gene, comple... 61 3e-08 dbj|E00783|E00783 DNA encoding for structural gene MOX(alcoholox... 58 1e-07 30 emb|A11156|A11156 MOX structural gene and it's 5'and 3'-flanking... 58 1e-07 emb|X02425|HPMOXG Hansenula polymorpha MOX gene for methanol oxi... 58 1e-07 emb|X70035|CHTRP1 C.heterostrophus gene for trifunctional trypto... 58 2e-07 gb[M64473]PHTTRP1 Phytophthora parasitica N-(5'-phosphoribosyl)a... 57 3e-07 gblJ01252|NEUTRP1 n.crassa trifunctional tryptophan biosynthesis... 57 3e-07 35 emb|X56047|PDTRPC P. chrysosporium trpC gene for trifunctional p... 55 1e-06 emb[X75951]SC6ORF S.cerevisiae URA1, SAC1, RSD1 and TRP3 genes a... 50 1e-06 gb|K01386|YSCTRP3 Yeast (S.cerevisiae) TRP3 gene coding for anth... 50 1e-06 emb|Z28211|SCYKL211C S.cerevisiae chromosome XI reading frame OR... 50 1e-06 emb|A86135|A86135 Sequence 794 from Patent EP0866129. 40 gb[M74901|CPCTRP1 Cryprococcus neoformans phosphoribosyl anthran... 40 0.042 emb|X75652|ALRIBT A.longa plastid genes for tRNAs, ribosomal pro... 36 0.74 emb[X14385]ALCRPEF Astasia longa chloroplast rps7 and tufA genes... 36 0.74 emb[X51672|SCRSD1 Saccharomyces cerevisiae rsd1 gene. 35 1.0 emb|AL049180|PFMAL13P1 Plasmodium falciparum chromosome 13 strai... 34 2.6 45 gb[U33057|SCD9717 Saccharomyces cerevisiae chromosome IV cosmids... 34 2.6 emblAV423493|AV423493 AV423493 Lotus japonicus young plants (two... 34 3.6 emb|AL353822|NC15E6 Neurospora crassa DNA linkage group V Cosmid... 34 3.6 emb|AW180222|AW180222 MgA0308f MgA Library Mycosphaerella gramin... 34 3.6 emblAJ133598|PSA133598 Pisum sativum chloroplast gF16P gene, exo... 33 5.0 50 emb|AI054479|AI054479 coau0001B20 Cotton Boll Abscission Zone cD... 33 5.0 gb|L34806|PEAFRUBISP Pisum sativum fructose-1,6-bisphosphate mRN... 33 5.0 emb|X68826|PSF16B P.sativum mRNA for fructose 1,6 biphosphatase. emb|AV391683|AV391683 AV391683 Chlamydomonas reinhardtii C9 Chla... 33 5.0 emb|AJ231281|BSAJ1281 Brachythecium salebrosum partial mRNA for ... 33 6.8 55 emb|AQ902358|AQ902358 LMAJFV1\_lm85h07.x1 Leishmania major FV1 ra... 33 6.8 emb|AQ654520|AQ654520 Sheared DNA-10A10.TF Sheared DNA Trypanoso... 32 9.3 emb|AF180022|AF180022 Adiantum capillus-veneris photosystem I P7... 32 9.3 emb|AQ649354|AQ649354 Sheared DNA-30F1.TF Sheared DNA Trypanosom... 32 9.3 gb|L44013|BLYCH Hordeum vulgare (clone WG241) STS mRNA, sequence... 32 9.3 60 emb|AA451566|AA451566 AOB168f Onion seedling leaf cDNA library A... 32 9.3 dbj|D87301|D87301 D87301 Trypanosoma cruzi Y (Tomoo Tanaka) Tryp... 28 10.0

Query= atu66345\_s\_at 17097 s\_at /id\_source genbank /description gb|aac49697.1| (u66345) calreticulin [arabidopsis thaliana] /blast score 0 5 (1424 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Searching......done Score E Sequences producing significant alignments: (bits) Value 15 emb|AJ002057|BVMRNAC Beta vulgaris mRNA for calreticulin. 577 e-163 emb|Z71395|NPCAL1MNR N.plumbaginifolia mRNA for calreticulin. 575 e-163 gb|U74630|RCU74630 Ricinus communis calreticulin mRNA, complete ... 575 e-163 emb|AF052040|AF052040 Berberis stolonifera calreticulin mRNA, co... 571 e-162 emb|X85382|NTRNATCAL N.tabacum mRNA for calreticulin. 20 emb|AF134733|AF134733 Prunus armeniaca calcium-binding protein c... 565 e-160 gb|L27349|BLYCRH2A Hordeum vulgare calreticulin (CRH2) mRNA, par... 560 e-159 gb|L27348|BLYCRH1A Hordeum vulgare calreticulin (CRH1) mRNA, par... 560 e-159 emb|AF019376|AF019376 Brassica napus calreticulin mRNA, complete... 465 e-156 emb|X80756|CAPCRTC C.annuum PCRTC mRNA. 25 gb|U74631|RCU74631 Ricinus communis calreticulin gene, complete ... 149 e-123 emb|AJ000765|CRAJ765 Chlamydomonas reinhardtii mRNA for calretic... 369 e-122 emb|AI728389|AI728389 BNLGHi10665 Six-day Cotton fiber Gossypium... 438 e-122 emb|AI782264|AI782264 EST263143 tomato susceptible, Cornell Lyco... 418 e-116 emb|AB018243|AB018243 Solanum melongena EEF22 mRNA for calreticu... 402 e-111 30 emb|AW667951|AW667951 GA\_\_Ea0012A11 Gossypium arboreum 7-10 dpa ... 402 e-111 emb|AW216727|AW216727 EST295441 tomato callus, TAMU Lycopersicon... 391 e-108 emb|AW509432|AW509432 si37b03.y1 Gm-r1030 Glycine max cDNA clone... 389 e-107 emb|AW039860|AW039860 EST282333 tomato mixed elicitor, BTI Lycop... 382 e-105 emb|AW624478|AW624478 EST322423 tomato flower buds 3-8 mm, Corne... 378 e-104 35 emb|AW626316|AW626316 EST320223 tomato radicle, 5 d post-imbibit... 375 e-103 emb|AW622049|AW622049 EST312847 tomato root during/after fruit s... 363 3e-99 emb|AW568477|AW568477 si59c07.yl Gm-r1030 Glycine max cDNA clone... 361 9e-99 emb|AW441195|AW441195 EST310591 tomato fruit red ripe, TAMU Lyco... 359 2e-98 40 gb|BE059929|BE059929 sn38h08.yl Gm-c1016 Glycine max cDNA clone ... 350 1e-95 emb|AW944954|AW944954 EST337004 tomato flower buds 3-8 mm, Corne... 349 3e-95 emb|AW932476|AW932476 EST358319 tomato fruit mature green, TAMU ... 348 5e-95 emb|AW930392|AW930392 EST340945 tomato fruit mature green, TAMU ... 348 7e-95 emb|AW649196|AW649196 EST327650 tomato germinating seedlings, TA... 347 2e-94 45 emb|AW650460|AW650460 EST328914 tomato germinating seedlings, TA... 344 8e-94 emb|AW560421|AW560421 EST315469 DSIR Medicago truncatula cDNA cl... 330 1e-93 emb|AW650947|AW650947 EST329401 tomato germinating seedlings, TA... 343 2e-93 emb[AW685878]AW685878 NF031C11NR1F1000 Nodulated root Medicago t... 341 6e-93 emb|AW756722|AW756722 sl26d08.yl Gm-c1027 Glycine max cDNA clone... 336 2e-91 emb|AW934135|AW934135 EST359978 tomato fruit mature green, TAMU ... 333 2e-90 50 emb|AW621571|AW621571 EST312369 tomato root during/after fruit s... 333 2e-90 emb|Y09816|EGCALRPR E.gracilis mRNA for calreticulin precursor. emblAW731454|AW731454 GA Ea0030H17 Gossypium arboreum 7-10 dpa ... 329 4e-89 emb|AI960982|AI960982 sc93e09.yl Gm-c1019 Glycine max cDNA clone... 328 6e-89 55 emb|AW725609|AW725609 GA\_ Ea0018P14 Gossypium arboreum 7-10 dpa ... 326 3e-88 emb|AW033447|AW033447 EST277018 tomato callus, TAMU Lycopersicon... 323 2e-87 emb|AW201052|AW201052 se97c10.yl Gm-c1027 Glycine max cDNA clone... 316 2e-85 60 emb|AW773947|AW773947 EST332933 KV3 Medicago truncatula cDNA clo... 316 2e-85

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60

Database: plantfungal 661,018 sequences: 426,114,510 total letters Searching.....done Ε Score Sequences producing significant alignments: (bits) Value gb|U70666|BNU70666 Brassica napus pathogenesis-related protein P... 323 3e-89 10 gb|U21849|BNU21849 Brassica napus PR-1a (LSC94) mRNA, complete cds. 323 3e-89 gb|U64806|BNU64806 Brassica napus pathogenesis-related protein P... 209 4e-86 emb|AI352851|AI352851 MB69-8A PZ204.BNlib Brassica napus cDNA cl... 311 5e-84 emb|AI352893|AI352893 MB72-6D PZ204.BNlib Brassica napus cDNA cl... 165 3e-62 emb|AW217013|AW217013 EST295727 tomato callus, TAMU Lycopersicon... 152 4e-60 15 emblAW219671|AW219671 EST302153 tomato root during/after fruit s... 151 1e-59 emb|AW092403|AW092403 EST285583 tomato mixed elicitor, BTI Lycop... 151 1e-59 emb|AI895090|AI895090 EST264533 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW625930|AW625930 EST319825 tomato radicle, 5 d post-imbibit... 151 1e-59 emb|AW032514|AW032514 EST276073 tomato callus, TAMU Lycopersicon... 151 1e-59 20 emb|AW034260|AW034260 EST277831 tomato callus, TAMU Lycopersicon... 151 1e-59 emblAW040983|AW040983 EST283847 tomato mixed elicitor, BTI Lycop... 151 1e-59 emblAW034206|AW034206 EST277777 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW219480|AW219480 EST301878 tomato root during/after fruit s... 151 1e-59 emb|AW092623|AW092623 EST285803 tomato mixed elicitor, BTI Lycop... 151 1e-59 25 emb|AW040954|AW040954 EST283818 tomato mixed elicitor, BTI Lycop... 151 1e-59 emblAW034454|AW034454 EST278025 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW032723|AW032723 EST276282 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW033873|AW033873 EST277444 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AI894650|AI894650 EST264093 tomato callus, TAMU Lycopersicon... 151 1e-59 30 emb|AW033593|AW033593 EST277164 tomato callus, TAMU Lycopersicon... 151 3e-59 emb|AW034724|AW034724 EST278526 tomato callus, TAMU Lycopersicon... 148 7e-59 emb|AW041033|AW041033 EST283897 tomato mixed elicitor, BTI Lycop... 148 7e-59 emb|X66942|NTPRB1B N.tabacum prb-1b gene. 146, 9e-59 emblAW559895|AW559895 EST314943 DSIR Medicago truncatula cDNA cl... 160 1e-58 35 emb|AW329241|AW329241 N200453e rootphos(-) Medicago truncatula c... 160 1e-58 emb|AI352771|AI352771 MB58-5G PZ204.BNlib Brassica napus cDNA cl... 220 2e-58 emb|AW559894|AW559894 EST314942 DSIR Medicago truncatula cDNA cl... 159 5e-58 emblAW035820|AW035820 EST281974 tomato callus, TAMU Lycopersicon... 145 6e-58 emb|AW032139|AW032139 EST275593 tomato callus, TAMU Lycopersicon... 145 6e-58 40 emb|AW034330|AW034330 EST277901 tomato callus, TAMU Lycopersicon... 144 2e-57 emb|AW622143|AW622143 EST312941 tomato root during/after fruit s... 149 2e-57 emb|AW032749|AW032749 EST276308 tomato callus, TAMU Lycopersicon... 149 3e-57 emb|AI894391|AI894391 EST263846 tomato callus, TAMU Lycopersicon... 142 4e-57 emb|AI896249|AI896249 EST265692 tomato callus, TAMU Lycopersicon... 142 6e-57 45 emblAI782621|AI782621 EST263500 tomato susceptible, Cornell Lyco... 144 8e-57 emb|AW126362|AW126362 N100469e rootphos(-) Medicago truncatula c... 160 3e-56 emb|AW981257|AW981257 EST392410 DSIL Medicago truncatula cDNA cl... 128 1e-55 emb|AW033687|AW033687 EST277258 tomato callus, TAMU Lycopersicon... 149 2e-55 emb|AI352783|AI352783 MB59-12D PZ204.BNlib Brassica napus cDNA c... 198 3e-55 50 emb[X14065]NTPRP1 Nicotiana tabacum gene for basic form of patho... 140 5e-55 emb|AW625666|AW625666 EST319573 tomato radicle, 5 d post-imbibit... 135 6e-55 emblAW032727|AW032727 EST276286 tomato callus, TAMU Lycopersicon... 147 1e-53 emb|X52555|NTW381 Tobacco W38/1 gene for PR-1 pathogenesis-relat... 116 1e-52 gb|BE034214|BE034214 MH01C07 MH Mesembryanthemum crystallinum cD... 112 4e-52 emb|AW053720|AW053720 L30-1612T3 Ice plant Lambda Uni-Zap XR exp... 112 4e-52 emb|AW559969|AW559969 EST315017 DSIR Medicago truncatula cDNA cl... 81 4e-52 emb|AI352801|AI352801 MB61-9C PZ204.BNlib Brassica napus cDNA cl... 194 1e-51 emb|X17681|NTPR1CA Tobacco gene for pathogenesis-related protein... 111 2e-51 emb|X05454|NTPR1CR Nicotiana tabacum mRNA for PR-1c protein. 60 emb|X12487|NTPR1C Tobacco mRNA fragment for pathogenesis-related... 111 2e-51

emb|AJ011520|LES011520 Lycopersicon esculentum prla (P4) gene.

	gb M69247 TOMPRP4 Lycopersicon esculentum PR (pathogenesis relat 98 1e-50 emb A22634 LEPI4GENE L.esculentum P14 gene. 98 1e-50 emb AJ250136 STU250136 Solanum tuberosum mRNA for pathogenesis r 99 2e-50
5	emb AW218808 AW218808 EST301288 tomato root during/after fruit s 116 3e-50 emb AI896011 AI896011 EST265454 tomato callus, TAMU Lycopersicon 116 3e-50 emb AW094536 AW094536 EST287716 tomato mixed elicitor, BTI Lycop 116 3e-50 emb AW038553 AW038553 EST280236 tomato mixed elicitor, BTI Lycop 116 3e-50
•	dbj D90196 TOBPR1A Nicotiana tabacum mRNA for PR1a protein precu 104 3e-50 emb X05959 NTPR1AG Tobacco PR-1a gene for pathogenesis-related p 104 4e-50
10	emb X12737 NTPR1A1 Tobacco PR-1a gene for pathogenesis-related p 104 4e-50
	emb X06361 NTPR1AG1 Nicotiana tabacum gene for pathogenesis-rela 104 4e-50 emb X12485 NTPR1A Tobacco mRNA fragment for pathogenesis-related 104 4e-50
	emb AW033469 AW033469 EST277040 tomato callus, TAMU Lycopersicon 116 5e-50
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15	emb AI352858 AI352858 MB70-10D PZ204.BNlib Brassica napus cDNA c 197 7e-50 emb AI782416 AI782416 EST263295 tomato susceptible, Cornell Lyco 95 7e-50
	emb AW034882 AW034882 EST279111 tomato callus, TAMU Lycopersicon 95 1e-49
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	emb[X17680]NTPR1BA Tobacco gene for pathogenesis-related protein 103 4e-49 emb[X03465]NTPRPR Nicotiana tabacum mRNA for pathogenesis-relate 103 4e-49
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•	gb M69248 TOMPRP6 Lycopersicon esculentum PR (pathogenesis relat 92 7e-49 emb X68738 LEP1P14A L.esculentum mRNA for ethylene-induced P1(p1 92 7e-49
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	emb AI779425 AI779425 EST260304 tomato susceptible, Cornell Lyco 92 7e-49
	emb AI782545 AI782545 EST263424 tomato susceptible, Cornell Lyco 92 7e-49 emb AI782822 AI782822 EST263701 tomato susceptible, Cornell Lyco 92 7e-49
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35	emb AI778680 AI778680 EST259559 tomato susceptible, Cornell Lyco 92 7e-49
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	, , , , , , , , , , , , , , , , , , ,
50	Query= af107726_s_at 17499_s_at /id_source genbank /description
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60	Score E
	Sequences producing significant alignments: (bits) Value

emb|AF079872|AF079872 Nicotiana tabacum cyclic nucleotide-gated ... 380 0.0 emb|AF079871|AF079871 Nicotiana tabacum cyclic nucleotide-gated ... 373 0.0 emb|AJ002610|HVAJ2610 Hordeum vulgare mRNA for putative calmodul... 331 0.0 emb|AW625647|AW625647 EST319554 tomato radicle, 5 d post-imbibit... 255 6e-82 emb|AW038928|AW038928 EST280884 tomato mixed elicitor, BTI Lycop... 190 2e-80 emb|AI782780|AI782780 EST263659 tomato susceptible, Cornell Lyco... 259 4e-77 emb|AI486671|AI486671 EST244993 tomato ovary, TAMU Lycopersicon ... 241 4e-70 emb|AW725576|AW725576 GA Ea0018M08 Gossypium arboreum 7-10 dpa ... 196 3e-69 10 emb|AW265987|AW265987 L30-2643T3 Ice plant Lambda Uni-Zap XR exp... 150 3e-69 emb[AW033272[AW033272 EST276843 tomato callus, TAMU Lycopersicon... 260 2e-68 emb|AW126067|AW126067 N100263e rootphos(-) Medicago truncatula c... 258 1e-67 gb[U65390]NTU65390 Nicotiana tabacum CaMB-channel protein mRNA, ... 162 le-65 emb|AI898047|AI898047 EST267490 tomato ovary, TAMU Lycopersicon ... 241 2e-62 15 emb|AI487755|AI487755 EST246077 tomato ovary, TAMU Lycopersicon ... 241 2e-62 emb|AI776725|AI776725 EST257825 tomato resistant, Cornell Lycope... 240 3e-62 emb|AI729234|AI729234 BNLGHi12947 Six-day Cotton fiber Gossypium... 176 3e-57 emb|AW705372|AW705372 sk48b11.yl Gm-c1019 Glycine max cDNA clone... 133 4e-51 emb|AW029776|AW029776 EST273031 tomato callus, TAMU Lycopersicon... 156 4e-51 20 emb|AW220952|AW220952 EST297421 tomato fruit mature green, TAMU ... 118 3e-50 emb|AW626197|AW626197 EST320104 tomato radicle, 5 d post-imbibit... 132 1e-49 gb|BE059468|BE059468 sn32d07.yl Gm-c1016 Glycine max cDNA clone ... 133 5e-46 emblAW029909|AW029909 EST273164 tomato callus, TAMU Lycopersicon... 122 5e-45 emb|AI729590|AI729590 BNLGHi13704 Six-day Cotton fiber Gossypium... 180 4e-44 25 emb|AI490589|AI490589 EST249143 tomato ovary, TAMU Lycopersicon ... 178 1e-43 emb|AW830981|AW830981 sm31b10.y1 Gm-c1028 Glycine max cDNA clone... 176 8e-43 emb|AV414437|AV414437 AV414437 Lotus japonicus young plants (two... 112 2e-41 emb|AI494870|AI494870 sb06e03.yl Gm-c1004 Glycine max cDNA clone... 125 1e-40 emb|AI055360|AI055360 coau0003M11 Cotton Boll Abscission Zone cD... 123 8e-39 30 emb|AW737921|AW737921 EST339348 tomato flower buds, anthesis, Co... 128 1e-37 emb|AI967361|AI967361 Ljimpest02-095-a3 Ljimp Lambda HybriZap ... 94 2e-36 emb|AI776645|AI776645 EST257745 tomato resistant, Cornell Lycope... 70 3e-35 emb|AW775519|AW775519 EST334584 DSIL Medicago truncatula cDNA cl... 147 3e-34 emb|AI778171|AI778171 EST259050 tomato susceptible, Cornell Lyco... 76 4e-34 35 emb|AW309771|AW309771 sf24f01.x1 Gm-c1028 Glycine max cDNA clone... 143 7e-33 emb|AI960258|AI960258 sc81a02.y1 Gm-c1018 Glycine max cDNA clone... 122 2e-32 emb|AW563840|AW563840 LG1 272\_B06.b1\_A002 Light Grown 1 (LG1) So... 113 1e-30 emb|AW774513|AW774513 EST333664 KV3 Medicago truncatula cDNA clo... 135 1e-30 emb|AI967666|AI967666 Ljimpest08-671-e8 Ljimp Lambda HybriZap ... 116 2e-30 40. emb|AW671345|AW671345 LG1\_335\_D05.b1\_A002 Light Grown 1 (LG1) So... 100 2e-30 gb|C95565|C95565 C95565 Citrus unshiu Miyagawa-wase maturation s... 132 1e-29 emb|AW759050|AW759050 sl34a04.yl Gm-c1027 Glycine max cDNA clone... 129 9e-29 emb|AW286638|AW286638 LG1\_335\_D05.g1\_A002 Light Grown 1 (LG1) So... 127 2e-28 emb|AW831384|AW831384 sm24g05.y1 Gm-c1028 Glycine max cDNA clone... 87 9e-27 45 emb|AW037755|AW037755 EST279384 tomato mixed elicitor, BTI Lycop... 122 1e-26 emb|AW508715|AW508715 si35e10.yl Gm-r1030 Glycine max cDNA clone... 117 3e-25 emb|AW350588|AW350588 GM210009A10H2R Gm-r1021 Glycine max cDNA 3... 106 1eemb|AW350847|AW350847 GM210009B20G5R Gm-r1021 Glycine max cDNA 3... 106 2e-50 emb|AI776960|AI776960 EST251986 tomato callus, TAMU Lycopersicon... 111 2e-23 emb|AW781088|AW781088 sl88h09.yl Gm-c1037 Glycine max cDNA clone... 111 3e-23 emb|AW039011|AW039011 EST280984 tomato mixed elicitor, BTI Lycop... 111 3e-23 emb|AW617273|AW617273 EST323684 L. hirsutum trichome, Cornell Un... 106 7e-22 55 gb|BE022211|BE022211 sm72b10.yl Gm-c1028 Glycine max cDNA clone ... 106 7e-22 emb|AW668188|AW668188 GA\_\_Ea0013B18 Gossypium arboreum 7-10 dpa ... 105 2e-21 emb|AA824914|AA824914 CT202.SK Tomato Leaf cDNA from cv. VFNT ch... 93 6e-21 emb|AT000374|AT000374 AT000374 Apple peel cDNA library Malus x d... 102 1e-20 emb|AW623583|AW623583 EST321528 tomato flower buds 3-8 mm, Corne... 69 7e-20 60 emb|AI495394|AI495394 sa97d10.y1 Gm-c1004 Glycine max cDNA clone... 98 2e-19 emb|AI725563|AI725563 BNLGHi12127 Six-day Cotton fiber Gossypium... 67 5e-18

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	emb AC016528 AC016528 Leishmania major chromosome 35 clone L4123 35 1.7
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	Score E
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	emb AW102460 AW102460 sd88d10.y1 Gm-c1009 Glycine max cDNA clone 70 2e-22
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	emb AI487362 AI487362 EST245684 tomato ovary, TAMU Lycopersicon 72 3e-20
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•	emb AL115248 CNS01C9K Botrytis cinerea strain T4 cDNA library un 65 4e-18
•	dbj[D10521]TETCALW T.pyriformis mRNA for calmodulin. 73 7e-18
	gb]M76407 SLECALMODU Stylonychia lemnae calmodulin gene, complet 74 9e-18
	emb AF007889 AF007889 Symbiodinium microadriaticum calmodulin (S 74 le-17
10	emb AF078679 AF078679 Olea europaea calcium-binding protein (PCA 57 2e-17
	emb[X56511]TBCALUBG T. brucei genes for calmodulin, EFH5 and ubi 77 2e-17
	emb[X52096]TCCALB2 Trypanosoma cruzi CalA2 calmodulin gene. 77 2e-17
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	gb[M83535]PHTCALPIA P.infestans calmodulin (calA) gene, complete 75 2e-17 emb[Y08373]TGCM T.gondii mRNA for calmodulin. 74 2e-17
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20	gb C96396 C96396 C96396 Marchantia polymorpha immature sex organ 73 3e-17
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	emb AW625406 AW625406 EST319229 tomato radicle, 5 d post-imbibit 62 1e-16
40	gb K02944 TRBCMRSG Trypanosoma brucei gambiense calmodulin genes 73 2e-16 gb M88307 BNACALM Brassica juncea calmodulin mRNA, complete cds. 73 2e-16
TU	gb M88307 BNACALM Brassica juncea calmodulin mRNA, complete cds. 73 2e-16 gb U10150 BNU10150 Brassica napus Naehan calmodulin (bcm1) mRNA, 73 2e-16
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45	emb AW101324 AW101324 sd77e09.yl Gm-c1009 Glycine max cDNA clone 59 2e-16 emb AL112713 CNS01AB5 Botrytis cinerea strain T4 cDNA library un 73 2e-16
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	1 17740 4-4-1-1 - 114-4-4-4-4-4-4-4-4-4-4-4-4-4
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50	A Improved the contract of the
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Sequences producing significant alignments:

(bits) Value

gb|U64925|NTU64925 Nicotiana tabacum geranylgeranylated protein ... 260 1e-68 emb|AW625701|AW625701 EST319608 tomato radicle, 5 d post-imbibit... 228 6e-59 emb|AW720227|AW720227 LjNEST17c4r Lotus japonicus nodule library... 219 4e-56 emb|AW220184|AW220184 EST302667 tomato root during/after fruit s... 208 7e-53 emb|AI774580|AI774580 EST255680 tomato resistant, Cornell Lycope... 173 2e-42 40 emb|AW685484|AW685484 NF030E02NR1F1000 Nodulated root Medicago t... 171 2e-41 emblAI443867|AI443867 sa44d09.yl Gm-c1004 Glycine max cDNA clone... 170 3e-41 emb|AW397252|AW397252 sg76f06.yl Gm-c1007 Glycine max cDNA clone... 168 1e-40 emb|AW033368|AW033368 EST276939 tomato callus, TAMU Lycopersicon... 162 5e-39 emb|AV417858|AV417858 AV417858 Lotus japonicus young plants (two... 153 4e-36 45 emb|AI780050|AI780050 EST260929 tomato susceptible, Cornell Lyco... 138 9e-32 emb|AW039095|AW039095 EST281068 tomato mixed elicitor, BTI Lycop... 111 2e-23 emb|AI780139|AI780139 EST261018 tomato susceptible, Cornell Lyco... 86 7e-16 emb|AW164180|AW164180 Ljirnpest21-672-c8 Ljirnp Lambda HybriZap ... 84 2e-15 emblAI781596|AI781596 EST262475 tomato susceptible, Cornell Lyco... 75 1e-12 50 emb|AW774764|AW774764 EST333915 KV3 Medicago truncatula cDNA clo... 62 7e-09 emb|AW348781|AW348781 GM210003A22H11R Gm-r1021 Glycine max cDNA ... 54 2e-06 emb|AI491210|AI491210 EST241919 tomato shoot, Cornell Lycopersic... 29 0.010 emb|AW651526|AW651526 EST329980 tomato germinating seedlings, TA... 29 0.011 emb|AW220594|AW220594 EST296979 tomato fruit mature green, TAMU ... 29 0.011 55 emb|AW926585|AW926585 HVSMEg0007J19 Hordeum vulgare pre-anthesis... 29 0.025 emb|AW032321|AW032321 EST275775 tomato callus, TAMU Lycopersicon... 28 0.026 emb|AW736598|AW736598 EST333090 KV3 Medicago truncatula cDNA clo... 37 0.39 emb|AW736597|AW736597 EST333089 KV3 Medicago truncatula cDNA clo... 37 0.39 emb|AW713727|AW713727 h1f12ne.fl Neurospora crassa evening cDNA ... 35 1.0 60 emb|AW711542|AW711542 f3g07ne.f1 Neurospora crassa evening cDNA ... 35 1.0 emb|AB009972|AB009972 Aspergillus oryzae gene for beta-1,4-xylos... 35 1.0

	emb AW713709 AW713709 h1e09ne.fl Neurospora crassa evening cDNA 35 1.0
	emb AW712721 AW712721 g3a07ne.fl Neurospora crassa evening cDNA 35 1.0
	emb AW709185 AW709185 d3e02ne.fl Neurospora crassa evening cDNA 35 1.0
_	emb Y09354 SPABC1 S.pombe ABC1 gene. 35 1.4
5	emb Z99262 SPAC9E9 S.pombe chromosome I cosmid c9E9. 35 1.4
	emb AJ225108 STA225108 Solanum tuberosum (cultivar Bintje) mitoc 35 1.4
	emb AW933326 AW933326 EST359169 tomato fruit mature green, TAMU 26 1.5
	emb Z25870 CACDC10G C.albicans CDC10 gene for cell division cycl 31 1.5
10	emb AC008368 AC008368 Trypanosoma brucei chromosome II clone RPC 35 1.9
10	emb AQ942780 AQ942780 Sheared DNA-42B15.TR Sheared DNA Trypanoso 35 1.9
	emb AQ953508 AQ953508 Sheared DNA-33G8.TR Sheared DNA Trypanosom 35 1.9
	emb AC009463 AC009463 Trypanosoma brucei chromosome II clone RPC 35 1.9
	emb AQ950237 AQ950237 Sheared DNA-39C11.TF Sheared DNA Trypanoso 35 1.9
	emb AQ643883 AQ643883 RPCI93-DpnII-26C13.TV RPCI93-DpnII Trypano 35 1.9
1.5	
15	emb AQ948491 AQ948491 Sheared DNA-49H2.TF Sheared DNA Trypanosom 35 1.5
	emb AW729536 AW729536 GA_Ea0025E24 Gossypium arboreum 7-10 dpa 35 1.9
	emb AW727289 AW727289 GA
	gb L36856 PEAIAP34A Pisum sativum GTP-binding protein (IAP34) mR 28 2.1
	emb Z28341 PSCLOEP P.sativum (miranda) mRNA for chloroplast oute 28 2.1
20	
20	emb AW776020 AW776020 EST335085 DSIL Medicago truncatula cDNA cl 28 2.2
	emb AW685649 AW685649 NF032G04NR1F1000 Nodulated root Medicago t 28 2.2
	emb AW690755 AW690755 NF037H10ST1F1000 Developing stem Medicago 28 2.2
•	emb AW256896 AW256896 EST305033 KV2 Medicago truncatula cDNA clo 28 2.3
	emb AV422565 AV422565 AV422565 Lotus japonicus young plants (two 28 2.3
25	emb AW832303 AW832303 sm07c04.y1 Gm-c1027 Glycine max cDNA clone 27 2.3
23	
	emb AL136538 SPAC30 S.pombe chromosome I cosmid c30. 34 2.6
	emb A1724721 A1724721 RHIZ1_26_C05.y2_A001 Rhizomel Sorghum hale 34 2.6
	emb AW256609 AW256609 EST304746 KV2 Medicago truncatula cDNA clo 34 2.6
	emb AI775696 AI775696 EST256796 tomato resistant, Cornell Lycope 28 3.1
30	emb Z71682 SCYNR067C S.cerevisiae chromosome XIV reading frame O 34 3.6
	emb AI776315 AI776315 EST257415 tomato resistant, Cornell Lycope 34 3.6
	emb AF230371 AF230371 Lycopersicon esculentum allene oxide synth 34 3.6
	emb AA741645 AA741645 LmLv39p3/255A Leishmania major promastigot 34 3.6
	gb U08843 PPU08843 Porphyra purpurea putative polysaccharide bin 34 3.6
35	emb AF124792 AF124792 Sporothrix schenckii protein kinase C (PCK 29 4.7
	emb AQ399149 AQ399149 mgxb0019C01f CUGI Rice Blast BAC Library P 33 5.0
	emb AW615911 AW615911 EST325409 tomato flower buds 0-3 mm, Corne 33 5.0
	emb AW035453 AW035453 EST281191 tomato callus, TAMU Lycopersicon 33 5.0
	emb AF051695 AF051695 Trypanosoma cruzi sialidase homolog (P85.1 33 5.0
40	
40	emb Z37538 LTGRR4 L.tarentolae mRNA encoding putative NADH subun 28 5.5
	emb X85021 SCXLTORFS S.cerevisiae DNA from left arm of chromosom 33 6.8
	gb L25088 YSCMEF2MIT Saccharomyces cerevisiae nuclear-encoded mi 33 6.8
	emb AQ443826 AQ443826 GSSTc01287 Trypanosoma cruzi random genomi 33 6.8
	emb AQ502942 AQ502942 V47H4 mTn-3xHA/lacZ Insertion Library Sacc 33 6.8
45	emb Z85962 MSZ85962 Musa sp. DNA for sequence tagged microsatell 33 6.8
	emb Z49377 SCYJL102W S.cerevisiae chromosome X reading frame ORF 33 6.8
	emb AW224537 AW224537 EST302980 tomato root, plants pre-anthesis 31 7.5
	emb AW257183 AW257183 EST305320 KV2 Medicago truncatula cDNA clo 32 9.4
	emb AQ642922 AQ642922 RPCI93-DpnII-27E7.TV RPCI93-DpnII Trypanos 32 9.4
50	emb AW738509 AW738509 EST339936 tomato flower buds, anthesis, Co 32 9.4
	emb AW217351 AW217351 EST296174 tomato flower buds 0-3 mm, Corne 32 9.4
	emb AW219255 AW219255 EST301737 tomato root during/after fruit s 32 9.4
	emb AC007865 AC007865 Trypanosoma brucei chromosome II clone RPC 32 9.4
	emb AW684118 AW684118 NF012F02NR1F1000 Nodulated root Medicago t 32 9.4
55	emb AW622239 AW622239 EST313037 tomato root during/after fruit s 32 9.4
	emb AQ651543 AQ651543 Sheared DNA-7C22.TF Sheared DNA Trypanosom 32 9.4
	emb[Y11565]NC11565 N.crassa acu-15 gene. 32 9.4
	emb AW775944 AW775944 EST335009 DSIL Medicago truncatula cDNA cl 32 9.4
cΛ	emb AB014493 AB014493 Gibberella zeae gene for reductase, partia 32 9.4
60	

"emb|cab59428.1| (aj002295) inositol-1,4,5-trisphosphate 5-phosphatase [arabidopsis thaliana]" /blast\_score 0 (2083 letters)

5 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 10 Score E Sequences producing significant alignments: (bits) Value emb|AI898446|AI898446 EST267889 tomato ovary, TAMU Lycopersicon ... 209 e-107 emb|AW217248|AW217248 EST295962 tomato callus, TAMU Lycopersicon... 245 1e-88 15 emb|AI485780|AI485780 EST244101 tomato ovary, TAMU Lycopersicon ... 242 1e-85 emb|AI485695|AI485695 EST244016 tomato ovary, TAMU Lycopersicon ... 261 3e-85 emb|AI485239|AI485239 EST243543 tomato ovary, TAMU Lycopersicon ... 291 1e-77 emb|AI899197|AI899197 EST268640 tomato ovary, TAMU Lycopersicon ... 161 1e-77 emb|AW034573|AW034573 EST278257 tomato callus, TAMU Lycopersicon... 272 8e-72 20 emb|AI488812|AI488812 EST247151 tomato ovary, TAMU Lycopersicon ... 251 1e-65 emblAW687082|AW687082 NF005G09RT1F1071 Developing root Medicago ... 182 6e-65 emb|AI898248|AI898248 EST267691 tomato ovary, TAMU Lycopersicon ... 204 1e-63 emb|AI483614|AI483614 EST249464 tomato ovary, TAMU Lycopersicon ... 238 1e-61 emb|A1897089|A1897089 EST266532 tomato ovary, TAMU Lycopersicon ... 226 7e-58 25 emb|AW774994|AW774994 EST334145 KV3 Medicago truncatula cDNA clo... 182 2e-56 emb|AW559604|AW559604 EST314652 DSIR Medicago truncatula cDNA cl... 177 3e-53 emb|AI485284|AI485284 EST243588 tomato ovary, TAMU Lycopersicon ... 204 3e-51 emb|AI485664|AI485664 EST243985 tomato ovary, TAMU Lycopersicon ... 195 8e-49 emb|AW774741|AW774741 EST333892 KV3 Medicago truncatula cDNA clo... 90 2e-48 30 emb|AI487608|AI487608 EST245930 tomato ovary, TAMU Lycopersicon ... 187 3e-46 emb|AI483438|AI483438 EST249259 tomato ovary, TAMU Lycopersicon ... 169 6e-41 emb|AW775688|AW775688 EST334753 DSIL Medicago truncatula cDNA cl... 157 2e-37 emb|AI485008|AI485008 EST243271 tomato ovary, TAMU Lycopersicon ... 157 2e-37 emb|AI938737|AI938737 sb58c06.yl Gm-c1018 Glycine max cDNA clone... 151 2e-35 35 emb|AI967736|AI967736 Ljirnpest11-837-a7 Ljirnp Lambda HybriZap ... 147 4e-34 emb|Z50161|SCC14ORFS S.cerevisiae orfs and LEU4, MET4, POL1, RAS... 60 1e-33 emb|Z71382|SCYNL106C S.cerevisiae chromosome XIV reading frame O... 60 1e-33 emblAI973618|AI973618 sd07d04.yl Gm-c1020 Glycine max cDNA clone... 141 2e-32 emb|AI485392|AI485392 EST243713 tomato ovary, TAMU Lycopersicon ... 138 2e-31 40 emb[X94335]SC130KBXV S.cerevisiae 130kb DNA fragment from chromo... 51 1e-29 emb|Z75017|SCYOR109W S.cerevisiae chromosome XV reading frame OR... 51 1e-29 emb|AL022103|SPBC2G2 S.pombe chromosome II cosmid c2G2. 79 le-27 emb[X79743]SCBPPS S.cerevisiae BET1, PAN1 PRI1 and STS1 genes. 72 2e-27 emb|Z38062|SC9687 S.cerevisiae chromosome IX cosmid 9687. 72 2e-27 45 emb|AI488810|AI488810 EST247149 tomato ovary, TAMU Lycopersicon ... 121 3e-26 gb|BE020060|BE020060 sm38e06.yl Gm-c1028 Glycine max cDNA clone ... 119 9e-26 emb|AW685248|AW685248 NF028A10NR1F1000 Nodulated root Medicago t... 116 2e-24 emb|AW257207|AW257207 EST305344 KV2 Medicago truncatula cDNA clo... 75 2e-24 emblAV406995|AV406995 AV406995 Lotus japonicus young plants (two... 67 2e-23 50 emb|AW034093|AW034093 EST277588 tomato callus, TAMU Lycopersicon... 111 3e-23 emb|Z98763|SPAC9G1 S.pombe chromosome I cosmid c9G1. emb|AQ502761|AQ502761 V84A11 mTn-3xHA/lacZ Insertion Library Sac... 52 2e-22 emb|AI897134|AI897134 EST266577 tomato ovary, TAMU Lycopersicon ... 107 4e-22 emb|AI771644|AI771644 EST252744 tomato ovary, TAMU Lycopersicon ... 104 3e-21 55 emb|AV413397|AV413397 AV413397 Lotus japonicus young plants (two... 100 8e-20 emb|AW616540|AW616540 EST322951 L. hirsutum trichome, Cornell Un... 98 2e-19 emb|AL110506|SPBC577 S.pombe chromosome II cosmid c577. 74 le-18 emb|AW126841|AW126841 ga16f04.y1 Moss EST library PPU Physcomitr... 94 4e-18 emb|AI780067|AI780067 EST260946 tomato susceptible, Cornell Lyco... 94 5e-18 60 emb|AW687035|AW687035 NF005C05RT1F1037 Developing root Medicago ... 73 2e-17

emblAW685930|AW685930 NF036F03NR1F1000 Nodulated root Medicago t... 73 2e-17 . emb|AV415287|AV415287 AV415287 Lotus japonicus young plants (two... 90 4e-17 gb|BE124605|BE124605 EST393640 GVN Medicago truncatula cDNA clon... 65 5e-15 emb|AW220291|AW220291 EST302774 tomato root during/after fruit s... 68 1e-14 5 emb|AQ644913|AQ644913 RPCI93-EcoRI-2C7.TV RPCI93-EcoRI Trypanoso... 53 1e-13 emb|AA495505|AA495505 c430 Zhou and Ragan 1993 Gracilaria gracil... 49 4e-13 emb|AW830613|AW830613 sm04c07.y1 Gm-c1027 Glycine max cDNA clone... 77 4e-13 emb|AW666237|AW666237 sk34b04.y1 Gm-c1028 Glycine max cDNA clone... 77 4e-13 emb|AW285758|AW285758 LG1\_223\_C03.b1\_A002 Light Grown 1 (LG1) So... 74 4e-12 10 emb|AQ658256|AQ658256 Sheared DNA-13C5.TR Sheared DNA Trypanosom... 73 1e-11 emb|AW704333|AW704333 sk18a03.y1 Gm-c1028 Glycine max cDNA clone... 71 4e-11 emb|AV390446|AV390446 AV390446 Chlamydomonas reinhardtii C9 Chla... 70 5e-11 emb|AL160371|LMFLCHR15 Leishmania major Friedlin assembled chrom... 64 7e-11 emb|AZ048449|AZ048449 PSB67 Barley PstI genomic clones Hordeum v... 69 1e-10 emb|AW695904|AW695904 NF099H04ST1F1043 Developing stem Medicago ... 49 2e-10 15 emb|AW760518|AW760518 sl51d02.yl Gm-c1027 Glycine max cDNA clone... 66 9e-10 emb|AW332143|AW332143 S4G7 AGS-1 Pneumocystis carinii f. sp. car... 36 1e-09 emb|AW687790|AW687790 NF013E04RT1F1034 Developing root Medicago ... 65 2e-09 emb|AI486692|AI486692 EST245014 tomato ovary, TAMU Lycopersicon ... 54 3e-09 20 emb|AW333870|AW333870 S27C11 AGS-1 Pneumocystis carinii f. sp. c... 36 4e-09 emb|AQ850639|AQ850639 LMAJFV1\_lm41a04.x1 Leishmania major FV1 ra... 44 5e-09 emb|AW127178|AW127178 M110117 GVN Medicago truncatula cDNA clone... 63 8e-09 emb|AW399291|AW399291 EST309791 L. pennellii trichome, Cornell U... 62 2e-08 emb|AW616547|AW616547 EST322958 L. hirsutum trichome, Cornell Un... 62 2e-08 25 emb|AW693686|AW693686 NF068A05ST1F1036 Developing stem Medicago ... 62 2e-08 emb|AW208046|AW208046 M111077e DSIR Medicago truncatula cDNA clo... 61 4e-08 emb|AQ642482|AQ642482 RPCI93-EcoRI-1A17.TV RPCI93-EcoRI Trypanos... 53 6e-08 emb|AW686583|AW686583 NF039G02NR1F1000 Nodulated root Medicago t... 45 5e-07 emb|AQ849089|AQ849089 LMAJFV1 lm41a04.yl Leishmania major FV1 ra... 44 5e-06 30 emb|AI781410|AI781410 EST262277 tomato susceptible, Cornell Lyco... 52 2e-05 emb|AW617564|AW617564 EST323975 L. hirsutum trichome, Cornell Un... 49 2e-04 emb|AQ946427|AQ946427 Sheared DNA-49M8.TF Sheared DNA Trypanosom... 47 4e-04 emb|AW099113|AW099113 sd34f04.yl Gm-c1012 Glycine max cDNA clone... 46 0.001 emb|Z74807|SCYOL065C S.cerevisiae chromosome XV reading frame OR... 29 0.002 35 emblAQ655271|AQ655271 Sheared DNA-27A17.TR Sheared DNA Trypanoso... 38 0.004 emb|AW685854|AW685854 NF031A06NR1F1000 Nodulated root Medicago t... 43 0.007 emblAW437996|AW437996 ST83C09 Pine TriplEx shoot tip library Pin... 42 0.014 emb|AI440709|AI440709 sa62e11.yl Gm-c1004 Glycine max cDNA clone... 31 0.021 emb|AW979881|AW979881 EST341528 tomato root deficiency, Cornell ... 37 0.45 gb|M28064|PFAHRKP Plasmodium brasilianum DNA homologous to the h... 37 0.45 40 emb|AF263282|AF263282 Filobasidiella neoformans var. neoformans ... 34 .0.54 emb|AW702543|AW702543 TgESTzz85a12.y1 TgRH\*-Tachyzoite cDNA Toxo... 32 0.58 emb|AQ651205|AQ651205 Sheared DNA-30N15.TF Sheared DNA Trypanoso... 37 0.62 emb|AQ945454|AQ945454 Sheared DNA-54H22.TF Sheared DNA Trypanoso... 37 0.62 45 emb|AZ214873|AZ214873 Sheared DNA-109E2.TR Sheared DNA Trypanoso... 37 0.62 emb|AQ939979|AQ939979 Sheared DNA-42A15.TF Sheared DNA Trypanoso... 37 0.62 emb|AL049180|PFMAL13P1 Plasmodium falciparum chromosome 13 strai... 36 1.2 emb|Z74892|BO15H11 B.oleracea mRNA for glycine-rich protein. 36 1.2 emb[AW774740]AW774740 EST333891 KV3 Medicago truncatula cDNA clo... 36 1.2 50 emb|Z38060|SC5610 S.cerevisiae chromosome IX sequence derived fr... 36 1.2 emb|AL031745|PFMAL1P2 Plasmodium falciparum chromosome 1 strain ... 35 2.2

Query= AJ010971\_at 18022\_at /id\_source genbank /description
55 emb|cab52675.1| (aj010971) glucose-6-phosphate 1-dehydrogenase
[arabidopsis thaliana] /blast\_score 0
(1690 letters)

Database: plantfungal
60 661,018 sequences; 426,114,510 total letters

Searching......done Score Ε Sequences producing significant alignments: (bits) Value 5 emb[X74421]STG6PDH S.tuberosum mRNA for glucose-6-phosphate dehy... 911 0.0 emb|AF012862|AF012862 Petroselinum crispum cytosolic glucose-6-p... 841 0.0 emb|AJ001770|NTTCG9 Nicotiana tabacum mRNA for cytosolic glucose... 893 0.0 gb|U18238|MSU18238 Medicago sativa glucose-6-phosphate dehydroge... 900 0.0 10 emb|AF012863|AF012863 Petroselinum crispum cytosolic glucose-6-p... 887 0.0 emb|AJ001769|NTTCG6 Nicotiana tabacum mRNA cytosolic glucose-6-p... 895 0.0 emb|AF097663|AF097663 Mesembryanthemum crystallinum cytoplasmic ... 754 0.0 emb|AB011441|AB011441 Triticum aestivum WESR5 mRNA for glucose-6... 500 e-141 emb|X70373|KLLETZWF K.lactis LET1 gene and ZWF gene for glucose-... 231 e-138 15 emb|Z69381|SCCXIV39K S.cerevisiae 38,855 bp segment of chromosom... 244 e-136 gb|M34709|YSCG6PD S.cerevisiae glucose-6-phosphate dehydrogenase... 244 e-136 emb|X57336|SCMET19 S. cerevisiae MET19 gene for glucose-6-phosph... 244 e-136 emb|Z71517|SCYNL241C S.cerevisiae chromosome XIV reading frame O... 244 e-136 emb|AJ010712|STU010712 Solanum tuberosum mRNA for glucose-6-phos... 240 e-136 20 emb|X99405|NTG6PD N.tabacum mRNA for chloroplast glucose-6-phosp... 242 e-135 emb|AF012861|AF012861 Petroselinum crispum plastidic glucose-6-p... 248 e-135 emb|AI730607|AI730607 BNLGHi7371 Six-day Cotton fiber Gossypium ... 481 e-135 emb|X87942|ANG6PDHSE A.niger mRNA for glucose-6-phosphate dehydr... 225 e-132 emb|AW686120|AW686120 NF038D09NR1F1000 Nodulated root Medicago t... 472 e-132 25 emb|AJ132346|DBI132346 Dunaliella bioculata mRNA for plastidic g... 235 e-132 emb|AW925642|AW925642 HVSMEg0005C04 Hordeum vulgare pre-anthesis... 465 e-130 emb|AJ001772|NTTPG18 Nicotiana tabacum mRNA for plastidic glucos... 243 e-129 emb|AW930385|AW930385 EST340938 tomato fruit mature green, TAMU ... 444 e-124 emb|AW831416|AW831416 sm22g09.yl Gm-c1028 Glycine max cDNA clone... 434 e-121 30 emb|X83923|STG6PDHPI S.tuberosum mRNA for glucose-6-phosphate de... 249 e-118 emb|AJ000182|SO000182 Spinacia oleracea mRNA for glucose-6-phosp... 242 e-117 emb|AJ001771|NTTPG16 Nicotiana tabacum mRNA for plastidic glucos... 247 e-116 emb|AI491202|AI491202 EST241911 tomato shoot, Cornell Lycopersic... 368 e-116 emb|AL121764|SPAC9 S.pombe chromosome I cosmid c9. 196 e-116 35 emb|AW233801|AW233801 sf26h03.y1 Gm-c1028 Glycine max cDNA clone... 415 e-115 emb|Z95395|SPAC3A12 S.pombe chromosome I cosmid c3A12. 196 e-108 emb|AW288048|AW288048 N100892e rootphos(-) Medicago truncatula c... 387 e-106 emb|AW685333|AW685333 NF027C04NR1F1000 Nodulated root Medicago t... 356 e-105 emb[X77830]ANWGGSDA A.nidulans (WG096) gsdA gene. 130 e-102 emb|AJ006246|CCA6246 Cyanidium caldarium mRNA for glucose-6-phos... 239 e-102 emb|AJ000184|SO000184|Spinacia oleracea mRNA for glucose-6-phosp... 242 e-100 emb|AJ000183|SO000183 Spinacia oleracea mRNA for glucose-6-phosp... 190 3e-99 emb[X84001]ANDNAG6PD A.nidulans g6pd gene. 130 2e-98 emb|AW744917|AW744917 LG1 384 F04.b1 A002 Light Grown 1 (LG1) So... 356 3e-97 45 emb|AW761534|AW761534 sl68h05.y1 Gm-c1027 Glycine max cDNA clone... 356 4e-97 emb|X74988|PFGLPH P.falciparum gene for glucose-6-phosphate dehy... 192 5e-89 emb|AW560329|AW560329 EST315377 DSIR Medicago truncatula cDNA cl... 323 2e-87 gb]M80655[PFAG6PD Plasmodium falciparum glucose-6-phosphate dehy... 192 1e-86 emb|AW497059|AW497059 ga53c08.yl Moss EST library PPU Physcomitr... 319 5e-86 50 emb|AW216550|AW216550 EST295264 tomato callus, TAMU Lycopersicon... 188 6e-84 emb|AW219903|AW219903 EST302386 tomato root during/after fruit s... 305 7e-82 emb|AW180861|AW180861 MgA1030f MgA Library Mycosphaerella gramin... 226 5e-80 emb|AW616585|AW616585 EST322996 L. hirsutum trichome, Cornell Un... 175 7e-80 emb|AW309937|AW309937 sf26h03.x1 Gm-c1028 Glycine max cDNA clone... 294 1e-78 emb|AW031447|AW031447 EST274901 tomato callus, TAMU Lycopersicon... 166 6e-78 emb|AI894720|AI894720 EST264163 tomato callus, TAMU Lycopersicon... 291 1e-77 emb|AW690515|AW690515 NF030E09ST1F1000 Developing stem Medicago ... 174 3e-77 emb|AW736245|AW736245 EST332231 KV3 Medicago truncatula cDNA clo... 278 7e-76

emb|AW455246|AW455246 EST311906 tomato root during/after fruit s... 155 6e-75

emb|AL023595|SPCC794 S.pombe chromosome III cosmid c794.

emb|AW980010|AW980010 EST310488 tomato root deficiency, Cornell ... 153 3e-74

60

emb|AW255521|AW255521 ML551 peppermint glandular trichome Mentha... 270 3e-71 emb|AW567621|AW567621 si65f02.yl Gm-r1030 Glycine max cDNA clone... 264 1e-69 emb|AW689492|AW689492 NF019H04ST1F1000 Developing stem Medicago ... 128 3e-69 emb|AW737104|AW737104 EST338531 tomato flower buds, anthesis, Co... 141 1e-64 emb|AW255222|AW255222 ML216 peppermint glandular trichome Mentha... 142 2e-64 emb|AW737079|AW737079 EST338506 tomato flower buds, anthesis, Co... 141 3e-64 emb|AW565200|AW565200 LG1\_328\_D07.b1\_A002 Light Grown 1 (LG1) So... 119 1e-60 emb|AW684970|AW684970 NF023F11NR1F1000 Nodulated root Medicago t... 128 1e-60 emb|AQ952407|AQ952407 Sheared DNA-32N19.TF Sheared DNA Trypanoso... 84 2e-60 10 emb|AA787466|AA787466 n3f11a1.rl Aspergillus nidulans 24hr asexu... 229 4e-59 emb|AW254983|AW254983 ML1164 peppermint glandular trichome Menth... 134 2e-58 emb|AW617385|AW617385 EST323796 L. hirsutum trichome, Cornell Un... 141 2e-57 emb|AI773327|AI773327 EST254427 tomato resistant, Cornell Lycope... 222 5e-57 emb|AW704079|AW704079 sk27e10.y1 Gm-c1028 Glycine max cDNA clone... 219 6e-56 15 emb[X77829]ANNGSDA A.niger (N400) gsdA gene. 131 4e-55 emb|AW286177|AW286177 LG1\_328\_D07.g1\_A002 Light Grown 1 (LG1) So... 216 4e-55 emb|AW617091|AW617091 EST323502 L. hirsutum trichome, Cornell Un... 138 5e-55 emb|AI777244|AI777244 EST258209 tomato resistant, Cornell Lycope... 150 le-54 emb|AV424412|AV424412 AV424412 Lotus japonicus young plants (two... 205 9e-52 20 emb|AQ875204|AQ875204 V123H11 mTn-3xHA/lacZ Insertion Library, s... 192 le-49 emb|AV416113|AV416113 AV416113 Lotus japonicus young plants (two... 196 4e-49 emb|Z99568|SPAC3C7 S.pombe chromosome I cosmid c3C7. emb|AQ500574|AQ500574 V36G4 mTn-3xHA/lacZ Insertion Library Sacc... 127 6e-48 emb|AI187687|AI187687 p280 Porphyra yezoensis in Lambda UniZAP P... 87 1e-47 25 emb|AJ279688|BPE279688 Betula pendula partial mRNA for Glucose-6... 93 7e-47 emb|AW700124|AW700124 gb34g09.yl Moss EST library PPN Physcomitr... 187 3e-46 emb|AW223852|AW223852 EST300663 tomato fruit red ripe, TAMU Lyco... 141 6e-46 emb|AW774199|AW774199 EST333350 KV3 Medicago truncatula cDNA clo... 143 3e-45 emblAW621290|AW621290 EST312088 tomato root during/after fruit s... 138 3e-43 30 emb|AW980083|AW980083 EST341594 tomato root deficiency, Cornell ... 133 2e-42 emb[AZ217409]AZ217409 Sheared DNA-90C8.TR Sheared DNA Trypanosom... 90 3e-42 emb|AI941197|AI941197 sb85g11.yl Gm-c1010 Glycine max cDNA clone... 169 7e-41 emb|AW224136|AW224136 EST300947 tomato fruit red ripe, TAMU Lyco... 141 3e-40 emb|AW616535|AW616535 EST322946 L. hirsutum trichome, Cornell Un... 139 2e-38 35 emblAU036687|AU036687 Schizosaccharomyces pombe genomic clone ha... 98 3e-38 emb|AI416414|AI416414 NCW07G12T3 Westergaards Neurospora crassa ... 103 3e-38 emb|AW286271|AW286271 LG1\_329\_F06.g1\_A002 Light Grown 1 (LG1) So... 149\_4e-35 emblAI055328|AI055328 coau0003K24 Cotton Boll Abscission Zone cD... 78 3e-32 emb|AW698775|AW698775 r358 non-glandular-haired subtracted cDNA ... 133 5e-30 40 emblAJ130773 LDI130773 Laminaria digitata mRNA for glucose-6-pho... 131 2e-29 emb|AI974714|AI974714 T113172e KV2 Medicago truncatula cDNA clon... 118 2e-28 emb|AF173650|AF173650 Beta vulgaris clone GPD109UNI glucose-6-ph... 77 1e-27 emb|AJ004900|GMJ004900 Glycine max mRNA for glucose-6-phosphate-... 121 1e-26 emb|AW831248|AW831248 sm13f09.yl Gm-c1027 Glycine max cDNA clone... 104 8e-26 45 Query= AJ238846 at 18054 at /id source genbank /desccription emb|cab54517.1| (aj238846) sgp1 monomeric g-protein [arabidopsis thaliana]/blast score 1.00e-150 50 (887 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 55 Searching......done Score Ε Sequences producing significant alignments: (bits) Value 60 emb|AW725454|AW725454 GA\_\_Ea0018A14 Gossypium arboreum 7-10 dpa ... 238 8e-81 emb|Z38114|SC9745 S.cerevisiae chromosome XIII cosmid 9745.

122 2e-33

	gb[U34860]SCU34860 Saccharomyces cerevisiae origin recognition c 122 2e-33
	dbj D38172 YSCTEM1P Yeast gene for GTP-binding protein Tem1p, co 122 2e-33
	emb AI988573 AI988573 sd04c11.yl Gm-c1020 Glycine max cDNA clone 142 3e-33
	emb Y12314 SPSPG1GEN S.pombe spg1 gene. 67 9e-20
5	A LANGE A GOVERNMENT OF THE STATE OF THE STA
,	emb AJ001587 SPAJ1587 Schizosaccharomyces pombe sid3 gene. 67 9e-20
*	emb AW776339 AW776339 EST335404 DSIL Medicago truncatula cDNA cl 67 5e-14
•	emb AI416417 AI416417 NCW07E2T3 Westergaards Neurospora crassa c 55 1e-06
	gb U17243 YSCL8003 Saccharomyces cerevisiae chromosome XII cosmi 32 2e-06
	emb Z75093 SCYOR185C S.cerevisiae chromosome XV reading frame OR 32 2e-06
10	gb L08690 YSCGSP1X Yeast GTP-binding protein (GSP1) gene, comple 32 2e-06
	gb L08691 YSCGSP2X Yeast GTP-binding protein (GSP2) gene, comple 32 2e-06
	emb[X71946]SCCNR2A S.cerevisiae CNR2 gene. 32 2e-06
	emb X71945 SCCNR1A S.cerevisiae CNR1 gene. 32 2e-06
15	dbj D17748 TETTRAN Tetrahymena thermophila mRNA for Ran/TC4, com 33 3e-06
13	dbj D21825 TETPRAN Tetrahymena pyriformis mRNA for Ran/TC4, comp 33 3e-06
	gb[U17086]TBU17086 Trypanosoma brucei rhodesiense RAN/TC4 GTPase 30 6e-05
	emb AC008031 AC008031 Trypanosoma brucei chromosome II clone RPC 35 0.001
	emb AQ661102 AQ661102 Sheared DNA-15E18.TF Sheared DNA Trypanoso 32 0.002
	emb AW671770 AW671770 LG1_351_B08.b1_A002 Light Grown 1 (LG1) So 42 0.00
20	emb W68882 W68882 T2862 MVAT4 bloodstream form of serodeme WRATa 30 0.00
	emb AW685566 AW685566 NF031H02NR1F1000 Nodulated root Medicago t 36 0.03
	emb AB024996 AB024996 Cicer arietinum mRNA for rac-type small GT 36 0.091
	emb AW832298 AW832298 sm07b06.yl Gm-c1027 Glycine max cDNA clone 38 0.13
25	
23	emb AI437767 AI437767 sa39c11.yl Gm-c1004 Glycine max cDNA clone 37 0.17
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	emb AW690945 AW690945 NF034H11ST1F1000 Developing stem Medicago 35 0.23
	emb AW621657 AW621657 EST312455 tomato root during/after fruit s 37 0.24
	emb AI775563 AI775563 EST256663 tomato resistant, Cornell Lycope 37 0.24
30	emb AW738459 AW738459 EST339886 tomato flower buds, anthesis, Co 36 0.33
	emb AI731040 AI731040 BNLGHi8453 Six-day Cotton fiber Gossypium 36 0.33
	emb AI727570 AI727570 BNLGHi8420 Six-day Cotton fiber Gossypium 36 0.33
	emb AV419645 AV419645 AV419645 Lotus japonicus young plants (two 36 0.33
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35	emb AW930158 AW930158 EST340615 tomato fruit mature green, TAMU 36 0.33
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	emb AI440994 AI440994 sa63e10.yl Gm-c1004 Glycine max cDNA clone 36 0.45
	gb U03624 PTU03624 Paramecium tetraurelia clone plg-14 guanine n 36 0.45
40	gb U64924 NTU64924 Nicotiana tabacum geranylgeranylated protein 35 0.55
40	emb AW040005 AW040005 EST282496 tomato mixed elicitor, BTI Lycop 35 0.56
	emb AW039993 AW039993 EST282484 tomato mixed elicitor, BTI Lycop 35 0.56
	emb AW705028 AW705028 sk41f03.yl Gm-c1019 Glycine max cDNA clone 35 0.57
	emb AW929161 AW929161 EST337949 tomato flower buds 8 mm to pre-a 35 0.57
	emb AW705209 AW705209 sk43a11.yl Gm-c1019 Glycine max cDNA clone 35 0.57
45	emb AV414769 AV414769 AV414769 Lotus japonicus young plants (two 35 0.62
	gb U64923 NTU64923 Nicotiana tabacum geranylgeranylated protein 35 0.62
	emb Z73962 LJRAC2 L.japonicus mRNA for small GTP-binding protein 35 0.62
	gb BE021064 BE021064 sm47d01.yl Gm-c1028 Glycine max cDNA clone 35 0.62
	emb AW573660 AW573660 EST316251 GVN Medicago truncatula cDNA clo 35 0.62
50	emb AV413108 AV413108 AV413108 Lotus japonicus young plants (two 35 0.62
	emb AW202293 AW202293 sf13c10.y1 Gm-c1027 Glycine max cDNA clone 35 0.62
	emb AI937960 AI937960 sc06b11.yl Gm-c1012 Glycine max cDNA clone 35 0.62
	omb/773061/t TD A CL Librarious TDNA for small CTD Librarious 25 0.02
	emb Z73961 LJRAC1 L.japonicus mRNA for small GTP-binding protein 35 0.62
c	emb AI162198 AI162198 A013P52U Hybrid aspen plasmid library Popu 35 0.62
55	emb AW694335 AW694335 NF075C06ST1F1049 Developing stem Medicago 35 0.62
	emb AW109094 AW109094 gate0002P07f Gossypium arboreum 7-10 dpa f 35 0.62
	gb L19093 PEARHOGTPP Pisum sativum rho (ras-related) GTP-binding 35 0.62
	emb AW565277 AW565277 LG1_332_G03.b1_A002 Light Grown 1 (LG1) So 35 0.62
	emb AV412205 AV412205 AV412205 Lotus japonicus young plants (two 35 0.62
60	gb BE054534 BE054534 GA_Ea0033M19f Gossypium arboreum 7-10 dpa 35 0.62
	emb AW108667 AW108667 gate0001G03f Gossypium arboreum 7-10 dpa f 35 0.62

	emb AI495724 AI495724 sb15e06.yl Gm-c1004 Glycine max cDNA clone 35 0.62 gb S79309 S79309 Rac9=21.5 kda GTP-binding protein [Gossypium hi 35 0.62 gb S79308 S79308 Rac13=21.8 kda GTP-binding protein [Gossypium h 35 0.62
5	emb AJ250174 NTA250174 Nicotiana tabacum mRNA for putative rac p 35 0.62 gb BE055015 BE055015 GA_Ea0001G03f Gossypium arboreum 7-10 dpa 35 0.62
	emb AW690086 AW690086 NF028B10ST1F1000 Developing stem Medicago 35 0.62 emb AW573665 AW573665 EST316256 GVN Medicago truncatula cDNA cl 35 0.62 emb AW559248 AW559248 EST306084 DSIR Medicago truncatula cDNA cl 35 0.62
	emb AI460950 AI460950 sa78f02.y1 Gm-c1004 Glycine max cDNA clone 35 0.62
10	emb AW692052 AW692052 NF047B03ST1F1000 Developing stem Medicago 35 0.62
	emb AW394676 AW394676 sh34a02.y1 Gm-c1017 Glycine max cDNA clone 35 0.62
	gb BE054555 BE054555 GA_Ea0002P07f Gossypium arboreum 7-10 dpa 35 0.62
	emb AF146340 AF146340 Physcomitrella patens Rac-like GTP binding 35 0.85
	emb AF233446 AF233446 Physcomitrella patens rac 1 protein (rac1) 35 0.85
15	emb AF115476 AF115476 Physcomitrella patens rac-like GTP binding 35 0.85
•	gb U06051 PFU06051 Plasmodium falciparum Dd2 Ran/TC4 nuclear GTP 35 0.85
	emb AW729788 AW729788 GA_Ea0026A18 Gossypium arboreum 7-10 dpa 35 0.8:
	emb AF146341 AF146341 Physcomitrella patens Rac-like GTP binding 35 0.85
	emb AI731831 AI731831 BNLGHi11032 Six-day Cotton fiber Gossypium 35 0.85
<b>20</b> .	emb AF051223 AF051223 Picea mariana Rac-like GTP binding protein 35 0.85
	gb BE055583 BE055583 GA_Ea0001I02f Gossypium arboreum 7-10 dpa 35 0.85
	emb AW108575 AW108575 gate0001102f Gossypium arboreum 7-10 dpa f 35 0.85
	emb X73954 PFRAN1 P.falciparum gene for ras-related nuclear prot 35 0.85
25	emb AW056772 AW056772 ST55F07 Pine TriplEx shoot tip library Pin 35 1.2
23	emb AW559842 AW559842 EST314890 DSIR Medicago truncatula cDNA cl 34 1.6
	emb AW217581 AW217581 EST296295 tomato flower buds 3-8 mm, Corne 34 1.6
	emb AW217573 AW217573 EST296287 tomato flower buds 3-8 mm, Corne 34 1.6 emb Z36133 SCYBR264C S.cerevisiae chromosome II reading frame OR 34 1.6
	emb Z36133 SCYBR264C S.cerevisiae chromosome II reading frame OR 34 1.6 emb X70529 SCCIIORF S. cerevisiae chromosome II sequence for ORF 34 1.6
30	emb AI778114 AI778114 EST258993 tomato susceptible, Cornell Lyco 34 1.6
	emb AF042330 AF042330 Brassica rapa rac-like small GTP binding p 34 1.6
	emb AW625569 AW625569 EST319476 tomato radicle, 5 d post-imbibit 34 1.6
	emb AW931490 AW931490 EST357333 tomato fruit mature green, TAMU 34 1.6
	gb BE033825 BE033825 MG04D04 MG Mesembryanthemum crystallinum cD 34 2.2
35	gb BE036577 BE036577 MP01F02 MP Mesembryanthemum crystallinum cD 34 2.2
	emb AQ399302 AQ399302 mgxb0015O18f CUGI Rice Blast BAC Library P 34 2.2
	emb AW697756 AW697756 Str1-D6 Sugar Beet germination cDNA librar 34 2.2
40	Query= X91916_at 18716_at /id_source genbank /description
-	gb aaf26754.1 ac007396_3 (ac007396) t4012.6 [arabidopsis thaliana]
	/blast_score 3.00e-36
	(403 letters)
45	Database: plantfungal
	661,018 sequences; 426,114,510 total letters
	Searchingdone
50	Score E
	Sequences producing significant alignments: (bits) Value
	gb U18557 RSU18557 Raphanus sativus antifungal protein 1 preprot 166 7e-43
	emb A26875 A26875 R.sativus AFP1 gene. 166 7e-43
55	emb X97318 RSAFP4 R.sativus mRNA for antifungal protein 4. 162 6e-42
	gb U18556 RSU18556 Raphanus sativus antifungal protein 2 preprot 160 1e-38
	emb X97319 RSEFP3 R.sativus mRNA for antifungal protein 3. 133 8e-37
	gb U59459 BNU59459 Brassica napus antifungal protein mRNA, compl 135 2e-36
60	emb AT001728 AT001728 AT001728 Flower bud cDNA Brassica rapa sub 82 2e-15
UU	gb L47901 L47901 BNAF1712 Mustard flower buds Brassica rapa cDNA 81 5e-15
	emb A26963 A26963 D.merkii AMP1 sequence. 74 5e-13

	emb A27062 A27062 C.benedictus AMP2 sequence. 73 1e-12
	emb X53375 HASF18 Sunflower anther-specific mRNA SF18. 72 3e-12
_	emb AT001738 AT001738 AT001738 Flower bud cDNA Brassica rapa sub 69 2e-11
5	emb[A27064]A27064 C.ternatea AMP1 sequence. 58 8e-10
	emb X91487 PASPIIGEN P.abies mRNA for gamma-thionin protein (put 40 1e-05
	emb AW870017 AW870017 NXNV_122_A10_F Nsf Xylem Normal wood Verti 35 6e-05
	and I Wild Are I I A Wild Are I Cross Do A P
	emb AW064751 AW064751 ST35D04 Pine TriplEx shoot tip library Pin 36 7e-05
	emb AW775186 AW775186 EST331908 GVN Medicago truncatula cDNA clo 41 0.001
10	dbj D29679 TOBPIT1A Tobacco pit1 mRNA (which expression is induc 41 0.003
	emb AW621708 AW621708 EST312506 tomato root during/after fruit s 36 0.007
	4 1 4 7774 4 10 4 7 1 4 7 1 7 1 7 1 7 1 7 1 7 1 7 1 7 1
	emb AW220215 AW220215 EST302698 tomato root during/after fruit s 36 0.007
	emb AW623112 AW623112 EST321057 tomato flower buds 3-8 mm, Corne 40 0.008
	emb AW220231 AW220231 EST302714 tomato root during/after fruit s 36 0.008
15	emb AW219164 AW219164 EST301646 tomato root during/after fruit s 36 0.008
	emb AW219793 AW219793 EST302275 tomato root during/after fruit s 36 0.009
	111-10-0-11-1-10-0-1
	emb AI487824 AI487824 EST246146 tomato ovary, TAMU Lycopersicon 40 0.009
	emb AI483999 AI483999 EST249870 tomato ovary, TAMU Lycopersicon 40 0.010
	gb[U20591 SLU20591 Solanum lycopersicum flower-specific gamma-th 40 0.010
20	emb AW622923 AW622923 EST306993 tomato flower buds 3-8 mm, Corne 40 0.010
	emb AW929929 AW929929 EST354199 tomato flower buds 8 mm to pre-a 40 0.010
	111777000000117770000000000000000000000
	emb AW929939 AW929939 EST354209 tomato flower buds 8 mm to pre-a 40 0.010
	emb AW217552 AW217552 EST296266 tomato flower buds 3-8 mm, Corne 40 0.012
	emb AI490062 AI490062 EST248401 tomato ovary, TAMU Lycopersicon 40 0.012
25	emb AW217379 AW217379 EST296063 tomato flower buds 0-3 mm, Corne 40 0.012
	emb AI484887 AI484887 EST243150 tomato ovary, TAMU Lycopersicon 40 0.012
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	emb AI897392 AI897392 EST266835 tomato ovary, TAMU Lycopersicon 40 0.012
	emb AW219347 AW219347 EST301829 tomato root during/after fruit s 33 0.014
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	emb AW622375 AW622375 EST313174 tomato root during/after fruit s 35 0.026
40	
40	emb AW622390 AW622390 EST313189 tomato root during/after fruit s 35 0.029
	emb AI486656 AI486656 EST244977 tomato ovary, TAMU Lycopersicon 38 0.031
	emb AW399651 AW399651 EST310151 L. pennellii trichome, Cornell U 38 0.043
	emb AW621171 AW621171 EST311969 tomato root during/after fruit s 34 0.048
	emb AW622051 AW622051 EST312849 tomato root during/after fruit s 34 0.053
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	emb AW621962 AW621962 EST312760 tomato root during/after fruit s 33 0.070
50	emb AF153353 AF153353 Dimocarpus longan ribulose 1,5-bisphosphat 36 0.15
-	
	emb AW621634 AW621634 EST312432 tomato root during/after fruit s 31 0.32
	emb AW255597 AW255597 ML641 peppermint glandular trichome Mentha 35 0.40
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55	emb Y15139 BOY15139 Bixa orellana chloroplast rbcL gene. 34 0.54
	· i · · · · · · · · · · · · · · · · ·
	emb AF022128 AF022128 Bixa orellana ribulose 1,5-bisphosphate ca 34 0.54
	emb AF022125 AF022125 Theobroma cacao ribulose 1,5-bisphosphate 34 0.54
	gb L12568 AKARBC Akania bidwillii ribulosebisphosphate carboxyla 34 0.54
60	emb[A27063]A27063 L.cicera AFP sequence. 34 0.54
	gb M95753 BCNCPRBCL Bretschneidera sinensis chloroplast ribulose 34 0.54
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	emb AF022120 AF022120 Camptostemon schultzii ribulose 1,5-bispho 33 1.4
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35	lipase/acylhydrolase with gdsl-motif family. ests gbht44453,
-	gb t04815, gb t45993, gb r30138, gb ai099570 and gb t22281 come from
	this gene. [arabidopsis thaliana]" /blast score 0 /ec number /family
	/chip nova /gb link
	http://www3.ncbi.nlm.nih.gov/htbin-
40	post/entrez/query?db=n&form=6&dopt=g&uid=gb ac006577  /ncgi
	http://www.ncgr.org/cgi-bin/ff?ac006577
	(1194 letters)
	Database: plantfungal
45	661,018 sequences; 426,114,510 total letters
	Constino
	Searchingdone
	Score E
50	Sequences producing significant alignments: (bits) Value
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	emb Y10155 BNMYAP9 B.napus for myrosinase-associated protein, cl 224 e-160
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60	emb AW774945 AW774945 EST334096 KV3 Medicago truncatula cDNA clo 46 5e-13
JU	emb AW685185 AW685185 NF025E06NR1F1000 Nodulated root Medicago t 62 1e-10 emb AW922141 AW922141 LG1 323 G11 b1 A002 Light Grown 1 (LG1) So. 65 5e-16
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emblAF022157|AF022157 Glycine max cytochrome P450 monooxygenase ... . 158 7e-53 emb|AW774664|AW774664 EST333815 KV3 Medicago truncatula cDNA clo... 208 9e-53 emb|AF029857|AF029857 Sorghum bicolor cytochrome P450 CYP99A1 (C... 98 1e-52 emb|AB001380|AB001380 Glycyrrhiza echinata CYP93B1 mRNA for cyto... 109 1e-52 5 emb|AW616345|AW616345 EST322756 L. hirsutum trichome, Cornell Un... 208 1e-52 emb|AW690420|AW690420 NF033B04ST1F1000 Developing stem Medicago ... 129 1e-52 emblAF000403|AF000403 Lotus japonicus putative cytochorome P450 ... 145 2e-52 emblAW922446|AW922446 DG1\_19\_F10.g1\_A002 Dark Grown 1 (DG1) Sorg... 204 2e-51 emb|AI729430|AI729430 BNLGHi13332 Six-day Cotton fiber Gossypium... 171 2e-51 10 emb|AJ010324|PAJ10324 Populus trichocarpa cv trichobel mRNA for ... 93 4e-51 emb|AF156976|AF156976 Gerbera hybrida flavone synthase II (CYP93... 110 4e-51 emb|AB025016|AB025016 Lotus japonicus mRNA for cytochrome P450, ... 152 6e-51 emb|AI779369|AI779369 EST260248 tomato susceptible, Cornell Lyco... 202 8e-51 emb|AI727414|AI727414 BNLGHi7936 Six-day Cotton fiber Gossypium ... 181 1e-50 15 Query= AC000375.44\_at 13115\_at /id\_source genbank /description "gb|aab60774.1| (ac000375) ests gb|u75592,gb|t13956,gb|t43869 come from from this gene. [arabidopsis thaliana]" /blast score 0 /ec number 20 /family /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|ac000375| /ncgi http://www.ncgr.org/cgi-bin/ff?ac000375 (1533 letters) 25 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching.....done 30 Ε Score Sequences producing significant alignments: (bits) Value gb|BE020282|BE020282 sm42g12.y1 Gm-c1028 Glycine max cDNA clone ... 91 2e-39 35 emb|AW831934|AW831934 sm18a01.yl Gm-c1027 Glycine max cDNA clone... 91 3e-38 emb|AI895084|AI895084 EST264527 tomato callus, TAMU Lycopersicon... 88 2e-37 emblAW731252lAW731252 GA Ea0030E03 Gossypium arboreum 7-10 dpa ... 93 2e-31 emb|AW650703|AW650703 EST329157 tomato germinating seedlings, TA... 88 2e-28 emb|AW648696|AW648696 EST327066 tomato germinating seedlings, TA... 85 7e-25 40 emb|AT000481|AT000481 AT000481 Brassica rapa guard cell Brassica... 62 3e-23 emb|AI895460|AI895460 EST264903 tomato callus, TAMU Lycopersicon... 77 2e-22 emb|AW776692|AW776692 EST335757 DSIL Medicago truncatula cDNA cl... 68 7e-21 emb|AI774644|AI774644 EST255744 tomato resistant, Cornell Lycope... 65 4e-19 emblAW560992|AW560992 EST316040 DSIR Medicago truncatula cDNA cl... 68 2e-18 45 emb|AW034968|AW034968 EST279197 tomato callus, TAMU Lycopersicon... 61 2e-15 emb|AW034372|AW034372 EST277943 tomato callus, TAMU Lycopersicon... 62 1e-13 gb|BE021411|BE021411 sm48g02.y1 Gm-c1028 Glycine max cDNA clone ... 60 6e-13 emb|AW032312|AW032312 EST275766 tomato callus, TAMU Lycopersicon... 69 9e-12 emb|AW399297|AW399297 EST309797 L. pennellii trichome, Cornell U... 46 2e-11 50 emb|AW032231|AW032231 EST275685 tomato callus, TAMU Lycopersicon... 62 6e-11 emb|AW032133|AW032133 EST275587 tomato callus, TAMU Lycopersicon... 62 6e-11 emb|AW030333|AW030333 EST273588 tomato callus, TAMU Lycopersicon... 55 8e-11 gb|BE020267|BE020267 sm42f07.yl Gm-c1028 Glycine max cDNA clone ... 69 1e-10 emb|AW773655|AW773655 EST332641 KV3 Medicago truncatula cDNA clo... 55 1e-10 55 emb|AI896893|AI896893 EST266336 tomato callus, TAMU Lycopersicon... 62 1e-10 emb|AW278353|AW278353 sf43a06.yl Gm-c1009 Glycine max cDNA clone... 49 2e-09 emb|AW687210|AW687210 NF007B09RT1F1076 Developing root Medicago ... 55 1e-08 emb|AT000955|AT000955 AT000955 Brassica rapa guard cell Brassica... 60 3e-08 emb|AW029692|AW029692 EST272947 tomato callus, TAMU Lycopersicon... 48 4e-08 60 emb|AT000818|AT000818 AT000818 Brassica rapa guard cell Brassica... 59 7e-08 emb|AW774597|AW774597 EST333748 KV3 Medicago truncatula cDNA clo... 47 2e-07

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emb|AI776153|AI776153 EST257241 tomato resistant, Cornell Lycope... 177 5e-56 emb|AI898279|AI898279 EST267722 tomato ovary, TAMU Lycopersicon ... 141 2e-55 gb|BE033502|BE033502 ME03F10 ME Mesembryanthemum crystallinum cD... 86 3e-55 emblAW030814|AW030814 EST274069 tomato callus, TAMU Lycopersicon... 140 6e-55 emb|AI485982|AI485982 EST244303 tomato ovary, TAMU Lycopersicon ... 138 3e-54 emblAW035013|AW035013 EST279284 tomato callus, TAMU Lycopersicon... 137 4e-54 emb|AW684552|AW684552 NF018C03NR1F1000 Nodulated root Medicago t... 98 1e-53 emb|AW029733|AW029733 EST272988 tomato callus, TAMU Lycopersicon... 134 2e-53 emb|AW924422|AW924422 WS1\_69\_C06.b1\_A002 Water-stressed 1 (WS1) ... 139 4e-53 10 emb|Z46950|SNCHJET19 S.nigra mRNA chitinase class II, pathogenes... 130 7e-53 gb|BE034447|BE034447 MH05A08 MH Mesembryanthemum crystallinum cD... 86 8e-53 emb[Y10373[MTCHITIN1 M.truncatula mRNA for chitinase. 116 2e-52 gb|U83592|MSU83592 Medicago sativa class I chitinase mRNA, compl... 116 1e-51 gb|U83591|MSU83591 Medicago sativa class I chitinase mRNA, compl... 116 1e-51 15 gb|L37876|PEACHI2I Pisum sativum chitinase class I (chi2) gene, ... 117 9e-51 emb|AW648023|AW648023 EST326477 tomato germinating seedlings, TA... 124 2e-50 gb|BE034481|BE034481 MH05F02 MH Mesembryanthemum crystallinum cD... 141 6e-50 emb|AF090336|AF090336 Citrus sinensis chitinase CHI1 (chi1) mRNA... 149 1e-49 emblAI897657|AI897657 EST267100 tomato ovary, TAMU Lycopersicon ... 165 4e-49 20 emb|AW033122|AW033122 EST276681 tomato callus, TAMU Lycopersicon... 120 4e-49 emb[X15494|STCHITIN Potato endochitinase gene (EC 3,2,1,14). · 115 9e-49 emb|A16119|A16119 Intracellular chitinase mRNA (SEQ ID NO: 2). gb|M15173|TOBECH Tobacco (N.tabacum) endochitinase mRNA, partial... 112 1e-48 emb|X07130|STCHIT Solanum tuberosum mRNA for endochitinase (EC 3... 115 1e-48 25 emb|AW745819|AW745819 WS1\_37\_H11.g1\_A002 Water-stressed I (WS1) ... 140 2e-48 gb|BE034450|BE034450 MH05B01 MH Mesembryanthemum crystallinum cD... 83 3e-48 gb|S44869|S44869 basic chitinase [Nicotiana tabacum=tobacco, cv... 112 5e-48 emb|X16939|NTECHITR Nicotiana tabacum mRNA for endochitinase (EC... 112 5e-48 emb|AW922735|AW922735 DG1\_45\_B06.g1\_A002 Dark Grown 1 (DG1) Sorg... 139 9e-48 30 emb|AW746695|AW746695 WS1\_54\_E02.g1\_A002 Water-stressed 1 (WS1) ... 139\_9e-48 emb|AJ012821|CAR012821 Cicer arietinum mRNA for class I chitinase. 118 3e-47 emblAF061806|AF061806 Elaeagnus umbellata basic chitinase mRNA, ... 118 4e-47 gb|M94105|ALCCHITIN Allium sativum chitinase mRNA, 3' end. emb|AF000965|AF000965 Poa pratensis chitinase (Chi3) pseudogene ... 120 2e-46 35 gb|M13968|PHVCHM P.vulgaris chitinase mRNA, complete cds. emb|AW922596|AW922596 DG1 46\_C01.b1 A002 Dark Grown 1 (DG1) Sorg... 79 3e-46 emb|AF000966|AF000966 Poa pratensis chitinase (Chi2) gene, compl... 116 3e-46 gb|BE034267|BE034267 MH02D06 MH Mesembryanthemum crystallinum cD... 74 3e-46 gb|U78888|GHU78888 Gossypium hirsutum class I endochitinase mRNA... 122 4e-46 40 emb|X76041|TACHIG T.aestivum (Chinese spring) chi gene for endoc... 116 4e-46 emb|X88800|VURNACHI1 V.unguiculata mRNA for chitinase clase 1 (p... 112 7e-46 emb|AF034566|AF034566 Gossypium hirsutum class I chitinase mRNA,... 121 9e-46 emb|X14133|STENCHIT Potato mRNA fragment for endochitinase (EC 3... 109 1e-45 emb|Z15140|LECHI9 L.esculentum mRNA for chitinase. 112 2e-45 45 emb X74919 PVGEC9 P. vulgaris gene for endochitinase. 135 2e-45 gb|S43926|S43926 CH5B=chitinase [Phaseolus vulgaris=beans, cv Sa... 116 2e-45 gb|U02607|STU02607 Solanum tuberosum chitinase (chtB3) mRNA, par... 109 2e-45 gb|M94106|ALCCHINTIA Allium sativum chitinase mRNA, 3' end. gb|U02605|STU02605 Solanum tuberosum chitinase (chtB1) mRNA, par... 109 6e-45 50 gblU02606|STU02606 Solanum tuberosum chitinase (chtB2) mRNA, par... 108 8e-45 emb|AW922776|AW922776 DG1 46 C01.g1\_A002 Dark Grown 1 (DG1) Sorg... 147 2e-44 gb|BE033398|BE033398 ME01A01 ME Mesembryanthemum crystallinum cD... 141 4e-44 emblAA739579|AA739579 344 PtIFG2 Pinus taeda cDNA clone 8562M 3'... 107 1e-43 emb|AW560048|AW560048 EST315096 DSIR Medicago truncatula cDNA cl... 116 1e-43 emb|A23396|A23396 B.vulgaris gene for chitinase 76. 105 3e-43 emb|AW267781|AW267781 EST305909 DSIR Medicago truncatula cDNA cl... 115 4e-43 emb|Z78202|PACHI1 Persea americana mRNA for endochitinase. emb|AF098302|AF098302 Brassica juncea chitinase mRNA, complete cds. 125 1e-42 emb|AI352718|AI352718 MB46-29 PZ204.BNlib Brassica napus cDNA cl... 144 4e-42 60 gb|BE034616|BE034616 ML04B04 ML Mesembryanthemum crystallinum cD... 80 5e-42 emb|AI729668|AI729668 BNLGHi13889 Six-day Cotton fiber Gossypium... 111 2e-41

emb|AF061805|AF061805 Elaeagnus umbellata acidic chitinase mRNA,... 116 3e-41 gb|U02287|HVU02287 Hordeum vulgare cultivar NK1558 chitinase gen... 117 4e-41 emb|Z15138|LECHI14 L.esculentum mRNA for chitinase (partial). gb|U30465|LEU30465 Lycopersicon esculentum class II chitinase (C... 114 2e-40 emb|Z54234|VVCHIT1MR V.vinifera mRNA for chitinase. 100 le-39 Query= AL031394.56 at 13176 at /id source genbank /description emblcaa20567.1 (al031394) putative protein [arabidopsis thaliana] 10 /blast\_score 0 /ec number /family /chip nova /gb\_link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|al031394| /ncgi http://www.ncgr.org/cgi-bin/ff?al031394 (1116 letters) 15 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 20 Score Sequences producing significant alignments: (bits) Value emb|AW671006|AW671006 LG1\_278\_H12.b1\_A002 Light Grown 1 (LG1) So... 81 6e-28 25 emb|AA520813|AA520813 TgESTzz64d07.r1 TgME49 invivo Bradyzoite c... 38 0.16 gb|U43491|SCU43491 Saccharomyces cerevisiae cosmid clone pEOA156... 37 0.31 emb|Z74916|SCYOR008C S.cerevisiae chromosome XV reading frame OR... 37 0.31 gb|U39481|SCU39481 Saccharomyces cerevisiae Slglp (SLG1) gene, c... 37 0.31 emb|AA680906|AA680906 LmFrAm0494 Leishmania major Amastigote ful... 33 1.0 30 emb|AF193903|AF193903 Cafeteria roenbergensis mitochondrial DNA,... 35 1.1 emb|AF229795|AF229795 Vigna radiata beta galactosidase mRNA, com... 35 1.5 emb|AW683786|AW683786 NF001A06NR1F1038 Nodulated root Medicago t... 35 1.5 emb|AC005802|AC005802 Leishmania major chromosome 3 clone L6202 ... 34 2.1 emb|AC005893|AC005893 Leishmania major chromosome 3 clone L822 s... 34 2.1 emb|AA520166|AA520166 TgESTzz39b08.s1 TgME49 invivo Bradyzoite c... 34 2.9 35 emb|AW618793|AW618793 EST320779 L. pennellii trichome, Cornell U... 34 2.9 emb|AW306460|AW306460 se51a02.y1 Gm-c1017 Glycine max cDNA clone... 34 2.9 emblAQ502036|AQ502036 V10E12 mTn-3xHA/lacZ Insertion Library Sac... 33 3.9 gb[U87148]HVU87148 Hordeum vulgare nucellin mRNA, complete cds. 40 emb|AF017430|AF017430 Hordeum vulgare EEA1 mRNA, complete cds. emb|AQ500344|AQ500344 V41B12 mTn-3xHA/lacZ Insertion Library Sac... 33 3.9 emb|Z71686|SCYNR071C S.cerevisiae chromosome XIV reading frame O... 33 3.9 emb|AQ501720|AQ501720 V15F6 mTn-3xHA/lacZ Insertion Library Sacc... 33 3.9 emb|AQ872965|AQ872965 V56D9 mTn-3xHA/lacZ Insertion Library, str... 33 3.9 45 emb|AW126050|AW126050 N100246e rootphos(-) Medicago truncatula c... 33 5.4 emb|AW127669|AW127669 M110413 DSLC Medicago truncatula cDNA clon... 33 5.4 emb|AW126177|AW126177 N100022e rootphos(-) Medicago truncatula c... 33 5.4 emb|AW126137|AW126137 N100334e rootphos(-) Medicago truncatula c... 33 5.4 emb|AW509583|AW509583 ga61f07.y1 Moss EST library PPU Physcomitr... 33 5.4 50 emb|AQ503147|AQ503147 V44D10 mTn-3xHA/lacZ Insertion Library Sac... 33 5.4 emb|AQ501343|AQ501343 V23B5 mTn-3xHA/lacZ Insertion Library Sacc... 33 5.4 emb|Z98559|SPAC23C11 S.pombe chromosome I cosmid c23C11. emb|AW672219|AW672219 LG1\_358\_B09.b1\_A002 Light Grown 1 (LG1) So... 33 5.4 emb|AW925272|AW925272 HVSMEg0001G19 Hordeum vulgare pre-anthesis... 33 5.4 55 emb|AA783066|AA783066 a1h02c9.rl Aspergillus nidulans 24hr asexu... 26 5.7 emb|AC013353|AC013353 Trypanosoma brucei chromosome VI clone RPC... 32 7.4 emb|AV409938|AV409938 AV409938 Lotus japonicus young plants (two... 32 7.4 emb|AW780906|AW780906 sl86c03.yl Gm-c1037 Glycine max cDNA clone... 32 7.4 emb|AI443278|AI443278 sa46f11.yl Gm-c1004 Glycine max cDNA clone... 32 7.4 60 emb|AW156670|AW156670 se29c01.yl Gm-c1015 Glycine max cDNA clone... 32 7.4 emb|AW277786|AW277786 sf86e12.yl Gm-c1019 Glycine max cDNA clone... 32 7.4

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5	emb AW704821  AW704821 sk40a01.yl Gm-c1019 Glycine max cDNA clone 32 7.4 emb AW277436  AW277436 sf82a10.yl Gm-c1019 Glycine max cDNA clone 32 7.4 emb AW459299  AW459299 sh23c01.yl Gm-c1016 Glycine max cDNA clone 32 7.4 emb AW407743  AV407743 AV407743 Lotus japonicus young plants (two 32 7.4 emb AW757240  AW757240 sl30h11.yl Gm-c1027 Glycine max cDNA clone 32 7.4
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15	http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb al021961  /ncgi http://www.ncgr.org/cgi-bin/ff?al021961  (1812 letters)
20	Database: plantfungal 661,018 sequences; 426,114,510 total letters
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25	Score E Sequences producing significant alignments: (bits) Value
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      emblAJ001429|RGDMANDER Rhodotorula graminis mRNA for D-mandelate... 70 6e-11
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emb|AW040337|AW040337 EST283201 tomato mixed elicitor, BTI Lycop... emb|AW037837|AW037837 EST279466 tomato mixed elicitor, BTI Lycop... 70 6e-11 emb|AL033497|CAC49C10 C.albicans cosmid Ca49C10. emb|AI897727|AI897727 EST267170 tomato ovary, TAMU Lycopersicon ... 69 9e-11 5 emb|AI898454|AI898454 EST267897 tomato ovary, TAMU Lycopersicon ... 69 9e-11 Query= AL035394.196 at 13627 at /id source genbank /description emb|caa23036.1| (al035394) putative na+/h+-exchanging protein 10 [arabidopsis thaliana] /blast score 0 /ec number /family /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|a1035394| /ncgi http://www.ncgr.org/cgi-bin/ff?al035394 (2463 letters) 15 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 20 Score Sequences producing significant alignments: (bits) Value emb|AW736388|AW736388 EST332307 KV3 Medicago truncatula cDNA clo... 243 5e-63 25 emb|AW622441|AW622441 EST313229 tomato root during/after fruit s... 204 2e-51 emb|AL157994|SPAC105 S.pombe chromosome I cosmid c105. emb|AQ841817|AQ841817 T134068 Soybean RFLP probe Glycine max gen... 186 8e-46 emb|X77087|SJ100567 S.cerevisiae (S288C) J0909, J0911, J0914 and... 150 5e-35 emb|Z49369|SCYJL094C S.cerevisiae chromosome X reading frame ORF... 150 5e-35 30 emb|AW774205|AW774205 EST333356 KV3 Medicago truncatula cDNA clo... 120 8e-28 emb|AW559356|AW559356 EST314404 DSIR Medicago truncatula cDNA cl... 104 2e-23 emb|AQ361449|AQ361449 mgxb0004G06r CUGI Rice Blast BAC Library P... 59 4e-16 emb|AT000678|AT000678 AT000678 Brassica rapa guard cell Brassica... 79 2e-13 emb|AV409975|AV409975 AV409975 Lotus japonicus young plants (two... 59 5e-11 35 emb|AT000552|AT000552 AT000552 Brassica rapa guard cell Brassica... 57 6e-07 emb|AW980088|AW980088 EST341613 tomato root deficiency, Cornell ... 45 8e-07 emb|AW307241|AW307241 sf54e09.y1 Gm-c1009 Glycine max cDNA clone... 48 1e-06 emblAW334422|AW334422 S34F3 AGS-1 Pneumocystis carinii f. sp. ca... 52 2e-05 emb|AQ842018|AQ842018 T134304 Soybean RFLP probe Glycine max gen... 47 1e-04 40 emb|AV410314|AV410314 AV410314 Lotus japonicus young plants (two... 40 0.010 emb|AW163886|AW163886 Ljirnpest17-312-a12 Ljirnp Lambda HybriZap... 34 0.019 emb|AQ849208|AQ849208 LMAJFV1\_lm43f12.yl Leishmania major FV1 ra... 41 0.042 emb|AW690188|AW690188 NF029E07ST1F1000 Developing stem Medicago ... 35 0.044 emb[Y15086]CFHEPABC Cylindrotheca fusiformis hepA, hepB, hepC ge... 39 0.21 45 emb|AL112906|CNS01AGI Botrytis cinerea strain T4 cDNA library un... 38 0.28 emb|AW725682|AW725682 GA\_Ea0019E05 Gossypium arboreum 7-10 dpa ... 37 0.54 emb|AW727060|AW727060 GA\_Ea0011B07 Gossypium arboreum 7-10 dpa ... 36 1.0 emb|AL034559|PFMAL3P7 Plasmodium falciparum MAL3P7, complete seq... 36 1.4 emb|AF127331|AF127331 Euplotes crassus histone H1-1 (H1-1) gene,... 36 1.4 50 emb|AW729298|AW729298 GA Ea0024K04 Gossypium arboreum 7-10 dpa ... 28 1.6 emb|AL139794|LMFP1105 Leishmania major Friedlin chromosome 4 PAC... 35 1.9 emb|AB028188|AB028188 Penicillium digitatum DNA fragment contain... 35 2.6 emb|Z48093|GVEITS133 G.verna DNA for internal transcribed spacer... 28 3.2 emb|AL353012|SPBC1711 S.pombe chromosome II cosmid c1711. 55 gb|BE036059|BE036059 MO19C02 MO Mesembryanthemum crystallinum cD... 35 3.6 gb|BE036176|BE036176 MO20A07 MO Mesembryanthemum crystallinum cD... 35 3.6 emb|AW693672|AW693672 NF067B05ST1F1044 Developing stem Medicago ... 35 3.6 emb|AJ274013|AJ274013 AJ274013 Metarhizium anisopliae ARSEF 2575... 35 3.6 gb|U80041|AFU80041 Avena fatua Af10-protein mRNA, complete cds. 60 emb|AI612607|AI612607 TENG0370 T. Cruzi epimastigote normalised ... emb|X04693|SOPCG Spinach gene for plastocyanin.

	emb AW010989 AW010989 ST15D02 Pine TriplEx shoot tip library Pin 34 5.0
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	emb AZ215593 AZ215593 Sheared DNA-74E9.TR Sheared DNA Trypanosom 34 6.8
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10	emb AF183445 CLITSYDK1 Codonopsis lanceolata internal transcribe 34 6.8
	emb AC005893 AC005893 Leishmania major chromosome 3 clone L822 s 34 6.8
	gb L06030 TRBVSGH Trypanosoma brucei MVAT5-RX2 variant surface g 34 6.8
	gb L04971 TRBVSGM Trypanosoma brucei variant surface glycoprotei 34 6.8
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	emb AA274264 AA274264 TgESTzz25f05.s1 TgME49 invivo Bradyzoite c 33 9.4
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	/gb_link http://www3.ncbi.nlm.nih.gov/htbin-
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	http://www.ncgr.org/cgi-bin/ff?ac002387
35	(2371 letters)
	(2571 100015)
	Database: plantfungal
	661,018 sequences; 426,114,510 total letters
	)
40	Searchingdone
	Score E
	Sequences producing significant alignments: (bits) Value
	(0.0)
45	emb Z50099 STTKETMR S.tuberosum mRNA for transketolase. 1372 0.0
	gb L76554 SPITRAN Spinacia oleracea transketolase mRNA, chloropl 1350 0.0
	emb Y15781 CAY15781 Capsicum annuum mRNA for plastid transketola 1344 0.0
	emb A52295 A52295 Sequence 1 from Patent EP0723017. 1283 0.0
	emb Z46648 CPTKT7 C.plantagineum tkt7 gene for transketolase. 1063 0.0
50	emb Z46647 CPTKT10 C.plantagineum tkt10 gene for transketolase. 1038 0.0
	emb Z46646 CPTKT3 C.plantagineum tkt3 gene for transketolase. 1014 0.0
	emb AJ249787 CPA249787 Cyanophora paradoxa mRNA for putative tra 557 0.0
	emb AL033501 CAC41C10 C.albicans cosmid Ca41C10. 253 e-175
	emb AC007872 AC007872 The sequence of an Aspergillus parasiticus 167 e-158
55	gb H55032 H55032 HHU58a Sorghum bicolor cv. TX430 Sorghum bicolo 233 e-151
<i>33</i> ···	emb AC005299 AC005299 emericella nidulans chromosome viii cosmid 166 e-147
	emb AC004395 AC004395 Emericella nidulans Chromosome VIII Cosmid 166 e-147
	emb AW443928 AW443928 EST308858 tomato mixed elicitor, BTI Lycop 468 e-131
60	emb AW667519 AW667519 GA
vv	emb AI778813 AI778813 EST259692 tomato susceptible, Cornell Lyco 453 e-126 emb AW776720 AW776720 EST335785 DSU, Medicago truncatula cDNA cl. 450 e-125
	— CHIUZA W 7 / O / ZUIA W 7 / O / ZUI EN 1337 / X3 LINI I. MICHICADO MINCAMIA CLINIA CLI — ASO △ 175

emb|AW695194|AW695194 NF092E10ST1F1082 Developing stem Medicago ... 370 e-124 emb|AI774685|AI774685 EST255785 tomato resistant, Cornell Lycope... 444 e-123 emb|AW618386|AW618386 EST320372 L. pennellii trichome, Cornell U... 434 e-120 5 emb|AW737315|AW737315 EST338838 tomato flower buds, anthesis, Co... 409 e-113 emb|AW694020|AW694020 NF071G12ST1F1099 Developing stem Medicago ... 406 e-112 gb|BE060293|BE060293 HVSMEg0011O04f Hordeum vulgare pre-anthesis... 405 e-112 emb|AW649769|AW649769 EST328223 tomato germinating seedlings, TA... 377 e-111 emb|AW234850|AW234850 sf19h07.y1 Gm-c1028 Glycine max cDNA clone... 399 e-110 10 emb|AW760220|AW760220 sl59e08.y1 Gm-c1027 Glycine max cDNA clone... 388 e-106 emb|AW929526|AW929526 EST338314 tomato flower buds 8 mm to pre-a... 386 e-106 emb|AW596420|AW596420 sj12c01.y1 Gm-c1032 Glycine max cDNA clone... 380 e-104 emb|AI730390|AI730390 BNLGHi6849 Six-day Cotton fiber Gossypium ... 302 e-102 emb|A1778812|A1778812 EST259691 tomato susceptible, Cornell Lyco... 372 e-102 15 emb|AW720123|AW720123 LjNEST14e4r Lotus japonicus nodule library... 369 e-101 emb|AW697121|AW697121 NF115C09ST1F1069 Developing stem Medicago ... 334 e-101 emb|AQ841855|AQ841855 T134114 Soybean RFLP probe Glycine max gen... 358 4e-98 emb|AW564025|AW564025 LG1 281 F10.b1 A002 Light Grown 1 (LG1) So... 359 4e-98 20 emblAW979915|AW979915 EST341564 tomato root deficiency, Cornell ... 349 6e-95 emblAI781665|AI781665 EST262544 tomato susceptible, Cornell Lyco... 348 1e-94 emb|Z26486|PSTKTG P.stipitis TKT gene for transketolase. 261 8e-93 emb|AW039152|AW039152 EST281387 tomato mixed elicitor, BTI Lycop... 339 7e-92 gb|BE053034|BE053034 GA\_Ea0031O21f Gossypium arboreum 7-10 dpa ... 338 9e-92 25 emb|AW691000|AW691000 NF040C09ST1F1000 Developing stem Medicago ... 338 9e-92 emb|AW290149|AW290149 NXNV012F12F Nsf Xylem Normal wood Vertical... 337 2e-91 emb|AW203636|AW203636 sf36e03.y1 Gm-c1028 Glycine max cDNA clone... 332 6e-90 emb|AI966148|AI966148 sc34c04.y1 Gm-c1014 Glycine max cDNA clone... 330 4e-89 emb|Z71255|SCCHRXVI S.cerevisiae chromosome XVI 165536 bp sequen... 225 2e-88 30 gb[U51033]YSCP9513 Saccharomyces cerevisiae chromosome XVI cosmi... 225 2e-88 emb|Z49219|SC9499X S.cerevisiae chromosome XVI cosmid 9499. 225 2e-88 emb|X73224|SCTKL1 S.cerevisiae TKL1 gene for transketolase. 225 2e-88 emb|AL033385|SPBC2G5 S.pombe chromosome II cosmid c2G5. 198 2e-87 emb|AV388967|AV388967 AV388967 Chlamydomonas reinhardtii C9 Chla... 322 8e-87 35 emb|AV391940|AV391940 AV391940 Chlamydomonas reinhardtii C9 Chla... 321 1e-86 emb[AW587453]AW587453 IPPGHZ0039 Cotton fiber and embryo Lambda ... 276 3e-86 gb|U65983|KLU65983 Kluyveromyces lactis transketolase (TKL1) gen... 211 1e-85 emb|AW035937|AW035937 EST282796 tomato callus, TAMU Lycopersicon... 316 5e-85 emb|AW156809|AW156809 se31b02.yl Gm-c1015 Glycine max cDNA clone... 315 1e-84 emb|AA660853|AA660853 00748 MtRHE Medicago truncatula cDNA 5' si... 282 1e-83 40 emb|AV393839|AV393839 AV393839 Chlamydomonas reinhardtii C9 Chla... 230 1e-83 emb|AA556878|AA556878 720 Loblolly pine C Pinus taeda cDNA clone... 289 2e-83 emb|AW689533|AW689533 NF021G12ST1F1000 Developing stem Medicago ... 308 8e-83 emblAW776795]AW776795 EST335860 DSIL Medicago truncatula cDNA cl... 307 2e-82 45 emb|AW694632|AW694632 NF078D05ST1F1045 Developing stem Medicago ... 285 4e-82 emb|AV391577|AV391577 AV391577 Chlamydomonas reinhardtii C9 Chla... 304 1e-81 emb|AV388182|AV388182 AV388182 Chlamydomonas reinhardtii C9 Chla... 304 2e-81 emb|AW688641|AW688641 NF009H10ST1F1000 Developing stem Medicago ... 282 8e-80 emb|AW508848|AW508848 si41b02.yl Gm-r1030 Glycine max cDNA clone... 298 9e-80 50 emb|AW695046|AW695046 NF082H06ST1F1059 Developing stem Medicago ... 297 3e-79 emb|X78993|SCRACII S.cerevisiae genomic DNA 70kb region of the r... 200 3e-79 emb[X73532]SCTKL2 S.cerevisiae gene for transketolase. 200 3e-79 emb|Z35985|SCYBR116C S.cerevisiae chromosome II reading frame OR... 200 3e-79 emb|AW564318|AW564318 LG1\_290\_F06.b1\_A002 Light Grown 1 (LG1) So... 293 4e-78 55 emb|AW201472|AW201472 sf03e09.yl Gm-c1027 Glycine max cDNA clone... 292 6e-78 emb|AB025004|AB025004 Cicer arietinum mRNA for transketolase, pa... 292 8e-78 emb|AW746874|AW746874 WS1 56 C06.b1 A002 Water-stressed 1 (WS1) ... 291 2e-77 emb|AW776336|AW776336 EST335401 DSIL Medicago truncatula cDNA cl... 290 4e-77 emb|AW696241|AW696241 NF104D02ST1F1016 Developing stem Medicago ... 288 1e-76 60 emb|AV395290|AV395290 AV395290 Chlamydomonas reinhardtii C9 Chla... 288 1e-76 emb|AW287516|AW287516 LG1 242 A07.b1 A002 Light Grown 1 (LG1) So... 283 4e-75

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(2037 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

35

40

Searching......done

Score E

Sequences producing significant alignments:

(bits) Value

emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 385 e-125 emb|X98520|BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 343 e-111 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. 348 e-109 emb|Y18260|BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 346 e-109 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 347 e-109 emb|Y14285|BOY14285 Brassica oleracea mRNA for SFR1 protein. 333 e-108 emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 342 e-107 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 328 e-106 gb|M97667|BNASTKR Brassica napus ssp. oleifera serine/threonine ... 334 e-105 gb|U00443|BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 329 e-104 emb|Y14286|BOY14286 Brassica oleracea SFR3 gene, partial. 205 e-104 emb|AB012106|AB012106 Brassica rapa mRNA for SRK45, complete cds. 326 e-103 cb|U00407|BNASPEOKIN Brassica pages S recent to lice at the lice a

gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 330 e-103 gb|M76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 336 e-102 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 325 e-101 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 290 e-100 emb|Y12530|BOARLKGEN B.oleraceae gene encoding serine/threonine ... 131 1e-95 emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 282 3e-94 dbj|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 281 3e-93

dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 276 4e-93 emb|AB000970|AB000970 Brassica campestris gene for receptor kina... 129 1e-91

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40	Database: plantfungal 661,018 sequences; 426,114,510 total letters
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25 (1612 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

30 Searching.....done

Score F

Sequences producing significant alignments: (bits) Value 35 gb[M32885]AVOCYP Avocado cytochrome P-450LXXIA1 (cyp71A1) mRNA, ... 137 e-103 emb[X70981|SMCYPEG2 S.melongena CYP71A1 mRNA for P450 hydroxylase. 127 6e-99 emb[Y09423]NRCYP71A5 Nepeta racemosa mRNA for cytochrome P450, C... 136 8e-98 emb|X71654|SMCYP71B3 S.melongena CYP71A2 mRNA for hydroxylase. dbj|D14990|POTCPEG4 Eggplant mRNA for cytochrome P-450EG4, compl... 124 9e-95 40 emb[Y10489]GMC450CP1 G.max mRNA for putative cytochrome P450, cl... 90 1e-84 emb[X70982|SMCYPEG3 S.melongena CYP71A3 mRNA for P450 hydroxylase. 126 3e-74 emb|Y09424|NRCYP71A6 N.racemosa mRNA for cytochrome P450, CYP71A... 154 3e-65 dbi|E13663|E13663 cDNA encoding cytochrome P450 which is induced... 71 9e-63 dbj[D83968|SOYCYP93A1 Soybean mRNA for cytochrome P450 (CYP93A1)... 71 9e-63 45 emb|AF022157|AF022157 Glycine max cytochrome P450 monooxygenase ... 126 3e-61 emb|AW053855|AW053855 L30-2274T3 Ice plant Lambda Uni-Zap XR exp... 103 1e-53 emb|Y09920|HT7EC0DET Helianthus tuberosus mRNA for 7-ethoxycouma... 94 1e-50 emb[Y10098]HTCYP76B1 H.tuberosus mRNA for 7-ethoxycoumarin O-dee... 94 1e-50 emb|AF022459|AF022459 Glycine max cytochrome P450 monooxygenase ... 105 4e-50 50 emb|AJ238612|CRO238612 Catharanthus roseus mRNA for cytochrome P... 89 2e-49 emb|AF029858|AF029858 Sorghum bicolor cytochrome P450 CYP71E1 (C... 139 3e-49 gb|U48434|SCU48434 Solanum chacoense cytochrome P450 mRNA, compl... 117 2e-48 emb|AF166332|AF166332 Nicotiana tabacum cytochrome P450 gene, co... 114 4e-47 emb|AW223719|AW223719 EST300530 tomato fruit red ripe, TAMU Lyco... 88 4e-46 55 emb|Y10490|GMC450CP3 G.max mRNA for putative cytochrome P450, cl... 104 6e-46 emb|X71658|SMCYPEG8 S.melongena CYP76A1 mRNA. 88 2e-45 emb|AF122821|AF122821 Capsicum annuum cytochrome P450 (PepCYP) m... 118 4e-45 emb|Z33875|CYMPCP450 M.piperita gene for cytochrome P-450 oxidase. 88 7e-45 emb|AI897763|AI897763 EST267206 tomato ovary, TAMU Lycopersicon ... 84 1e-44 60 emb|AW830233|AW830233 sm24f03.y1 Gm-c1028 Glycine max cDNA clone... 139 5e-44 gb U48435 SCU48435 Solanum chacoense putative cytochrome P450 ge... 95 2e-43

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5	emb AW034502 AW034502 EST278118 tomato callus, TAMU Lycopersicon 107 2e-42
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٠.	emb AI484957 AI484957 EST243220 tomato ovary, TAMU Lycopersicon 84 4e-42
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emb|AF134590|AF134590 Papaver somniferum (S)-N-methylcoclaurine ... 83 6e-33 emb|AW255181|AW255181 ML169 peppermint glandular trichome Mentha... 142 6e-33 emb|AF191772|AF191772 Papaver somniferum (S)-N-methylcoclaurine ... 82 1e-32 emb|AI777331|AI777331 EST263739 tomato seed, TAMU Lycopersicon e... 86 1e-32 5 dbj|D85184|D85184 Gentiana triflora mRNA for flavonoid 3',5'-hyd... 72 2e-32 emb[Y10492|GMC450CP5 G.max mRNA for putative cytochrome P450, cl... 67 2e-32 emb|AW733734|AW733734 sk77e06.yl Gm-c1016 Glycine max cDNA clone... 70 2e-32 emb|AW832405|AW832405 sm09e01.yl Gm-c1027 Glycine max cDNA clone... 88 2e-32 emb|AW254778|AW254778 ML1061 peppermint glandular trichome Menth... 141 2e-32 10 emb|AW255299|AW255299 ML307 peppermint glandular trichome Mentha... 88 2e-32 emb|AF195801|AF195801 Medicago sativa isoflavone synthase 2 (ifs... 64 3e-32 emb|AJ243804|CAR243804 Cicer arietinum mRNA for cytochrome P450 ... 66 3e-32 Query= AC004165.66 at 14614 at /id source genbank /description gb|aac16958.1| (ac004165) putative glucosyltransferase [arabidopsis 15 thaliana] /blast score 0 /ec number /family glucosyltransferase /chip nova/gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|ac004165| /ncgi http://www.ncgr.org/cgi-bin/ff?ac004165 20 (1368 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 25 Searching......done Score E Sequences producing significant alignments: (bits) Value 30 emb|AW256802|AW256802 EST304939 KV2 Medicago truncatula cDNA clo... 112 5e-46 emb|AW268009|AW268009 EST306231 DSIR Medicago truncatula cDNA cl... 102 3e-39 emb|AF190634|AF190634 Nicotiana tabacum UDP-glucose:salicylic ac... 101 3e-37 emb|AB000623|AB000623 Nicotiana tabacum mRNA for glucosyl transf... 101 3e-36 emb|AB012116|AB012116 Vigna mungo UFGlyT mRNA for UDP-glycose:fl... 82 2e-34 35 emb[AB013598]AB013598 Verbena hybrida HGT8 mRNA for UDP-glucose:... 104 2e-34 emb|AW776132|AW776132 EST335197 DSIL Medicago truncatula cDNA cl... 99 3e-33 emb|AW459541|AW459541 sh42h03.y1 Gm-c1017 Glycine max cDNA clone... 98 2e-32 emb|AV408145|AV408145 AV408145 Lotus japonicus young plants (two... 85 2e-32 emb|AW216808|AW216808 EST295522 tomato callus, TAMU Lycopersicon... 101 3e-32 40 emb|AW398421|AW398421 EST298268 L. pennellii trichome, Cornell U... 84 6e-32 emb|AW349414|AW349414 GM210007A20D2R Gm-r1021 Glycine max cDNA 3... 97 1e-31 emb|AW035896|AW035896 EST282403 tomato callus, TAMU Lycopersicon... 100 1e-31 gb|BE022282|BE022282 sm72b01.yl Gm-c1028 Glycine max cDNA clone ... 102 4e-31 emb|AB013596|AB013596 Perilla frutescens PF3R4 mRNA for UDP-gluc... 93 7e-31 45 emb|AB013597|AB013597 Perilla frutescens PF3R6 mRNA for UDP-gluc... 93 1e-30 emb|AI771830|AI771830 EST252930 tomato ovary, TAMU Lycopersicon ... 93 1e-30 emb|AB027455|AB027455 Petunia x hybrida PH1 mRNA for anthocyanin... 89 6e-30 emb|AW459258|AW459258 sh22f07.yl Gm-c1016 Glycine max cDNA clone... 69 4e-29 emb|AW928895|AW928895 EST337683 tomato flower buds 8 mm to pre-a... 90 4e-29 50 emb|X85138|LETWII L.esculentum twi1 mRNA. 81 1e-28 emb|AB033758|AB033758 Citrus unshiu LGTase mRNA for limonoid UDP... 89 2e-28 emb[X72729]LEERT1B L.esculentum (ERT 1b) ripening-related mRNA. 87 9e-28 emb|A62529|A62529 Sequence 34 from Patent WO9716559. 96 1e-27 emb|X77461|MECGT2 M.esculenta Crantz CGT2 mRNA for UTP-glucose g... 78 1e-27 55 emb|AQ580287|AQ580287 T135903b shotgun sub-library of BAC clone ... 93 1e-27 emb|X77460|MECGT4 M.esculenta Crantz CGT4 mRNA for UTP-glucose g... 84 2e-27 emb|AW621210|AW621210 EST312008 tomato root during/after fruit s... 98 4e-27 emb|AW036493|AW036493 EST282992 tomato seed, TAMU Lycopersicon e... 92 5e-27 emb|AW695243|AW695243 NF093A08ST1F1056 Developing stem Medicago ... 74 6e-27 60 emblAO368131|AO368131 toxb0001H06r CUGI Tomato BAC Library Lycop... 84 7e-27

emb|AF199453|AF199453 Sorghum bicolor UDP-glucose glucosyltransf... 76 9e-27

	emb AI488782 AI488782 EST247121 tomato ovary, TAMU Lycopersicon 80 1e-26 emb AI729108 AI729108 BNLGHi12670 Six-day Cotton fiber Gossypium 82 1e-26 gb BE126076 BE126076 DG1 65 E03.gl A002 Dark Grown 1 (DG1) Sorgh 106 1e-26
5	dbj D85186 D85186 Gentiana triflora mRNA for UDP-glucose:flavono 86 3e-26 emb X77369 SMGT S.melongena GT mRNA for glycosyltransferase. 97 5e-26
	dbj E12713 E12713 Solanum melongena cDNA encoding flavonoid-3-gl 97 5e-26
	emb AW223197 AW223197 EST300008 tomato fruit red ripe, TAMU Lyco 79 6e-26
	emb AI485027 AI485027 EST243307 tomato ovary, TAMU Lycopersicon 84 6e-26 emb AF127218 AF127218 Forsythia x intermedia flavonoid 3-O-gluco 95 1e-25
10	emb AF101972 AF101972 Phaseolus lunatus zeatin O-glucosyltransfe 87 2e-25
	emb X77462 MECGT5 M.esculenta Crantz CGT5 mRNA for UTP-glucose g 83 2e-25
	emb Y18871 DBE18871 Dorotheanthus bellidiformis mRNA for betanid 81 3e-25
	emb X15694 HVBRNZ1H Barley bronze 1 homologue for UDPglucose fla 80 5e-25 emb AB027454 AB027454 Petunia x hybrida PGT8 mRNA for anthocyani 99 8e-25
15	emb AF116858 AF116858 Phaseolus vulgaris zeatin O-xylosyltransfe 84 8e-25
•	emb AW573699 AW573699 EST316290 GVN Medicago truncatula cDNA clo 78 1e-24'
	emb AW442772 AW442772 EST307702 tomato mixed elicitor, BTI Lycop 84 1e-24
	emb AI898951 AI898951 EST268394 tomato ovary, TAMU Lycopersicon 83 1e-24 emb AW040941 AW040941 EST283805 tomato mixed elicitor, BTI Lycop 83 1e-24
20	emb AW040941 AW040941 EST283805 tomato mixed elicitor, BTI Lycop 83 1e-24 emb AW221893 AW221893 EST298704 tomato fruit red ripe, TAMU Lyco 84 2e-24
	emb AW933199 AW933199 EST359042 tomato fruit mature green, TAMU 93 2e-24
	emb AW981156 AW981156 EST392350 DSIL Medicago truncatula cDNA cl 83 4e-24
	emb AW625781 AW625781 EST319688 tomato radicle, 5 d post-imbibit 84 4e-24 emb X77459 MECGT1 M.esculenta Crantz CGT1 mRNA for UDP-glucose g 66 4e-24
25	emb AW034671 AW034671 EST278402 tomato callus, TAMU Lycopersicon 81 5e-24
	emb AI487283 AI487283 EST245605 tomato ovary, TAMU Lycopersicon 89 1e-23
	emb AQ580273 AQ580273 T135889b shotgun sub-library of BAC clone 93 2e-23
	emb AW329251 AW329251 N200469e rootphos(-) Medicago truncatula c 93 2e-23 emb AI780684 AI780684 EST261479 tomato susceptible, Cornell Lyco 79 2e-23
30	emb AI486974 AI486974 EST245296 tomato ovary, TAMU Lycopersicon 83 3e-23
	emb AI483783 AI483783 EST249654 tomato ovary, TAMU Lycopersicon 79 5e-23
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	emb AW649880 AW649880 EST328334 tomato germinating seedlings, TA 87 9e-23 emb AJ400861 CAR400861 Cicer arietinum partial mRNA for putative 80 1e-22
35	emb X77463 MECGT6 M.esculenta Crantz CGT6 mRNA for UTP-glucose g 74 2e-22
	emb AI487582 AI487582 EST245904 tomato ovary, TAMU Lycopersicon 82 2e-22
	emb AW759836 AW759836 sl54f11.yl Gm-c1027 Glycine max cDNA clone 91 2e-22 emb AW350921 AW350921 GM210010B10C8R Gm-r1021 Glycine max cDNA 3 82 3e-22
	emb AF165148 AF165148 Petunia x hybrida UDP-galactose:flavonol 3 86 4e-22
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	emb AB009370 AB009370 Vigna mungo UF3GaT mRNA for flavonoid 3-O 80 1e-21 emb AF000372 AF000372 Vitis vinifera UDP glucose:flavonoid 3-o-g 85 1e-21
	emb AF000371 AF000371 Vitis vinifera UDP glucose:flavonoid 3-o-g 85 1e-21
م	emb X77464 MECGT7 M.esculenta Crantz CGT7 mRNA for UTP-glucose g 88 1e-21
45	emb AB012114 AB012114 Vigna mungo UFGlyT mRNA for UDP-glycose:fl 73 2e-21 emb AI894979 AI894979 EST264422 tomato callus, TAMU Lycopersicon 74 4e-21
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50	gb U82367 STU82367 Solanum tuberosum UDP-glucose glucosyltransfe 73 9e-21
50	emb AW650188 AW650188 EST328642 tomato germinating seedlings, TA 89 9e-21 emb AW617410 AW617410 EST323821 L. hirsutum trichome, Cornell Un 81 1e-20
	emb AF006081 AF006081 Solanum berthaultii UDPG glucosyltransfera 69 3e-20
	emb AI487484 AI487484 EST245806 tomato ovary, TAMU Lycopersicon 74 3e-20
55	emb AW776615 AW776615 EST335680 DSIL Medicago truncatula cDNA cl 100 4e-20
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	emb AW317745 AW317745 sg56g04.yl Gm-c1007 Glycine max cDNA clone 57 8e-20
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.60	emb AW092049 AW092049 EST285325 tomato mixed elicitor, BTI Lycop 88 4e-19
.00	gb BE055475 BE055475 GA_Ea0035112f Gossypium arboreum 7-10 dpa 69 5e-19 emb AF117267 AF117267 Malus domestica UDP glucose:flavonoid 3-O 80 7e-19

emb|AW781424|AW781424 s178a07.y1 Gm-c1037 Glycine max cDNA clone... 95 9e-19

emb|Z25802|PHUDPRHAX P.hybrida mRNA for UDP rhamnose: anthocyani... 68 9e-19 emb|AI967768|AI967768 Ljimpest11-882-f3 Ljimp Lambda HybriZap ... 95 1e-18 emb|AB002818|AB002818 Perilla frutescens mRNA for flavonoid 3-O-... 90 1e-18 5 emblAB012115|AB012115 Vigna mungo UFGlyT mRNA for UDP-glycose:fl... 89 1e-18 emb|AI485737|AI485737 EST244058 tomato ovary, TAMU Lycopersicon ... 71 2e-18 emb|AW677595|AW677595 DG1 8 E07.g1 A002 Dark Grown 1 (DG1) Sorgh... 94 2e-18 10 Query= AL021961.3 at 15042 at /id source genbank /description emb|caa17549.1| (al021961) cinnamyl alcohol dehydrogenase - like protein [arabidopsis thaliana] /blast score 1.00e-167 /ec number /family /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|al021961| /ncgi 15 http://www.ncgr.org/cgi-bin/ff?al021961 (1074 letters) Database: plantfungal 20 661,018 sequences; 426,114,510 total letters Searching......done Score Ε 25 Sequences producing significant alignments: (bits) Value dbj|D13991|AAICAD A. cordata mRNA for cinnamyl alcohol dehydroge... 279 e-152 emb[X62343]NTCAD14MR N.tabacum CAD14 mRNA for cinnamyl alcohol d... 314 e-150 emb|Z19568|PDCIALDHA P.deltoides encoding cinnamyl alcohol dehyd... 310 e-146 30 emb|A24083|A24083 pPOPCAD1 cinnamyl alcohol dehydrogenase cDNA. emb|AF217957|AF217957 Populus tremuloides cinnamyl alcohol dehyd... 309 e-145 emb[X62344]NTCAD19MR N.tabacum CAD19 mRNA for cinnamyl alcohol d... 302 e-145 emb|AF038561|AF038561 Eucalyptus globulus cinnamyl alcohol dehyd... 318 e-141 emb[X65631]EGCADMR E.gunnii mRNA cad for cinnamyl alcohol dehydr... 319 e-141 35 emb|A24084|A24084 pEUCAD1 cinnamyl alcohol dehydrogenase cDNA. 319 e-141 emb|AF083332|AF083332 Medicago sativa cinnamyl-alcohol dehydroge... 326 e-140 emb[Z19573]MSCIALDHA M.sativa encoding cinnamyl alcohol dehydrog... 325 e-140 emb|AJ231135|SOF231135 Saccharum officinarum mRNA for cinnamyl a... 384 e-134 emb|AF010290|AF010290 Lolium perenne cinnamyl alcohol dehydrogen... 262 e-133 40 emb|X72675|PACINALDA P.abies mRNA for cinnamyl alcohol dehydroge... 240 e-124 dbi|D86590|D86590 Zinnia elegans mRNA for cinnamyl alcohol dehyd... 264 e-123 gb|U62394|PRU62394 Pinus radiata cinnamyl alcohol dehydrogenase ... 242 e-122 emb|Z37992|PTCADBMR P.taeda mRNA for cinnamyl alcohol dehydrogen... 237 e-121 emb|Z37991|PTCADAMR P.taeda mRNA for cinnamyl alcohol dehydrogen... 237 e-121 45 gb|U63534|FXU63534 Fragaria x ananassa cinnamyl alcohol dehydrog... 251 9e-89 emb|AF083333|AF083333 Medicago sativa cinnamyl-alcohol dehydroge... 225 5e-84 gb|L36823|SSNCAD1A Stylosanthes humilis cinnamyl alcohol dehydro... 226 9e-84 gb|U24561|AGU24561 Apium graveolens mannitol dehydrogenase (Mtd)... 237 3e-79 emb|X67817|PCELI3 P.crispum mRNA for Eli3. 240 3e-77 50 emb|X92855|LEMTD L.esculentum exon 1 of MTD gene. 130 6e-77 emb|AW255506|AW255506 ML532 peppermint glandular trichome Mentha... 275 6e-73 emb|AI487388|AI487388 EST245710 tomato ovary, TAMU Lycopersicon ... 274 8e-73 emb|AF207552|AF207552 Brassica napus cinnamyl alcohol dehydrogen... 155 2e-72 emb|AF207553|AF207553 Brassica napus cinnamyl alcohol dehydrogen... 156 5e-72 emblAF207555|AF207555 Brassica rapa cinnamyl alcohol dehydrogena... 155 1e-71 55 emb|AF207554|AF207554 Brassica oleracea cinnamyl alcohol dehydro... 156 1e-71 emb|X92754|HVCADMRNA H.vulgare mRNA for cinnamyl alcohol dehydro... 270 2e-71 emb|AI489728|AI489728 EST248067 tomato ovary, TAMU Lycopersicon ... 269 3e-71 emblAW830216|AW830216 sm24b12.yl Gm-c1028 Glycine max cDNA clone... 264 6e-70 emb|AW350274|AW350274 GM210007B20B7R Gm-r1021 Glycine max cDNA 3... 262 1e-69 60 gb[U79770]MCU79770 Mesembryanthemum crystallinum cinnamyl-alcoho... 255 4e-69

emblAW031628|AW031628 EST275082 tomato callus, TAMU Lycopersicon... 182 7e-69 gb|BE021646|BE021646 sm60f08.yl Gm-c1028 Glycine max cDNA clone ... 259 4e-68 gb|BE123932|BE123932 EST394057 DSIL Medicago truncatula cDNA clo... 185 7e-65 emb|AW560160|AW560160 EST315208 DSIR Medicago truncatula cDNA cl... 169 3e-64 5 emb|AW684815|AW684815 NF021D07NR1F1000 Nodulated root Medicago t... 230 le-62 emb|AI162401|AI162401 A017P09U Hybrid aspen plasmid library Popu... 178 5e-62 emb|AI900147|AI900147 sc01e08.yl Gm-c1012 Glycine max cDNA clone... 142 2e-61 emb|AW101492|AW101492 sd78h08.y1 Gm-c1009 Glycine max cDNA clone... 176 3e-61 emb|AW568106|AW568106 si68e05.yl Gm-r1030 Glycine max cDNA clone... 136 3e-59 10 emb|AF146691|AF146691 Lycopersicon esculentum cultivar Rio Grand... 194 1e-58 emb|AI443116|AI443116 sa84e07.yl Gm-c1004 Glycine max cDNA clone... 170 1e-54 emb|AJ001925|PAAJ1925 Picea abies cad7 gene. 115 le-53 emb|AJ001926|PAAJ1926 Picea abies cad8 gene. 115 1e-53 emb|AJ001924|PAAJ1924 Picea abies cad2 gene. 114 le-53 15 emb|AI488134|AI488134 EST246456 tomato ovary, TAMU Lycopersicon ... 210 2e-53 emb|AW981164|AW981164 EST392358 DSIL Medicago truncatula cDNA cl... 133 3e-53 emb|AI965672|AI965672 sc76g06.yl Gm-c1018 Glycine max cDNA clone... 209 4e-53 emb|AW981208|AW981208 EST392298 DSIL Medicago truncatula cDNA cl... 149 3e-52 emb|AW692803|AW692803 NF059G11ST1F1087 Developing stem Medicago ... 125 4e-52 20 gb|BE123743|BE123743 NXNV\_153 E12\_F Nsf Xylem Normal wood Vertic... 187 4e-49 emb|AL121862|LMFL1063 Leishmania major Friedlin chromosome 23 co... 73 8e-48 emb|AF109157|AF109157 Eucalyptus globulus cinnamyl alcohol dehyd... 138 2e-46 emb|X75480|EGCAD E.gunnii CAD gene. 139 3e-46 dbj|D16624|EGCCAD Eucalyptus botryoides DNA for cinnamyl alcohol... 138 2e-45 25 emb|AW775567|AW775567 EST334632 DSIL Medicago truncatula cDNA cl... 105 7e-45 gb|L36456|SSNCAD3 Stylosanthes humilis cinnamyl alcohol dehydrog... 134 le-44. emb|AW706922|AW706922 sk08c02.y1 Gm-c1023 Glycine max cDNA clone... 113 1e-43 gb|BE123884|BE123884 EST394009 DSIL Medicago truncatula cDNA clo... 110 2e-43 emb|AW201173|AW201173 se98g05.yl Gm-c1027 Glycine max cDNA clone... 135 3e-43 30 emb|AI729035|AI729035 BNLGHi12406 Six-day Cotton fiber Gossypium... 174 9e-43 emb|AW666266|AW666266 sk34e03.y1 Gm-c1028 Glycine max cDNA clone... 108 2e-42 emb|AW218543|AW218543 EST303726 tomato radicle, 5 d post-imbibit... 88 6e-42 emb|AW100321|AW100321 sd23a02.y1 Gm-c1012 Glycine max cDNA clone... 130 2e-41 emb|AI727983|AI727983 BNLGHi9898 Six-day Cotton fiber Gossypium ... 95 2e-40 35 emb|AV412798|AV412798 AV412798 Lotus japonicus young plants (two... 137 6e-40 emb|AI731536|AI731536 BNLGHi9665 Six-day Cotton fiber Gossypium ... 94 8e-40 emb|AW775594|AW775594 EST334659 DSIL Medicago truncatula cDNA cl... 98 8e-40 emb|AI726959|AI726959 BNLGHi6955 Six-day Cotton fiber Gossypium ... 93 4e-39 gb|BE124304|BE124304 EST394429 DSIL Medicago truncatula cDNA clo... 94 7e-39 40 emb|AW776649|AW776649 EST335714 DSIL Medicago truncatula cDNA cl... 94 1e-38 emb|AI728121|AI728121 BNLGHi9734 Six-day Cotton fiber Gossypium ... 93 2e-38 emb|AF060491|AF060491 Pinus radiata cinnamyl alcohol dehydrogena... 118 2e-38 gb|BE036055|BE036055 MO19B09 MO Mesembryanthemum crystallinum cD... 81 4e-38 emb|AI161452|AI161452 A001P14U Hybrid aspen plasmid library Popu... 125 7e-38 45 emb|AW648829|AW648829 EST327283 tomato germinating seedlings, TA... 89 3e-37 gb|BE124289|BE124289 EST394414 DSIL Medicago truncatula cDNA clo... 154 1e-36 emb|AW348888|AW348888 GM210010A10H2R Gm-r1021 Glycine max cDNA 3... 154 1eemb|AI496395|AI496395 sb04b10.y1 Gm-c1004 Glycine max cDNA clone... 154 1e-36 50 emb|AW684143|AW684143 NF012H08NR1F1000 Nodulated root Medicago t... 121 3e-36 emblAI899871|AI899871 sb94e03.y1 Gm-c1017 Glycine max cDNA clone... 100 3e-36 emb|AW650552|AW650552 EST329006 tomato germinating seedlings, TA... 148 4e-36 emb|AW928461|AW928461 EST337249 tomato flower buds 8 mm to pre-a... 89 3e-34 emb|AV417794|AV417794 AV417794 Lotus japonicus young plants (two... 140 1e-32 emb|AF067082|AF067082 Apium graveolens mannitol dehydrogenase (M... 127 9e-32 emb|AW234172|AW234172 sf22a12.yl Gm-c1028 Glycine max cDNA clone... 134 1e-30 emb|AI165779|AI165779 A091p26u Hybrid aspen plasmid library Popu... 94 2e-30 emblAW686753|AW686753 NF042C07NR1F1000 Nodulated root Medicago t... 93 2e-30 emb|AW256719|AW256719 EST304856 KV2 Medicago truncatula cDNA clo... 78 3e-30 60 emb|AW625821|AW625821 EST319728 tomato radicle, 5 d post-imbibit... 132 4e-30 emb|AI166747|AI166747 xylem.est.549 Poplar xylem Lambda ZAPII li... 94 5e-30

emb|AW218822|AW218822 EST301302 tomato root during/after fruit s... 84 2e-29

emb|AI960981|AI960981 sc93e08.y1 Gm-c1019 Glycine max cDNA clone... 85 4e-29 emb|AW617779|AW617779 EST324178 L. hirsutum trichome, Cornell Un... 89 1e-28 emb|AI165405|AI165405 A083p34u Hybrid aspen plasmid library Popu... 127 1e-28 5 emb|AW648072|AW648072 EST326526 tomato germinating seedlings, TA... 79 2e-27 Query= AL078637.213 s at 15523 s at /id source genbank /description emb|cab45071.1| (al078637) putative protein [arabidopsis thaliana] 10 /blast\_score 8.00e-48 /ec\_number /family /chip nova /gb\_link /ncgi (336 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 15 Searching......done Score Ε Sequences producing significant alignments: (bits) Value 20 emb|AW394463|AW394463 sh32h06.yl Gm-c1017 Glycine max cDNA clone... 80 1e-17 gb|BE023782|BE023782 sm92d02.yl Gm-c1015 Glycine max cDNA clone ... 73 2e-15 emb|AW395908|AW395908 sh07c05.yl Gm-c1016 Glycine max cDNA clone... 80 7e-15 emb|AW733452|AW733452 sk73g05.y1 Gm-c1016 Glycine max cDNA clone... 77 4e-14 25 gb[BE058639]BE058639 sn18g09.y1 Gm-c1016 Glycine max cDNA clone ... 77 6e-14 emblAW156195|AW156195 se20g08.y1 Gm-c1015 Glycine max cDNA clone... 76 1e-13 emb|AW563817|AW563817 LG1\_261\_C02.g1\_A002 Light Grown 1 (LG1) So... 76 1e-13 emb|AW677327|AW677327 DG1\_5\_D03.g1\_A002 Dark Grown 1 (DG1) Sorgh... 76 1e-13 emb|AW424028|AW424028 sh59f09.y1 Gm-c1015 Glycine max cDNA clone... 62 1e-12 30 gb|BE022753|BE022753 sm88a02.yl Gm-c1015 Glycine max cDNA clone ... 62 2e-12 emb|AW677332|AW677332 DG1\_5\_F03.g1\_A002 Dark Grown 1 (DG1) Sorgh... 71 3e-12 gb|BE035944|BE035944 MO22E07 MO Mesembryanthemum crystallinum cD... 69 1e-11 emb|AW119934|AW119934 sd54g12.y1 Gm-c1016 Glycine max cDNA clone... 55 3e-10 gb|L20233|POPVEGSTRA P.trichocarpa x P.deltoides vegetative stor... 64 4e-10 35 emb|AW119941|AW119941 sd54h12.y1 Gm-c1016 Glycine max cDNA clone... 47 4e-08 emb|AW092074|AW092074 EST285350 tomato mixed elicitor, BTI Lycop... 57 4e-08 emb|AW127599|AW127599 M110330 DSLC Medicago truncatula cDNA clon... 43 5e-07 emb|AI822191|AI822191 L0-668T3 Ice plant Lambda Uni-Zap XR expre... 52 6e-07 gb|BE035204|BE035204 MO02A04 MO Mesembryanthemum crystallinum cD... 51 2e-06 40 gb|BE035225|BE035225 MO02E03 MO Mesembryanthemum crystallinum cD... 51 2e-06 gb|BE036341|BE036341 MO23E06 MO Mesembryanthemum crystallinum cD... 51 2e-06 gb[BE037208]BE037208 MP18A03 MP Mesembryanthemum crystallinum cD... 51 2e-06 gb|BE037538|BE037538 MP19G11 MP Mesembryanthemum crystallinum cD... 48 9e-06 gb|BE034872|BE034872 ML05E09 ML Mesembryanthemum crystallinum cD... 45 2e-04 45 emb|AW458345|AW458345 sh86h01.yl Gm-c1016 Glycine max cDNA clone... 31 9e-04 emb|AU036852|AU036852 AU036852 Cryptomeria japonica seedling lea... 41 0.003 gb|S59422|S59422 Populus x canadensis major storage protein mRNA... 41 0.003 gb|M77504|POPBSP Populus deltoides bark storage protein mRNA, co... 41 0.003 emb|AC000133|ENAC000133 Emericella nidulans cosmid SW06E08, comp... 33 0.77 50 emb|AW287856|AW287856 N100699e rootphos(-) Medicago truncatula c... 31 1.0 emb|AQ851083|AQ851083 LMAJFV1 lm38d08,y1 Leishmania major FV1 ra... 33 1.1 emb|AW329091|AW329091 N200297e rootphos(-) Medicago truncatula c... 31 1.2 emb|AW329193|AW329193 N200405e rootphos(-) Medicago truncatula c... 31 1.3 emb|AQ398028|AQ398028 mgxb0017N12f CUGI Rice Blast BAC Library P... 32 1.4 55 emb|AI068626|AI068626 mgae0003cD01f Magnaporthe grisea Appressor... 32 1.4 emb|AW981194|AW981194 EST392388 DSIL Medicago truncatula cDNA cl... 31 1.5 emb|AW695905|AW695905 NF100E05ST1F1038 Developing stem Medicago ... 31 1.5 dbj|D50414|D50414 Cannabis sativa male-associated DNA sequence. 29 1.7 emb|X70064|PDBSPA P.deltoides gene for poplar bark storage protein. 32 2.0 60 emb|Z64354|SPAC23D3 S.pombe chromosome I cosmid c23D3. 32 2.0 emb|X83853|NTPAR1A N.tabacum mRNA for PAR-1a. 32 2.0

	emb AW268020 AW268020 EST306242 DSIR Medicago truncatula cDNA cl 32 2.0 emb AA550547 AA550547 1710m3 gmbPfHB3.1, G. Roman Reddy Plasmodi 32 2.0 emb AL078627 SPBC365 S.pombe chromosome II cosmid c365. 31 2.7
_	gb U05613 CBU05613 Crepidomanes birmanicum chloroplast large sub 31 3.7
5	emb AI050216 AI050216 TENU1464 T. cruzi epimastigote normalized 31 3.7
	emb AI065375 AI065375 TENU2262 T. cruzi epimastigote normalized 31 3.7 emb AI065371 AI065371 TENU2258 T. cruzi epimastigote normalized 31 3.7
	• i •
	emb AT000167 AT000167 AT000167 Apple young fruit cDNA library Ma 29 4.0 emb AJ388714 AJ388714 AJ388714 Medicago truncatula R108 Medicago 29 4.3
10	emb AQ361816 AQ361816 mgxb0005A07f CUGI Rice Blast BAC Library P 30 5.1
-0	emb AQ162953 AQ162953 mgxb0021G21r CUGI Rice Blast BAC Library P 30 5.1
	emb Z67998 SPAC1F7 S.pombe chromosome I cosmid c1F7. 30 5.1
	emb AF148676 AF148676 Zizaniopsis villanensis maturase (matK) ge 30 7.1
	emb AW684956 AW684956 NF023E05NR1F1000 Nodulated root Medicago t 30 7.1
15	emb AZ216404 AZ216404 Sheared DNA-121G9.TF Sheared DNA Trypanoso 30 7.1
	emb AQ942697 AQ942697 Sheared DNA-42J8.TR Sheared DNA Trypanosom 30 7.1
•	emb AI163063 AI163063 A031p30u Hybrid aspen plasmid library Popu 30 7.1
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••	gb BE023592 BE023592 sm82e07.yl Gm-c1015 Glycine max cDNA clone 30 7.1
20	emb AZ220034 AZ220034 Sheared DNA-63D11.TR Sheared DNA Trypanoso 30 7.1
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	emb AI057841 AI057841 TENU1932 T. cruzi epimastigote normalized 30 7.1
25	emb X83851 NTPAR1B N.tabacum mRNA for PAR-1b. 30 7.1 gb M18538 POPPOP3A Populus balsamifera subsp. trichocarpa X Populus 30 7.1
2.5	emb AW203632 AW203632 sf36d11.yl Gm-c1028 Glycine max cDNA clone 30 7.1
	emb AQ640232 AQ640232 927P1-18B11.TV 927P1 Trypanosoma brucei ge 30 7.1
	emb AI779122 AI779122 EST260001 tomato susceptible, Cornell Lyco 27 8.3
	emb AW096641 AW096641 EST289821 tomato mixed elicitor, BTI Lycop 27 8.9
30	gb[N98085]N98085 2245C3 czapPFDd2.1, Debopam Chakrabarti Plasmod 29 9.7
	emb AW929160 AW929160 EST337948 tomato flower buds 8 mm to pre-a 29 9.7
	emb AW037807 AW037807 EST279436 tomato mixed elicitor, BTI Lycop 29 9.7
	emb AW729409 AW729409 GA_Ea0024O24 Gossypium arboreum 7-10 dpa 29 9.7
2 6	emb AF193903 AF193903 Cafeteria roenbergensis mitochondrial DNA, 29 9.7
35	
	Query= AL078637.191 r at 15532 r at /id source genbank /description
	emb[cab45069.1] (al078637) putative protein [arabidopsis thaliana]
	/blast_score 0 /ec_number /family /chip nova /gb_link
40	http://www3.ncbi.nlm.nih.gov/htbin-
	post/entrez/query?db=n&form=6&dopt=g&uid=gb al078637  /ncgi
	http://www.ncgr.org/cgi-bin/ff?al078637
	(990 letters)
45	Database: plantfungal
	661,018 sequences; 426,114,510 total letters
	Searching
	Searchingdone
50	Score E
- •	Sequences producing significant alignments: (bits) Value
•	(214)
	emb AW092074 AW092074 EST285350 tomato mixed elicitor, BTI Lycop 145 1e-69
	emb AI778762 AI778762 EST259641 tomato susceptible, Cornell Lyco 149 1e-49
55	gb BE035944 BE035944 MO22E07 MO Mesembryanthemum crystallinum cD 120 9e-49
	emb AW618184 AW618184 EST314234 L. pennellii trichome, Cornell U 138 2e-46
	emb AI778761 AI778761 EST259640 tomato susceptible, Cornell Lyco 133 6e-45
	emb AW618179 AW618179 EST314229 L. pennellii trichome, Cornell U 114 3e-39
60	gb S59422 S59422 Populus x canadensis major storage protein mRNA 99 6e-32
UU	gb M77504 POPBSP Populus deltoides bark storage protein mRNA, co 99 3e-30
	emb AW395908 AW395908 sh07c05.yl Gm-c1016 Glycine max cDNA clone 125 5e-28

emb|AW733452|AW733452 sk73g05.y1 Gm-c1016 Glycine max cDNA clone... 122 3e-27 gb|BE058639|BE058639 sn18g09.y1 Gm-c1016 Glycine max cDNA clone ... 122 4e-27 emb|AW394463|AW394463 sh32h06.y1 Gm-c1017 Glycine max cDNA clone... 111 4e-26 emb|AW563817|AW563817 LG1 261\_C02.g1\_A002 Light Grown 1 (LG1) So... 104 4e-25 5 emblAI822191|AI822191 LO-668T3 Ice plant Lambda Uni-Zap XR expre... 114 1e-24 gb|BE036341|BE036341 MO23E06 MO Mesembryanthemum crystallinum cD... 112 3e-24 gb|BE035204|BE035204 MO02A04 MO Mesembryanthemum crystallinum cD... 112 3e-24 gb|BE037538|BE037538 MP19G11 MP Mesembryanthemum crystallinum cD... 112 3e-24 gb|BE035225|BE035225 MO02E03 MO Mesembryanthemum crystallinum cD... 112 3e-24 10 gb|BE059090|BE059090 sn25b08.y1 Gm-c1016 Glycine max cDNA clone ... 75 2e-23 emb|AW395168|AW395168 sh40g07.yl Gm-c1017 Glycine max cDNA clone... 68 3e-21 gb|BE037512|BE037512 MP04B06 MP Mesembryanthemum crystallinum cD... 95 4e-20 emb|AW623629|AW623629 EST321574 tomato flower buds 3-8 mm, Corne... 92 3e-19 emb|AW625587|AW625587 EST319494 tomato radicle, 5 d post-imbibit... 92 3e-19 15 emb|AW929488|AW929488 EST338276 tomato flower buds 8 mm to pre-a... 92 3e-19 emb|AI778197|AI778197 EST259076 tomato susceptible, Cornell Lyco... 92 3e-19 emb|AW648720|AW648720 EST327090 tomato germinating seedlings, TA... 92 3e-19 emb|AW625287|AW625287 EST319290 tomato radicle, 5 d post-imbibit... 92 3e-19 emb|AW648718|AW648718 EST327088 tomato germinating seedlings, TA... 92 3e-19 20 emb|AW928879|AW928879 EST337667 tomato flower buds 8 mm to pre-a... 92 3e-19 gb|L20233|POPVEGSTRA P.trichocarpa x P.deltoides vegetative stor... 94 2e-18 emb|AW733620|AW733620 sk75h08.y1 Gm-c1016 Glycine max cDNA clone... 86 3e-18 emb|AW677327|AW677327 DG1 5 D03.gl A002 Dark Grown 1 (DG1) Sorgh... 93 4e-18 emb|AI163910|AI163910 A051P26U Hybrid aspen plasmid library Popu... 56 1e-17 25 emb|AW156195|AW156195 se20g08.yl Gm-c1015 Glycine max cDNA clone... 90 2e-17 emb|AW775932|AW775932 EST334997 DSIL Medicago truncatula cDNA cl... 84 2e-17 emb|AW692820|AW692820 NF056A09ST1F1000 Developing stem Medicago ... 84 2e-17 emb|AW685340|AW685340 NF027C09NR1F1000 Nodulated root Medicago t... 83 4e-17 emblAW684469|AW684469 NF017C09NR1F1000 Nodulated root Medicago t... 75 1e-16 30 gb|BE037208|BE037208 MP18A03 MP Mesembryanthemum crystallinum cD... 84 2e-15 gb|BE023782|BE023782 sm92d02.y1 Gm-c1015 Glycine max cDNA clone ... 72 4e-14 gb|BE036234|BE036234 MO21C12 MO Mesembryanthemum crystallinum cD... 78 1e-13 emb|AW286123|AW286123 LG1\_261\_F11.b1\_A002 Light Grown 1 (LG1) So... 64 5e-13 emb|AW626069|AW626069 EST319976 tomato radicle, 5 d post-imbibit... 69 1e-12 35 gb|BE095283|BE095283 00344 leafy spurge Lambda HybriZAP 2.1 two-... 62 1e-12 emb|AW677160|AW677160 DG1\_5 D03.b1 A002 Dark Grown 1 (DG1) Sorgh... 62 2e-12 emb|AW677332|AW677332 DG1 5 F03.g1 A002 Dark Grown 1 (DG1) Sorgh... 70 2e-11 gb|BE022753|BE022753 sm88a02.yl Gm-c1015 Glycine max cDNA clone ... 61 5e-11 emb|AW424028|AW424028 sh59f09.yl Gm-c1015 Glycine max cDNA clone... 62 8e-11 40 emb[X70064]PDBSPA P.deltoides gene for poplar bark storage protein. 63 3e-09 emb|AW119934|AW119934 sd54g12.y1 Gm-c1016 Glycine max cDNA clone... 54 4e-09 emb|AW677125|AW677125 DG1\_5\_F03.b1\_A002 Dark Grown 1 (DG1) Sorgh... 43 5e-07 gb|BE058421|BE058421 sn15h03.yl Gm-c1016 Glycine max cDNA clone ... 56 6e-07 emb|AW119941|AW119941 sd54h12.y1 Gm-c1016 Glycine max cDNA clone... 46 1e-06 45 gb|BE037054|BE037054 MP13H01 MP Mesembryanthemum crystallinum cD... 55 1e-06 gb|BE036004|BE036004 MO17H11 MO Mesembryanthemum crystallinum cD... 51 1e-06 gb|BE035382|BE035382 MO03G01 MO Mesembryanthemum crystallinum cD... 54 2e-06 emb|AW127599|AW127599 M110330 DSLC Medicago truncatula cDNA clon... 44 3e-06 gb|BE095282|BE095282 00343 leafy spurge Lambda HybriZAP 2.1 two-... 52 5e-06 50 emb|AW287592|AW287592 LG1 244 A09.b1 A002 Light Grown 1 (LG1) So... 40 6e-06 emb|AA557101|AA557101 943 Loblolly pine N Pinus taeda cDNA clone... 47 2e-04 gb|BE037091|BE037091 MP14F10 MP Mesembryanthemum crystallinum cD... 47 2e-04 gb|M25340|POPCHIC Populus sp. chitinase mRNA fragment, clone 4. 46 6e-04 gb|BE036753|BE036753 MP04H07 MP Mesembryanthemum crystallinum cD... 46 6e-04 55 gb|BE037437|BE037437 MP21A08 MP Mesembryanthemum crystallinum cD... 34 8e-04 gb|BE037490|BE037490 MP21G12 MP Mesembryanthemum crystallinum cD... 45 9e-04 emb|AU036852|AU036852 AU036852 Cryptomeria japonica seedling lea... 41 0.015 emb|AA556425|AA556425 280 Loblolly pine C Pinus taeda cDNA clone... 41 0.015 gb|BE036626|BE036626 MP02H12 MP Mesembryanthemum crystallinum cD... 40 0.021 60 emb|AW458345|AW458345 sh86h01.yl Gm-c1016 Glycine max cDNA clone... 30 0.052 gb|BE034872|BE034872 ML05E09 ML Mesembryanthemum crystallinum cD... 38 0.10

	emb AW729858 AW729858 GAEa0026H02 Gossypium arboreum 7-10 dpa 36 0.3 emb AQ953583 AQ953583 Sheared DNA-53H18.TF Sheared DNA Trypanoso 36 0.5
	gb BE036480 BE036480 MP03G10 MP Mesembryanthemum crystallinum cD 36 0.51
5	emb Z00044 CHNTXX Nicotiana tabacum chloroplast genome DNA. 35 0.70
J	emb AL160939 L1356CX Leishmania major Friedlin cosmid L1356.3 t3 34 1.8 gb BE036897 BE036897 MP08B09 MP Mesembryanthemum crystallinum cD 34 1.8
	gb BE036897 BE036897 MP08B09 MP Mesembryanthemum crystallinum cD 34 1.8 emb AI730722 AI730722 BNLGHi7729 Six-day Cotton fiber Gossypium 34 1.8
	emb AB030726 AB030726 Nicotiana tabacum mRNA for DNA (cytosine-5 34 2.5
	emb AW397331 AW397331 sg77e08.y1 Gm-c1007 Glycine max cDNA clone 34 2.5
10	emb AI210350 AI210350 i0c03a1.rl Aspergillus nidulans 24hr asexu 34 2.5
	emb AA786346 AA786346 13g09a1.fl Aspergillus nidulans 24hr asexu 34 2.5
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	emb AW672119 AW672119 LG1_357_F07.b1_A002 Light Grown 1 (LG1) So 33 3.4
15	emb AJ270207 ECA270207 Entodinium caudatum partial mRNA fro puta 33 3.4
	emb AW672133 AW672133 LG1_357_D07.b1_A002 Light Grown 1 (LG1) So 33 3.4
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	emb AZ212142 AZ212142 Sheared DNA-70G10.TF Sheared DNA Trypanoso 32 6.4
20	emb AI163630 AI163630 A045p06u Hybrid aspen plasmid library Popu 32 6.4
20	emb AL354532 LMFL1177 Leishmania major Friedlin chromosome 21 co 30 6.9 emb AJ243516 NCR243516 Neurospora crassa partial nca-3 gene for 32 8.8
	emb AL116648 CNS01DCG Botrytis cinerea strain T4 cDNA library un 27 9.3
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25	/blast score 0 /ec number /family /chip nova /gb link /ncgi
	(1236 letters)
	Database: plantfungal
• •	661,018 sequences; 426,114,510 total letters
30	
	Searchingdone
	Score E
	Sequences producing significant alignments: (bits) Value
35	to openious producing significant anginionis. (Dis) value
	gb M60729 HRAHRPCC A.rusticana peroxidase isoenzyme C (HPR C) ge 744 0.0
	emb A00741 A00741 A.rusticana synthetic gene (reverse complement 692 0.0
	emb A00740 A00740 A.rusticana synthetic gene for peroxidase. 692 0.0
	dbj E01651 E01651 cDNA encoding horseradish peroxidase. 692 0.0
40	gb[M37157[HRAHRPCB A.rusticana peroxidase isoenzyme C (HPR C) ge 371 e-133
	emb X97349 PTPXP2PER P.trichocarpa mRNA for anionic peroxidase P 473 e-132
	emb X97350 PTPXP3PER P.trichocarpa mRNA for anionic peroxidase P 469 e-131
	dbj D83224 POPP01 Populus nigra mRNA for peroxidase, complete cds. 468 e-131
45	gb M37156 HRAHRPCA A.rusticana peroxidase isoenzyme C (HPR C) ge 363 e-130 emb X97348 PTPXP1PER P.trichocarpa mRNA for anionic peroxidase P 463 e-129
7.5	dbj[D30652[POPPA Populus kitakamiensis mRNA for peroxidase, part 441 e-123
	emb[X97351]PTPXP4PER P.trichocarpa mRNA for anionic peroxidase P 383 e-120
	dbj D30653 POPPB Populus kitakamiensis mRNA for peroxidase, part 356 e-112
	emb AF149277 AF149277 Phaseolus vulgaris peroxidase 1 precursor 227 e-107
50	gb L36157 ALFPXDC Medicago sativa peroxidase (pxdC) mRNA, comple 229 e-107
	emb AF007211 AF007211 Glycine max peroxidase precursor (GMIPER1) 229 e-106
	gb L07554 LINPEROX Linum usitatissimum peroxidase (FLXPER1) mRNA 381 e-10
	emb X90692 MSRNAPE1A M.sativa mRNA for peroxidase 1A. 211 e-105
_	dbj D90115 HRAPRXC2 Horseradish prxC2 gene encoding peroxidase i 296 e-103
55 -	emb AJ242742 IBA242742 Ipomoea batatas mRNA for peroxidase (pod 138 2e-97
	emb AF149280 AF149280 Phaseolus vulgaris peroxidase 5 precursor 142 4e-97
	emb[X90693]MSRNAPE1B M.sativa mRNA for peroxidase 1B. 231 1e-95
	emb X90694 MSRNAPE1C M.sativa mRNA for peroxidase 1C. 226 7e-94
<b>5</b> 0	gb L36111 SSNPEROXIB Stylosanthes humilis peroxidase mRNA. 210 2e-89
50	emb AI959837 AI959837 sc94h07.y1 Gm-c1019 Glycine max cDNA clone 228 6e-89
	emb AF244924 AF244924 Spinacia oleracea peroxidase prx15 precurs 208 9e-88

emb|AW559660|AW559660 EST314772 DSIR Medicago truncatula cDNA cl... 236 1e-87 emb|AW774581|AW774581 EST333732 KV3 Medicago truncatula cDNA clo... 236 7e-87 emb|AF155124|AF155124 Gossypium hirsutum bacterial-induced perox... 215 2e-86 emb|AW981426|AW981426 EST392579 DSIL Medicago truncatula cDNA cl... 236 2e-85 5 emb|AB042103|AB042103 Asparagus officinalis AspPOX1 mRNA for per... 209 2e-85 emb|AF244923|AF244923 Spinacia oleracea peroxidase prx14 precurs... 205 9e-85 emb|AJ250121|PAB250121 Picea abies mRNA for SPI2 protein (spi2 g... 253 1e-83 emb|AW775762|AW775762 EST334827 DSIL Medicago truncatula cDNA cl... 236 6e-83 emb[Y10466]SOPRXR5 S.oleracea mRNA for peroxidase, clone PC18. 10 emb|AW775425|AW775425 EST334490 DSIL Medicago truncatula cDNA cl... 208 1e-81 emb|AW685437|AW685437 NF029D09NR1F1000 Nodulated root Medicago t... 230 9e-80 emb|AB024439|AB024439 Scutellaria baicalensis mRNA for peroxidas... 209 2e-79 emb|AF244922|AF244922 Spinacia oleracea peroxidase prx13 precurs... 212 3e-79 emb|X57564|ARNEUPERO A.rusticana mRNA for neutral peroxidase. 115 6e-78 emb|AW776273|AW776273 EST335338 DSIL Medicago truncatula cDNA cl... 222 1e-77 15 emb|AW267813|AW267813 EST305941 DSIR Medicago truncatula cDNA cl... 200 1e-76 emb|X91232|MARNAPRX M.annua mRNA for peroxidase. 196 5e-76 emb|AF049881|AF049881 Linum usitatissimum peroxidase FLXPER4 (PE... 207 7e-76 emb|AW256487|AW256487 EST304624 KV2 Medicago truncatula cDNA clo... 200 9e-76 emblAW775890lAW775890 EST334955 DSIL Medicago truncatula cDNA cl... 200 -3e-74 20 emb|AW257195|AW257195 EST305332 KV2 Medicago truncatula cDNA clo... 214 5e-74 gb|M91373|CUSPREPER Cucumis sativus peroxidase mRNA, complete cds. 86 2e-73 gb|U41657|GMU41657 Glycine max seed coat peroxidase isozyme (SPO... 159 3e-73 emb|AB027752|AB027752 Nicotiana tabacum mRNA for peroxidase, com... 197 5e-73 25 gb|L36110|SSNPEROXIA Stylosanthes humilis peroxidase mRNA. 199 5e-73 gb|M37636|ARCPNC1 Arachis hypogaea cationic peroxidase (PNC1) mR... 204 7e-73 emb|AW685235|AW685235 NF027H10NR1F1000 Nodulated root Medicago t... 236 9e-73 emb|AW278775|AW278775 sf97d02.y1 Gm-c1019 Glycine max cDNA clone... 155 1e-72 emb|X71593|LECEVI1A L.esculentum CEVI-1 mRNA. 99 2e-72 30 gb|J02979|TOBPXDLF Nicotiana tabacum lignin-forming peroxidase m... 100 2e-72 emb|AW559945|AW559945 EST314993 DSIR Medicago truncatula cDNA cl... 233 2e-72 emb|AW980744|AW980744 EST391897 GVN Medicago truncatula cDNA clo... 235 5e-70 emb[Y10467|SOPRXR6 S.oleracea mRNA for peroxidase, clone PC23. 105 le-69 dbj|D83225|POPP02 Populus nigra peroxidase gene, complete cds. 196 2e-69 35 gb[M74103]TOBANPER Nicotiana sylvestris anionic peroxidase mRNA,... 205 6e-69 emb|AF043234|AF043234 Striga asiatica ferriprotein porphyrin-con... 211 1e-68 gb|BE034991|BE034991 MM01A12 MM Mesembryanthemum crystallinum cD... 202 6e-68 emb|AW574244|AW574244 EST316835 GVN Medicago truncatula cDNA clo... 202 1e-67 emb|Y10465|SOPRXR4 S.oleracea mRNA for peroxidase, clone PC44. 40 emb|AW686084|AW686084 NF038B07NR1F1000 Nodulated root Medicago t... 236 5e-67 emb|AW561032|AW561032 EST316080 DSIR Medicago truncatula cDNA cl... 203 7e-67 emb[X56011]TAPERO Wheat mRNA for peroxidase. 111 2e-66 emb|AW185769|AW185769 se59d08.y1 Gm-c1019 Glycine max cDNA clone... 219 4e-65 emb[Y17192]CPY17192 Cucurbita pepo mRNA for peroxidase. 96 2e-64 45 gb|BE033422|BE033422 ME01E09 ME Mesembryanthemum crystallinum cD... 184 4e-64 emb|AB024438|AB024438 Scutellaria baicalensis mRNA for peroxidas... 205 7e-64 gb[M91374]CUSPREPERA Cucumis sativus peroxidase mRNA, complete cds. 76 2e-63 gb|L24120|LINFLXP Linum usitatissimum peroxidase precursor (FLXP... 167 2e-63 gb|M91372|CUSPREPERB Cucumis sativus peroxidase mRNA, complete cds. 134 2e-63 50 emb|AF043235|AF043235 Striga asiatica ferriprotein porphyrin-con... 199 3e-63 emb|AW288002|AW288002 N100846e rootphos(-) Medicago truncatula c... 182 7e-63 emb|AW126121|AW126121 N100318e rootphos(-) Medicago truncatula c... 216 1e-62 gb|L36093|BLYPRX Barley peroxidase mRNA, complete cds. emb|AW687443|AW687443 NF009F07RT1F1062 Developing root Medicago ... 235 1e-61 55 emb|AW687957|AW687957 NF001D11ST1F1000 Developing stem Medicago ... 236 3e-61 emb|AB024437|AB024437 Scutellaria baicalensis mRNA for peroxidas... 163 3e-61 emb|X58396|HVPEROXI Barley mRNA for peroxidase (EC=1.11.1.7). emb|AI496388|AI496388 sb04a11.yl Gm-c1004 Glycine max cDNA clone... 224 6e-61 emb|AW686765|AW686765 NF042E07NR1F1000 Nodulated root Medicago t... 222 8e-61 60 dbj|D38050|POPP1 Aspen prxA3a gene for peroxidase, complete cds. emb|AW704659|AW704659 sk54h10.y1 Gm-c1019 Glycine max cDNA clone... 149 9e-61

emb[Y10464|SOPRXR3 S.oleracea mRNA for peroxidase, clone PC42. emb|AW705730|AW705730 sk51b02.yl Gm-c1019 Glycine max cDNA clone... 149 8e-60 gb|U51191|GMU51191 Glycine max peroxidase precursor (sEPa1) mRNA... 100 9e-60 gb|U12314|CCU12314 Cenchrus ciliaris clone PX7 peroxidase mRNA. ... 105 9e-60 emb|AI938533|AI938533 sb46h09.yl Gm-c1015 Glycine max cDNA clone... 145 1e-59 emb|AW705617|AW705617 sk50d03.yl Gm-c1019 Glycine max cDNA clone... 149 2e-59 emb|AI781859|AI781859 EST262738 tomato susceptible, Cornell Lyco... 102 2e-59 emb|AF149278|AF149278 Phaseolus vulgaris peroxidase 3 precursor ... 90 3e-59 gb|U51192|GMU51192 Glycine max peroxidase precursor (sEPa2) mRNA... 102 4e-59 10 emb|AW441632|AW441632 EST311028 tomato fruit red ripe, TAMU Lyco... 105 7e-59 emb|AF244921|AF244921 Spinacia oleracea peroxidase prx12 precurs... 156 8e-59 gb|M32742|CUSCUPER C.sativus peroxidase (put.) (CuPer2) mRNA, 3'... 107 2e-58 emb|AW705946|AW705946 sk52h07.yl Gm-c1019 Glycine max cDNA clone... 224 6e-58 15 Query= X68592.6\_at 15978\_at /id\_source genbank/description emb|caa48579.1| (x68592) adenosine nucleotide translocator [arabidopsis thaliana] /blast\_score 0 /ec number /family translocase /chip nova /gb link 20 http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|x68592|/ncgi http://www.ncgr.org/cgi-bin/ff?x68592 (1530 letters) 25 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 30 Score E Sequences producing significant alignments: (bits) Value emb|AF006489|AF006489 Gossypium hirsutum adenine nucleotide tran... 325 0.0 emb|X62123|STANTG S.tuberosum ant gene for ADP/ATP translocator. 35 gb|U89839|LEU89839 Lycopersicon esculentum ADP/ATP translocator ... 327 0.0 emb|X57557|STANT1 S.tuberosum PANT1 mRNA for adenine nucleotide ... 635 0.0 emb|AJ003197|LAAJ3197 Lupinus albus mRNA for adenine nucleotide ... 331 e-175 emb|X80023|TTADPATP T.turgidum mRNA for ADP/ATP carrier. emb|X65194|CRANT C.reinhardtii mRNA CRANT for mitochondrial ADP/... 348 e-149 40 emb|AL023634|SPBC530 S.pombe chromosome II cosmid c530. emb|Z49974|SPANC1GN S.pombe ANC1 gene for adenine nucleotide car... 265 e-139 emb|AF085429|AF085429 Candida parapsilosis ADP/ATP carrier prote... 311 e-134 dbj|D89102|D89102 Schizosaccharomyces pombe mRNA, partial cds, c... 265 e-133 gb|L33797|YSKAAC Kluyveromyces lactis ADP/ATP translocase (AAC) ... 259 e-132 45 emb|AF237675|AF237675 Yarrowia lipolytica ADP/ATP carrier protei... 247 e-132 emb|AJ277099|CUT277099 Candida utilis anc gene for mitochondria1... 253 e-131 emb|AJ277098|CUT277098 Candida utilis anc gene for mitochondrial... 253 e-131 gb[M34075]YSCAAC3 S.cerevisiae ADP/ATP-translocator protein (AAC... 252 e-129 emb|X77291|SCIILDNA S.cerevisiae YBL0421, YBL0438, YBL0418, YBL0... 252 e-129 50 emb|Z35791|SCYBL030C S.cerevisiae chromosome II reading frame OR... 252 e-129 emb|X74427|SCADNUCA S.cerevisiae gene for adenine nucleotide car... 252 e-129 gb|J04021|YSCAAC2 S.cerevisiae ADP/ATP carrier protein (AAC2) ge... 252 e-129 emb|X00363|NCADPATP Neurospora crassa mRNA for mitochondrial ADP... 280 e-126 emb|Z49703|SC9796 S.cerevisiae chromosome XIII cosmid 9796. 55 gb M12514 YSCPET9 S.cerevisiae ADP/ATP translocator protein (AAC... 248 e-125 emb|Z35954|SCYBR085W S.cerevisiae chromosome II reading frame OR... 247 e-125 gb|M34076|YSCAAC2A S.cerevisiae ADP/ATP-translocator protein (AA... 247 e-125 emb|AW774326|AW774326 EST333477 KV3 Medicago truncatula cDNA clo... 288 e-124 emb|AL111975|CNS019QN Botrytis cinerea strain T4 cDNA library un... 254 e-122 60 emb|AW647699|AW647699 EST307178 tomato germinating seedlings, TA... 299 e-120

emb|AW349848|AW349848 GM210006A11G9R Gm-r1021 Glycine max cDNA 3... 326 eemb|AW624842|AW624842 EST313671 tomato radicle, 5 d post-imbibit... 229 e-116 emb|X95863|TTANT1 T.turgidum ant gene (1549bp). 195 e-114 5 emb|AW041186|AW041186 EST284050 tomato mixed elicitor, BTI Lycop... 239 e-114 emb|X95864|TTANT2 T.turgidum ant gene (1494bp). 191 e-112 emb|AW706324|AW706324 sj54h05.yl Gm-c1033 Glycine max cDNA clone... 245 e-110 emb|AL157416|LMFL6066 Leishmania major Friedlin chromosome 19 co... 233 e-108 emb|AW160172|AW160172 EST290029 L. pennellii trichome, Cornell U... 328 e-108 10 emb|AW928728|AW928728 EST337516 tomato flower buds 8 mm to pre-a... 247 e-107 emb|AW218544|AW218544 EST303727 tomato radicle, 5 d post-imbibit... 325 e-105 emb|AW830381|AW830381 sm26a12.yl Gm-c1028 Glycine max cDNA clone... 268 e-105 emblAW201674|AW201674 sf05h11.yl Gm-c1027 Glycine max cDNA clone... 309 e-104 emb|AI812944|AI812944 22G12 Pine Lambda Zap Xylem library Pinus ... 251 e-103 15 emb|AI777865|AI777865 EST258744 tomato susceptible, Cornell Lyco... 284 e-102 emb|AW234033|AW234033 sf33d01.yl Gm-c1028 Glycine max cDNA clone... 237 e-101 emb[AW831587]AW831587 sm28b02.y1 Gm-c1028 Glycine max cDNA clone... 300 e-101 emb|AW668198|AW668198 GA Ea0013C13 Gossypium arboreum 7-10 dpa ... 331 e-100 emb|AF049130|AF049130 Trypanosoma brucei brucei ADP/ATP carrier ... 238 1e-98 20 emb|AI898886|AI898886 EST268329 tomato ovary, TAMU Lycopersicon ... 213 2e-98 gblU32987|TBU32987 Trypanosoma brucei rhodesiense ADP/ATP carrie... 238 3e-98 emb|AW223982|AW223982 EST300793 tomato fruit red ripe, TAMU Lyco... 327 1e-96 emb|AW624951|AW624951 EST313780 tomato radicle, 5 d post-imbibit... 327 1e-96 emb|AW348348|AW348348 GM210002A13A5R Gm-r1021 Glycine max cDNA 3... 328 2e-25 emblAW441243|AW441243 EST310639 tomato fruit red ripe, TAMU Lyco... 324 1e-95 emb|AW223973|AW223973 EST300784 tomato fruit red ripe, TAMU Lyco... 321 7e-95 emb|AW931569|AW931569 EST357412 tomato fruit mature green, TAMU ... 328 2e-94 emb|AW218871|AW218871 EST301353 tomato root during/after fruit s... 327 8e-94 30 emb|AL116444|CNS01D6S Botrytis cinerea strain T4 cDNA library un... 267 4e-93 emb|AW831561|AW831561 sm34f06.y1 Gm-c1028 Glycine max cDNA clone... 312 2e-92 emb|AI484151|AI484151 EST248958 tomato resistant, Cornell Lycope... 264 3e-91 emb|AW925414|AW925414 HVSMEg0001L02 Hordeum vulgare pre-anthesis... 299 6e-91 emb|AI731594|AI731594 BNLGHi10182 Six-day Cotton fiber Gossypium... 335 7e-91 35 emb|AW647665|AW647665 EST307143 tomato germinating seedlings, TA... 317 1e-90 emb|AW395111|AW395111 sh40a06.yl Gm-c1017 Glycine max cDNA clone... 303 6e-90 emb|AI725588|AI725588 BNLGHi12376 Six-day Cotton fiber Gossypium... 288 1e-89 emb|AW934656|AW934656 EST353548 tomato flower buds 0-3 mm, Corne... 328 2e-89 40 emb|AW757478|AW757478 874001D11.yl C. reinhardtii CC-1690, Lambd... 202 3e-89 emb|AW725897|AW725897 GA\_Ea0020B06 Gossypium arboreum 7-10 dpa ... 328 8e-89 emb|AW647757|AW647757 EST326211 tomato germinating seedlings, TA... 301 1e-88 emb|AW509174|AW509174 sh92b04.y1 Gm-c1016 Glycine max cDNA clone... 297 4e-88 emblAL114553|CNS01BO9 Botrytis cinerea strain T4 cDNA library un... 231 3e-87 45 gb|BE021489|BE021489 sm59b05.y1 Gm-c1028 Glycine max cDNA clone ... 322 5e-87 emb|AW507801|AW507801 si45c02.yl Gm-r1030 Glycine max cDNA clone... 259 2e-86 emb|AW733916|AW733916 sk85a11.yl Gm-c1035 Glycine max cDNA clone... 306 2e-86 emblAW096468|AW096468 EST289648 tomato mixed elicitor, BTI Lycop... 292 6e-86 50 emb|AW156741|AW156741 se30b08.yl Gm-c1015 Glycine max cDNA clone... 184 1e-85 emb|AI729625|AI729625 BNLGHi13824 Six-day Cotton fiber Gossypium... 276 4e-85 emb|AW034214|AW034214 EST277785 tomato callus, TAMU Lycopersicon... 304 5e-85 emblAJ273864|AJ273864 AJ273864 Metarhizium anisopliae ARSEF 2575... 184 1e-84 emb|AI775647|AI775647 EST256747 tomato resistant, Cornell Lycope... 297 1e-84 emb|AI822682|AI822682 L0-1204T3 Ice plant Lambda Uni-Zap XR expr... 314 2e-84 emb|AW625107|AW625107 EST313924 tomato radicle, 5 d post-imbibit... 314 2e-84 emb|AI726147|AI726147 BNLGHi5085 Six-day Cotton fiber Gossypium ... 263 6e-83 emb|AW755396|AW755396 sl03d11.yl Gm-c1036 Glycine max cDNA clone... 306 3e-82 emb|AI728088|AI728088 BNLGHi9938 Six-day Cotton fiber Gossypium ... 259 5e-82 60 emb|AW979992|AW979992 EST310378 tomato root deficiency, Cornell ... 285 8e-82 emb|AW333018|AW333018 S16C3 AGS-1 Pneumocystis carinii f. sp. ca... 278 1e-81

emblAI780394|AI780394 EST261273 tomato susceptible, Cornell Lyco... 284 2e-81 emb|AW691046|AW691046 NF037A12ST1F1000 Developing stem Medicago ... 266 2e-81 emb|AW924717|AW924717 WS1 71 A04.b1 A002 Water-stressed I (WS1) ... 303 3e-81 emb|AI731287|AI731287 BNLGHi9093 Six-day Cotton fiber Gossypium ... 262 5e-81 5 emb|AW760027|AW760027 sl57b04.yl Gm-c1027 Glycine max cDNA clone... 301 1e-80 emb|AW622023|AW622023 EST312821 tomato root during/after fruit s... 280 4e-80 emblAV388663|AV388663 AV388663 Chlamydomonas reinhardtii C9 Chla... 284 8e-80 emb|AW650428|AW650428 EST328882 tomato germinating seedlings, TA... 278 4e-79 10 emb|AI775628|AI775628 EST256728 tomato resistant, Cornell Lycope... 295 7e-79 gb|BE024093|BE024093 sm96f04.yl Gm-c1015 Glycine max cDNA clone ... 293 2e-78 emb|AJ273749|AJ273749 AJ273749 Metarhizium anisopliae ARSEF 2575... 256 1e-77 emb|AW737446|AW737446 EST338789 tomato flower buds, anthesis, Co... 290 2e-77

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Query= Y14251.4 i at 16053 i at /id source genbank /description emb|caa74639.1| (y14251) glutathione s-transferase [arabidopsis thaliana] /blast\_score 1.00e-110 /ec\_number /family transferase /chip nova/gb link

http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb[y14251]/ncgi http://www.ncgr.org/cgi-bin/ff?y14251 (630 letters)

25 Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching.....done

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E Score

Sequences producing significant alignments:

(bits) Value

emb|X78203|HMGST H.muticus mRNA for glutathione S-transferase. emb|AW727692|AW727692 GA\_Ea0015I24 Gossypium arboreum 7-10 dpa ... 146 9e-71 emb|AF002692|AF002692 Solanum commersonii glutathione S-transfer... 264 3e-70 dbj|D10524|TOBPARB Nicotiana tabacum mRNA for glutathione S-tran... 262 2e-69 dbj|D29680|TOBAPI2B Tobacco api2 mRNA (which expression is induc... 261 4e-69 emblAW220064|AW220064 EST302547 tomato root during/after fruit s... 260 5e-69 emb|Z71749|NPGSTMR N.plumbaginifolia mRNA for glutathione S-tran... 260 8e-69 emb|AI774583|AI774583 EST255683 tomato resistant, Cornell Lycope... 258 3e-68 emb|AW728413|AW728413 GA\_Ea0016J18 Gossypium arboreum 7-10 dpa ... 142 7e-68 emb|AI725552|AI725552 BNLGHi12077 Six-day Cotton fiber Gossypium... 142 4e-67 emblAI728937|AI728937 BNLGHi12090 Six-day Cotton fiber Gossypium... 142 4e-67 gb|BE033971|BE033971 MG02G09 MG Mesembryanthemum crystallinum cD... 228 2e-65 emb|AW735791|AW735791 EST336559 tomato flower buds 0-3 mm, Corne... 248 2e-65 emb|AF242309|AF242309 Euphorbia esula glutathione S-transferase ... 130 3e-62 emb|AI726215|AI726215 BNLGHi5300 Six-day Cotton fiber Gossypium ... 142 5e-61 emb|AW218151|AW218151 EST303332 tomato radicle, 5 d post-imbibit... 232 1e-60 gb|M84968|SIPGTSTF Silene cucubalus glutathione-S-transferase mR... 115 5e-59 emb|AF133894|AF133894 Persea americana glutathione S-transferase... 220 8e-57 emb|AI352770|AI352770 MB58-4B PZ204.BNlib Brassica napus cDNA cl... 174 7e-55 emb|AW040790|AW040790 EST283654 tomato mixed elicitor, BTI Lycop... 210 6e-54 emb|AW649890|AW649890 EST328344 tomato germinating seedlings, TA... 202 2e-51 emb|AW862639|AW862639 00097 leafy spurge Lambda HybriZAP 2.1 two... 128 2e-48 emb|AI943409|AI943409 MF02B7 MF Mesembryanthemum crystallinum cD... 190 1e-47 emb|AV411409|AV411409 AV411409 Lotus japonicus young plants (two... 187 6e-47

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gb|BE033640|BE033640 MF05D04 MF Mesembryanthemum crystallinum cD... 185 2e-46 emb|AW777169|AW777169 Str3-D4 Sugar Beet germination cDNA librar... 185 3e-46 emb|AV414359|AV414359 AV414359 Lotus japonicus young plants (two... 182 2e-45 5 gb|BE034473|BE034473 MH05E04 MH Mesembryanthemum crystallinum cD... 181 4e-45 gb|BE033466|BE033466 ME02E04 ME Mesembryanthemum crystallimum cD... 170 7e-45 emb|AW623451|AW623451 EST321396 tomato flower buds 3-8 mm, Corne... 179 2e-44 emb|AV408851|AV408851 AV408851 Lotus japonicus young plants (two... 173 9e-43 emb|AV417287|AV417287 AV417287 Lotus japonicus young plants (two... 173 9e-43 10 emb|AI486597|AI486597 EST244918 tomato ovary, TAMU Lycopersicon ... 173 1e-42 emb|AV417605|AV417605 AV417605 Lotus japonicus young plants (two... 172 2e-42 emb|AW667380|AW667380 GA\_Ea0009C08 Gossypium arboreum 7-10 dpa ... 147 2e-42 gb|C96149|C96149 C96149 Marchantia polymorpha immature sex organ... 145 1e-41 emb|AI054936|AI054936 coau0002J02 Cotton Boll Abscission Zone cD... 142 2e-40 15 dbi|D49526|TOBPARBA Tobacco chimeric parB promoter/beta-glucuron... 163 1e-39 emb|AI774099|AI774099 EST255199 tomato resistant, Cornell Lycope... 163 1e-39 emblAV418074|AV418074 AV418074 Lotus japonicus young plants (two... 160 6e-39 gb|BE020725|BE020725 sm52b05.yl Gm-c1028 Glycine max cDNA clone ... 148 5e-38 emb|AI162894|AI162894 A026P40U Hybrid aspen plasmid library Popu... 155 4e-37 20 emb|Y07721|PHGLSTRAN P.hybrida mRNA for glutathione S-transferase. 151 5e-36 gb|BE053268|BE053268 GA\_Ea0035A09f Gossypium arboreum 7-10 dpa ... 137 7e-36 gb|M84969|SIPGTSTFA Silene cucubalus glutathione-S-transferase g... 78 1e-35 emb|AW507753|AW507753 si44f10.yl Gm-r1030 Glycine max cDNA clone... 149 2e-35 emb|AW596451|AW596451 sj12e11.yl Gm-c1032 Glycine max cDNA clone... 149 2e-35 25 emblAW926756|AW926756 HVSMEg0008B23 Hordeum vulgare pre-anthesis... 92 3e-35 emb|AJ279691|BPE279691 Betula pendula partial mRNA for glutathio... 144 8e-34 emb|AV427165|AV427165 AV427165 Lotus japonicus young plants (two... 142 3e-33 emb|AJ010451|AMY010451 Alopecurus myosuroides mRNA for glutathio... 97 4e-33 emb|AF184059|AF184059 Triticum aestivum glutathione S-transferas... 81 5e-33 30 emb|AW164336|AW164336 se71b09.yl Gm-c1023 Glycine max cDNA clone... 139 1e-32 emb|AJ010454|AMY010454 Alopecurus myosuroides mRNA for glutathio... 95 2e-32 emb|AJ010452|AMY010452 Alopecurus myosuroides mRNA for glutathio... 94 3e-32 emb|AW678048|AW678048 WS1 13 A09.b1 A002 Water-stressed 1 (WS1) ... 84 3e-32 gb|BE021219|BE021219 sm56c09.yl Gm-c1028 Glycine max cDNA clone ... 99 7e-32 35 emb|AW561921|AW561921 IPPGHZ0010 Cotton fiber and embryo Lambda ... 137 7e-32 emb|AW458936|AW458936 sh16g07.y1 Gm-c1016 Glycine max cDNA clone... 137 7e-32 emb|AW218152|AW218152 EST303333 tomato radicle, 5 d post-imbibit... 137 le-31 emb|AV426478|AV426478 AV426478 Lotus japonicus young plants (two... 137 1e-31 emb|AI823144|AI823144 L30-1027T3 Ice plant Lambda Uni-Zap XR exp... 112 4e-31 40 emb|AI823131|AI823131 L30-1014T3 Ice plant Lambda Uni-Zap XR exp... 112 4e-31 gb|BE058603|BE058603 sn18c06.yl Gm-c1016 Glycine max cDNA clone ... 134 5e-31 emb|AI773198|AI773198 EST254298 tomato resistant, Cornell Lycope... 133 1e-30 emb|AI778224|AI778224 EST259103 tomato susceptible, Cornell Lyco... 111 2e-30 emb|AW745644|AW745644 WS1\_36\_D11.b1\_A002 Water-stressed 1 (WS1) ... 84 4e-30 45 emb|AW279568|AW279568 sf95d06.yl Gm-c1019 Glycine max cDNA clone... 131 4e-30 emb|AW054044|AW054044 L30-2014T3 Ice plant Lambda Uni-Zap XR exp... 106 3e-29 emb|AW202090|AW202090 sf11a03.y1 Gm-c1027 Glycine max cDNA clone... 106 1e-28 emb|AI440830|AI440830 sa86f05.yl Gm-c1004 Glycine max cDNA clone... 126 1e-28 emblAW678628|AW678628 WS1 1 B05.b1 A002 Water-stressed 1 (WS1) S... 85 2e-28 50 emb|AW677650|AW677650 WS1\_10\_F04.b1\_A002 Water-stressed 1 (WS1) ... 80 6e-28 emb|AW678706|AW678706 WS1\_1\_B05.b2\_A002 Water-stressed 1 (WS1) S... 85 8e-28 emb|AW677798|AW677798 WS1\_11\_H05.b1\_A002 Water-stressed 1 (WS1) ... 76 2e-27 emb|AW053375|AW053375 L30-1572T3 Ice plant Lambda Uni-Zap XR exp... 99 2e-27 emb|AV426523|AV426523 AV426523 Lotus japonicus young plants (two... 122 2e-27 55 emb|AV427075|AV427075 AV427075 Lotus japonicus young plants (two... 122 2e-27 emb|AV415440|AV415440 AV415440 Lotus japonicus young plants (two... 122 2e-27 emb|AT000441|AT000441 AT000441 Brassica rapa guard cell Brassica... 122 3e-27 emb|AI442436|AI442436 sa26h04.yl Gm-c1004 Glycine max cDNA clone... 111 4e-27 gb|BE034032|BE034032 MG03G03 MG Mesembryanthemum crystallinum cD... 90 6e-26 60 emb|AW689646|AW689646 NF022G04ST1F1000 Developing stem Medicago ... 66 3e-25 gb|BE060798|BE060798 HVSMEg0013G13f Hordeum vulgare pre-anthesis... 92 6e-25

gb|BE060847|BE060847 HVSMEg0013J11f Hordeum vulgare pre-anthesis... 64 3e-24 emb|AW171715|AW171715 N100609e rootphos(-) Medicago truncatula c... 112 3e-24 emb|AW348131|AW348131 GM210001A21A6R Gm-r1021 Glycine max cDNA 3... 98 4e-24 emb|AW680604|AW680604 WS1\_6\_C03.b1\_A002 Water-stressed 1 (WS1) S... 84 4e-24 emb|AI352728|AI352728 MB47-17 PZ204.BNlib Brassica napus cDNA cl... 110 1e-23 emb|AW127163|AW127163 M110099 GVN Medicago truncatula cDNA clone... 110 1e-23 emb|AW924273|AW924273 WS1\_51\_A04.b1\_A002 Water-stressed 1 (WS1) ... 95 1e-23 emb|AW680779|AW680779 WS1\_7\_D01.b1\_A002 Water-stressed 1 (WS1) S... 97 1e-23 emb|AI444064|AI444064 sa31g12.y1 Gm-c1004 Glycine max cDNA clone... 109 2e-23 emb|AW459151|AW459151 sb21c07.y1 Gm-c1016 Glycine max cDNA clone... 66 6e-23

Query= X77500.2\_at 16522\_at /id\_source genbank /description emb|caa54631.1| (x77500) amino acid transporter [arabidopsis thaliana] /blast\_score 0 /ec\_number /family /chip nova /gb\_link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|x77500| /ncgi http://www.ncgr.org/cgi-bin/ff?x77500 (1619 letters)

20

Datābāse: plantfungal 661,018 sequences; 426,114,510 total letters

Searching done

25

Score E

Sequences producing significant alignments:

(bits) Value

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(735 letters)

5

10

Database: plantfungal

661,018 sequences; 426,114,510 total letters

Searching.....done

Sequences producing significant alignments:

20 Score E

(bits) Value

emb|AW685448|AW685448 NF029F08NR1F1000 Nodulated root Medicago t... 409 e-114 emb|AJ010501|CAR010501 Cicer arietinum L. mRNA for thaumatin-lik... 409 e-114 emb|AW573922|AW573922 EST316513 GVN Medicago truncatula cDNA clo... 409 e-113 emb|AW685184|AW685184 NF026H08NR1F1000 Nodulated root Medicago t... 338 e-105 emb|AW348587|AW348587 GM210002B22C8R Gm-r1021 Glycine max cDNA 3... 365 e-101 emb|AW685583|AW685583 NF029C04NR1F1000 Nodulated root Medicago t... 320 e-100 emb|X15224|NTE22TLP Tobacco E22 gene for a thaumatin-like protein. 201 4e-99 emb|X12739|NTPRRMAJ N. tabacum mRNA for pathogenesis-related pro... 201 4e-99 emb|AF003007|AF003007 Vitis vinifera thaumatin-like protein VVTL... 214 7e-98

emb|AF003007|AF003007 Vitis vinifera thaumatin-like protein VVTL... 214 7e-98
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emb|X03913|NTTHAUR Tobacco mRNA for TMV induced protein homologo... 195 2e-97
emb|AW684755|AW684755 NF021G01NR1F1000 Nodulated root Medicago t... 348 3e-95

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 emb|AW223507|AW223507 EST300318 tomato fruit red ripe, TAMU Lyco...
 193 1e-85

emb|AW222204|AW222204 EST299015 tomato fruit red ripe, TAMU Lyco... 193 1e-85 emb|AW217005|AW217005 EST295719 tomato callus, TAMU Lycopersicon... 193 1e-85 emb|AW622159|AW622159 EST312957 tomato root during/after fruit s... 193 1e-85 emb|AW684839|AW684839 NF022B04NR1F1000 Nodulated root Medicago t... 316 1e-85 5 emb|AI855542|AI855542 sc20e11.yl Gm-c1013 Glycine max cDNA clone... 185 1e-85 emb|X67244|CSOSLP S.commersonii mRNA for osmotin-like protein. 193 2e-85 emb|AW216590|AW216590 EST295304 tomato callus, TAMU Lycopersicon... 193 3e-85 emb|X72926|SCA81OLP S.commersonii (pA81) mRNA for osmotin-like p... 189 6e-85 emb|AF001528|AF001528 Musa acuminata ripening-associated protein... 307 8e-85 10 emb|AW622107|AW622107 EST312905 tomato root during/after fruit s... 192 9e-85 emb|AW218972|AW218972 EST301454 tomato root during/after fruit s... 191 1e-84 emb|AW033829|AW033829 EST277400 tomato callus, TAMU Lycopersicon... 193 2e-84 emb|AW221921|AW221921 EST298732 tomato fruit red ripe, TAMU Lyco... 187 4e-84 emb|AF109653|AF109653 AF109653 Capsicum annuum root susceptible ... 192 9e-84 15 emb|AW621924|AW621924 EST312722 tomato root during/after fruit s... 192 2e-83 emb|AW650675|AW650675 EST329129 tomato germinating seedlings, TA... 193 1e-82 emb|AW034088|AW034088 EST277583 tomato callus, TAMU Lycopersicon... 193 1e-81 emb|AW219330|AW219330 EST301812 tomato root during/after fruit s... 193 1e-81 emb|AW034433|AW034433 EST278004 tomato callus, TAMU Lycopersicon... 175 2e-80 emb|AI895910|AI895910 EST265353 tomato callus, TAMU Lycopersicon... 177 3e-80 emb|AW625009|AW625009 EST313838 tomato radicle, 5 d post-imbibit... 193 4e-80 emb|AI896330|AI896330 EST265773 tomato callus, TAMU Lycopersicon... 193 4e-80 emb|AJ277064|LES277064 Lycopersicon esculentum PR-5 gene for pat... 192 3e-79 gb[M29279]TOBOSM N.tabacum osmotin mRNA, complete cds. 242 4e-79 25 emblAW029746|AW029746 EST273001 tomato callus, TAMU Lycopersicon... 168 2e-78 emb|AW029798|AW029798 EST273053 tomato callus, TAMU Lycopersicon... 168 2e-78 emb|AW650717|AW650717 EST329171 tomato germinating seedlings, TA... 165 2e-77 emb|AW094053|AW094053 EST287233 tomato mixed elicitor, BTI Lycop... 166 3e-77 emb|X61679|NTOSMOTIN N.tabacum mRNA for osmotin. 193 8e-77 30 emb|AW686653|AW686653 NF043G01NR1F1000 Nodulated root Medicago t... 265 1e-76 emb|AW220061|AW220061 EST302544 tomato root during/after fruit s... 193 1e-76 emb|AW830631|AW830631 sm04e12.yl Gm-c1027 Glycine max cDNA clone... 230 2e-76 emblAW099440|AW099440 sd40e08.yl Gm-c1016 Glycine max cDNA clone... 227 2e-76 emblAW458142|AW458142 sh78g09.yl Gm-c1016 Glycine max cDNA clone... 185 2e-75 35 emb|AJ131731|PME131731 Pseudotsuga menziesii mRNA for Thaumatin-... 237 3e-74 gb|BE033983|BE033983 MG02H11 MG Mesembryanthemum crystallinum cD... 155 2e-73 gb[BE034303]BE034303 MH03A01 MH Mesembryanthemum crystallinum cD... 139 4e-73 emb|AW686245|AW686245 NF035F09NR1F1000 Nodulated root Medicago t... 274 7e-73 emb|A15671|A15671 proprothaumatine. 116 4e-72 40 gb|J01209|TDATHAU2 T.daniellii preprothaumatin-2 mRNA, complete ... 116 4e-72 emb|A15673|A15673 proprothaumatine and the coding strand of its ... 116 4e-72 emb|A15677|A15677 proprothaumatine and the coding stand of its s... 116 4e-72 emb|AF110151|AF110151 AF110151 Capsicum annuum root 1st-branched... 192 4e-72 emb|A15675|A15675 proprothaumatine and the coding strand of its ... 116 2e-71 45 emb|A15660|A15660 Mature thaumatin. 116 2e-71 emb|A46806|A46806 Sequence 3 from Patent EP0684312. emblAF121776|AF121776 Juniperus ashei allergen Jun a 3 mRNA, com... 144 1e-70 emb|AF016327|AF016327 Hordeum vulgare Barperm1 (perm1) mRNA, par... 221 3e-70 emb|AW031249|AW031249 EST274624 tomato callus, TAMU Lycopersicon... 200 3e-70 50 emb|AI896554|AI896554 EST265997 tomato callus, TAMU Lycopersicon... 140 5e-70 emb|AF108890|AF108890 AF108890 Capsicum annuum root 1st-branched... 177 9e-70 emb|AV423642|AV423642 AV423642 Lotus japonicus young plants (two... 263 1e-69 emb|AI055586|AI055586 coau0004G15 Cotton Boll Abscission Zone cD... 262 2e-69 emb|AW032915|AW032915 EST276474 tomato callus, TAMU Lycopersicon... 200 2e-69 55 emb|A46810|A46810 Sequence 7 from Patent EP0684312. emb|AW441774|AW441774 EST311170 tomato fruit red ripe, TAMU Lyco... 193 1e-68 emb|AW223623|AW223623 EST300434 tomato fruit red ripe, TAMU Lyco... 193 1e-68 emb|AW032317|AW032317 EST275771 tomato callus, TAMU Lycopersicon... 193 1e-68 emb|AW224329|AW224329 EST301140 tomato fruit red ripe, TAMU Lyco... 193 1e-68 60 emb|AI895353|AI895353 EST264796 tomato callus, TAMU Lycopersicon... 193 1e-68 emb|AA824730|AA824730 CT028.SK Tomato Leaf cDNA from cv. VFNT ch... 193 1e-68

Query= U35829.2 s at 16981 s at /id source genbank /description gb|aac49356.1| (u35829) thioredoxin h [arabidopsis thaliana] 5 /blast\_score 4.00e-64 /ec\_number /family /chip nova /gb\_link /ncgi (357 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Score Sequences producing significant alignments: (bits) Value 15 emb[X89759]BOPC17 B.oleracea mRNA for thioredoxin h-like, pollen... 194 2e-49 gblU59379|BNU59379 Brassica napus thioredoxin-h-like-1 (THL-1) m... 194 2e-49 emb|AB010434|AB010434 Brassica rapa PEC-2 mRNA for Thioredoxin, ... 192 9e-49 emb|AW255457|AW255457 ML480 peppermint glandular trichome Mentha... 190 3e-48 20 emb|AW569018|AW569018 si74e02.yl Gm-c1031 Glycine max cDNA clone... 185 1e-46 emb|AI988470|AI988470 sd02f07.yl Gm-c1020 Glycine max cDNA clone... 185 1e-46 emb|Z70677|RCTHIORXN R.communis mRNA for thioredoxin. emb|AI161830|AI161830 A007P52U Hybrid aspen plasmid library Popu... 182 9e-46 gb|BE053835|BE053835 GA\_Ea0009P21f Gossypium arboreum 7-10 dpa ... 182 9e-46 25 emb|AW677726|AW677726 WS1\_10\_F03.g1\_A002 Water-stressed 1 (WS1) ... 181 2e-45 emb|AW924685|AW924685 WS1\_71\_B11.b1\_A002 Water-stressed 1 (WS1) ... 181 2e-45 emb|AW565750|AW565750 LG1\_349\_G02.g1\_A002 Light Grown 1 (LG1) So... 181 2e-45 emb|AW671668|AW671668 LG1\_349\_G02.b1\_A002 Light Grown 1 (LG1) So... 181 2e-45 emb|AW677651|AW677651 WS1 10 F03.b1 A002 Water-stressed 1 (WS1) ... 181 2e-45 30 emb|AW349345|AW349345 GM210007A20B10R Gm-r1021 Glycine max cDNA ... 179 6eemb|AI938238|AI938238 sc41e05.yl Gm-c1014 Glycine max cDNA clone... 179 6e-45 emb|AI461219|AI461219 sa76f11.yl Gm-c1004 Glycine max cDNA clone... 179 6e-45 emb|AJ009762|TAE9762 Triticum aestivum mRNA for thioredoxin H. 35 emb|AW164730|AW164730 se77a02.yl Gm-c1023 Glycine max cDNA clone... 179 9e-45 gb|T24347|T24347 crs1466 lambdaZAPST Ricinus communis cDNA clone... 177 3e-44 dbj|D87984|D87984 Fagopyrum esculentum mRNA for thioredoxin, com... 177 4e-44 emb|AW983331|AW983331 HVSMEg0010D20f Hordeum vulgare pre-anthesi... 177 4e-44 40 emb|AW329521|AW329521 N200767e rootphos(-) Medicago truncatula c... 176 6e-44 gb|BE053246|BE053246 GA \_ Ea0021K08f Gossypium arboreum 7-10 dpa ... 176 8e-44 emb|AW255195|AW255195 ML185 peppermint glandular trichome Mentha... 176 8e-44 emb|X69915|TATHIORDH T.aestivum mRNA for thioredoxine H. 175 1e-43 emb|AW164347|AW164347 se71c11.yl Gm-c1023 Glycine max cDNA clone... 175 1e-43 45 emb|AW781479|AW781479 sl79e04.yl Gm-c1037 Glycine max cDNA clone... 175 1e-43 emb|A48516|A48516 Sequence 4 from Patent WO9603505. 175 le-43 emb|AW705063|AW705063 sk57b09.yl Gm-c1019 Glycine max cDNA clone... 175 1e-43 emb|AW568753|AW568753 si72f07.yl Gm-c1031 Glycine max cDNA clone... 175 1e-43 emb|AI165128|AI165128 A076P57U Hybrid aspen plasmid library Popu... 175 1e-43 50 emb|AJ001903|TDAJ1903 Triticum durum mRNA for thioredoxin H. 175 1e-43 emb|A48520|A48520 Sequence 8 from Patent WO9603505. 175 le-43 emb|A48514|A48514 Sequence 2 from Patent WO9603505. 175 1e-43 emb|A48519|A48519 Sequence 7 from Patent WO9603505. 175 1e-43 emb|AW329764|AW329764 N201033e rootphos(-) Medicago truncatula c... 174 2e-43 55 emb|AW983305|AW983305 HVSMEg0010C12f Hordeum vulgare pre-anthesi... 174 3e-43 emb|AW626018|AW626018 EST319925 tomato radicle, 5 d post-imbibit... 174 3e-43 emb|AW982237|AW982237 HVSMEg0002G18f Hordeum vulgare pre-anthesi... 174 3e-43 emb|X58527|NTTRNA N.tabacum mRNA for thioredoxin. 173 5e-43 gb[U59380]BNU59380 Brassica napus thioredoxin-h-like-2 (THL-2) m... 101 5e-43 60 gb|C95504|C95504 C95504 Citrus unshiu Miyagawa-wase maturation s... 172 7e-43 emb|AW277335|AW277335 sf80b02.yl Gm-c1019 Glycine max cDNA clone... 171 le-42

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http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb|ac002391| /ncgi http://www.ncgr.org/cgi-bin/ff?ac002391 (1896 letters)

5

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching.....done

10

## Score E

Sequences producing significant alignments:

(bits) Value

emb|AF123503|AF123503 Nicotiana tabacum Nt-gh3 deduced protein m... 484 0.0 15 emb|X60033|GMGH3G G.max GH3 gene for auxin-regulated protein. emb|AI489187|AI489187 EST247526 tomato ovary, TAMU Lycopersicon ... 421 e-116 emb|AI489278|AI489278 EST247617 tomato ovary, TAMU Lycopersicon ... 410 e-113 emb|AW223544|AW223544 EST300355 tomato fruit red ripe, TAMU Lyco... 385 e-106 emb|AI729810|AI729810 BNLGHi5260 Six-day Cotton fiber Gossypium ... 225 e-103 emb|AW455302|AW455302 EST311840 tomato root during/after fruit s... 358 6e-98 emb|AW307114|AW307114 sf52h11.y1 Gm-c1009 Glycine max cDNA clone... 357 2e-97 emb|AW730905|AW730905 GA\_ Ea0029G23 Gossypium arboreum 7-10 dpa ... 321 2e-95 emb|AW737090|AW737090 EST338517 tomato flower buds, anthesis, Co... 228 3e-94 gb|BE053589|BE053589 GA Ea0013N22f Gossypium arboreum 7-10 dpa ... 298 1e-79 25 emb[AW164615]AW164615 se74c04.yl Gm-c1023 Glycine max cDNA clone... 169 1e-78 emb|AW686313|AW686313 NF040C09NR1F1000 Nodulated root Medicago t... 211 7e-78 emb|AW035201|AW035201 EST280463 tomato callus, TAMU Lycopersicon... 153 2e-77 emb|AW720296|AW720296 LjNEST20d3r Lotus japonicus nodule library... 290 3e-77 emb|AW559498|AW559498 EST314546 DSIR Medicago truncatula cDNA cl... 169 3e-76 30 emb|AW774169|AW774169 EST333252 KV3 Medicago truncatula cDNA clo... 169 1e-75 emb|AW125947|AW125947 N100142e rootphos(-) Medicago truncatula c... 274 1e-72 emb|AW668277|AW668277 GA\_\_Ea0013G22 Gossypium arboreum 7-10 dpa ... 271 1e-71 emb|AW980820|AW980820 EST391973 GVN Medicago truncatula cDNA clo... 254 2e-66 emb|AW221739|AW221739 EST298550 tomato fruit red ripe, TAMU Lyco... 180 6e-63 35 emb|AW929039|AW929039 EST337743 tomato flower buds 8 mm to pre-a... 144 6e-62 emb|AV410002|AV410002 AV410002 Lotus japonicus young plants (two... 232 9e-61 emb|AW720379|AW720379 LjNEST22a11r Lotus japonicus nodule librar... 175 2e-59 emb|AW928468|AW928468 EST337256 tomato flower buds 8 mm to pre-a... 200 3e-59 emb|AW163964|AW163964 Ljirnpest17-383-g11 Ljirnp Lambda HybriZap... 198 6e-57 40 emb|AW185677|AW185677 se58c02.yl Gm-c1019 Glycine max cDNA clone... 219 5e-56 emb|AI773943|AI773943 EST255043 tomato resistant, Cornell Lycope... 133 5e-54 emb|AI054856|AI054856 coau0002F14 Cotton Boll Abscission Zone cD... 97 8e-53 emb|AV426547|AV426547 AV426547 Lotus japonicus young plants (two... 135 1e-51 emb|AW684512|AW684512 NF017G09NR1F1000 Nodulated root Medicago t... 197 2e-49 45 emb|AW428928|AW428928 Ljirnpest25-068-h9 Ljirnp Lambda HybriZap ... 107 5e-49 emb|AI484627|AI484627 EST242888 tomato ovary, TAMU Lycopersicon ... 194 1e-48 emb|AW676760|AW676760 DG1\_14\_F09.g1\_A002 Dark Grown 1 (DG1) Sorg... 167 2e-47 emb|AW647702|AW647702 EST307181 tomato germinating seedlings, TA... 130 2e-44 emb|AW160191|AW160191 EST290048 L. pennellii trichome, Cornell U... 137 6e-44 50 emb|AI775892|AI775892 EST256992 tomato resistant, Cornell Lycope... 177 2e-43 emb|AW031091|AW031091 EST274398 tomato callus, TAMU Lycopersicon... 104 4e-43 emb|AI437757|AI437757 sa39b12.yl Gm-c1004 Glycine max cDNA clone... 144 6e-43 emblAW647700|AW647700 EST307179 tomato germinating seedlings, TA... 129 7e-43 gb|BE122431|BE122431 Ljimpest26-072-a2 Ljimp Lambda HybriZap t... 107 1e-42 55 emb|AW349113|AW349113 GM210004B21A1R Gm-r1021 Glycine max cDNA 3... 131 4eemb|AI737532|AI737532 T110159e KV0 Medicago truncatula cDNA clon... 165 1e-39 emb|AV420807|AV420807 AV420807 Lotus japonicus young plants (two... 154 7e-39 emb|AW622684|AW622684 EST306821 tomato flower buds 3-8 mm, Corne... 105 7e-38 60 emb|AW217219|AW217219 EST295933 tomato callus, TAMU Lycopersicon... 157 2e-37

emb|AI771570|AI771570 EST252670 tomato ovary, TAMU Lycopersicon ... 112 3e-37

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55

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http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb|ac006585| /ncgi

http://www.ncgr.org/cgi-bin/ff?ac006585 (1582 letters)

Database: plantfungal
5 661,018 sequences; 426,114,510 total letters
Searching......done

Score II
Sequences producing significant alignments:

dbj|D88273|D88273 Hordeum vulgare naat-A mRNA for nicotianamine ... 446 e-124 emb|AB005788|AB005788 Hordeum vulgare mRNA for nicotianamine ami... 433 e-120 emb|AB024006|AB024006 Hordeum vulgare naat-B and naat-A genes fo... 184 3e-78

(bits) Value

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emb|AW832427|AW832427 sm10c03.y1 Gm-c1027 Glycine max cDNA clone... 233 3e-60 emb|AW760284|AW760284 sl48d01.y1 Gm-c1027 Glycine max cDNA clone... 206 6e-55 emb|AW620771|AW620771 sj09d03.y1 Gm-c1032 Glycine max cDNA clone... 209 5e-53 emb|AW568831|AW568831 si61g09.y1 Gm-r1030 Glycine max cDNA clone... 199 5e-50 emb|AW030722|AW030722 EST273977 tomato callus, TAMU Lycopersicon... 198 9e-50 emb|AW306460|AW306460 se51a02.y1 Gm-c1017 Glycine max cDNA clone... 178 8e-44

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 emb|AI487927|AI487927 EST246249 tomato ovary, TAMU Lycopersicon ...
 70 7e-23
 emb|AW737817|AW737817 EST339244 tomato flower buds, anthesis, Co...
 79 2e-22
 emb|AW924630|AW924630 WS1\_70\_A07.gl\_A002 Water-stressed 1 (WS1) ...
 106 5e-22
 emb|AW832131|AW832131 sm20a05.yl Gm-c1027 Glycine max cDNA clone...
 105 9e-22

emb|AW221912|AW221912 EST298723 tomato fruit red ripe, TAMU Lyco... 70 9e-19 emb|AI562691|AI562691 TENS2678 T. cruzi epimastigote normalized ... 92 1e-18 emb|AI562604|AI562604 TENS2761 T. cruzi epimastigote normalized ... 95 2e-18 emb|AW782202|AW782202 sm02g12.yl Gm-c1027 Glycine max cDNA clone... 70 1e-16 emb|Z48758|SC9727 S.cerevisiae chromosome IV cosmid 9727. 54 2e-16 emb|AV421843|AV421843 AV421843 Lotus janonicus young plants (two 86 6e-16

emb|AV421843|AV421843 AV421843 Lotus japonicus young plants (two... 86 6e-16 emb|Z69909|SPAC19G10 S.pombe chromosome I cosmid c19G10. 57 1e-15 emb|AL096788|SPBC582 S.pombe chromosome II cosmid c582. 57 1e-15 emb|AW034806|AW034806 EST278842 tomato callus, TAMU Lycopersicon... 85 1e-15 gb|U53880|YSCL9449 Saccharomyces cerevisiae chromosome XII cosmi... 51 2e-14

emb|Z73261|SCYLR089C S.cerevisiae chromosome XII reading frame O...
 emb|AW100534|AW100534 sd56d09.y1 Gm-c1016 Glycine max cDNA clone...
 emb|AW776160|AW776160 EST335225 DSIL Medicago truncatula cDNA cl...
 emb|Z49335|SCYJL060W S.cerevisiae chromosome X reading frame ORF...
 emb|AW329712|AW329712 N200976e rootphos(-) Medicago truncatula c...
 1e-12

emb|Z98531|SPAC6B12 S.pombe chromosome I cosmid c6B12. 63 2e-12 emb|AI057693|AI057693 TENU1783 T. cruzi epimastigote normalized ... 74 3e-12 emb|AV419799|AV419799 AV419799 Lotus japonicus young plants (two... 72 2e-11 emb|AW219060|AW219060 EST301542 tomato root during/after fruit s... 71 3e-11 emb|AW756163|AW756163 sl16f12.yl Gm-c1036 Glycine max cDNA clone... 62 2e-08

60 emb|AI667953|AI667953 TENG0919 T. Cruzi epimastigote normalised ... 61 2e-08 emb|AQ911130|AQ911130 GSSTc02564 Trypanosome cruzi random genomi... 61 2e-08

emblAQ904747|AQ904747 GSSTc03969 Trypanosome cruzi random genomi... 61 2e-08 emb|AQ905153|AQ905153 GSSTc03744 Trypanosome cruzi random genomi... 61 2e-08 emb|AI068927|AI068927 mgae0004dB02f Magnaporthe grisea Appressor... 45 5e-08 emb|AQ445624|AQ445624 GSSTc00112 Trypanosoma cruzi random genomi... 56 1e-06 5 emb[AII10334]AII10334 TENU3304 T. cruzi epimastigote normalized ... 39 1e-06 emb|AI055475|AI055475 coau0004B14 Cotton Boll Abscission Zone cD... 55 2e-06 gb|U37774|CMU37774 Cucurbita maxima 1-aminocyclopropane-1-carbox... 50 2e-06 dbj|D01033|CUCACCA Cucurbita maxima mRNA for 1-aminocyclopropane... 50 2e-06 emb|AF083816|AF083816 Antirrhinum majus ACC synthase 3 (ACS3) mR... 46 2e-06 10 emb|AI080889|AI080889 TENU3747 T. cruzi epimastigote normalized ... 52 9e-06 emb|AB006805|AB006805 Cucumis sativus CS-ACS3 mRNA for ACC synth... 48 1e-05 gb[U59813|CSU59813 Cucumis sativus 1-aminocyclopropane-1-carboxy... 48 1e-05 emb|AW934295|AW934295 EST360138 tomato fruit mature green, TAMU ... 52 1e-05 emb|AW101681|AW101681 sd68f11.yl Gm-c1008 Glycine max cDNA clone... 51 2e-05 15 emb|AW691076|AW691076 NF041A07ST1F1000 Developing stem Medicago ... 44 2e-05 emb|AW477177|AW477177 ga42h10.yl Moss EST library PPU Physcomitr... 44 2e-05 emb|AV424265|AV424265 AV424265 Lotus japonicus young plants (two... 51 2e-05 emb|AF074929|AF074929 Sinapis arvensis 1-aminocyclopropane-1-car... 46 3e-05 emb|AW033989|AW033989 EST277651 tomato callus, TAMU Lycopersicon... 51 3e-05 20 emb|AF074931|AF074931 Sinapis arvensis 1-aminocyclopropane-1-car... 46 3e-05 emb|AI896625|AI896625 EST266068 tomato callus, TAMU Lycopersicon... 50 5e-05 emb|AW695981|AW695981 NF100F04ST1F1042 Developing stem Medicago ... 50 5e-05 emblAI483496lAI483496 EST249317 tomato ovary, TAMU Lycopersicon ... 41 5e-05 emb|AI487017|AI487017 EST245339 tomato ovary, TAMU Lycopersicon ... 41 5e-05 25 emb|AW032141|AW032141 EST275595 tomato callus, TAMU Lycopersicon... 50 6e-05 emb|AQ906278|AQ906278 GSSTc02322 Trypanosome cruzi random genomi... 50 6e-05 emb|AF074930|AF074930 Sinapis arvensis 1-aminocyclopropane-1-car... 45 9e-05 emb|AI896469|AI896469 EST265900 tomato callus, TAMU Lycopersicon... 49 9e-05 emb|AW033411|AW033411 EST276982 tomato callus, TAMU Lycopersicon... 49 9e-05 30 emb[AW685223]AW685223 NF027D03NR1F1000 Nodulated root Medicago t... 49 9e-05 emb|AI896781|AI896781 EST266224 tomato callus, TAMU Lycopersicon... 49 9e-05 emb|AV426435|AV426435 AV426435 Lotus japonicus young plants (two... 49 1e-04 emb|AW690226|AW690226 NF030F01ST1F1000 Developing stem Medicago ... 40 2e-04 emb|AW093940|AW093940 EST287120 tomato mixed elicitor, BTI Lycop... 48 2e-04 35 emb|Z26322|HVALAAT H.vulgare mRNA for alanine aminotransferase. 48 2e-04 emb|AW727211|AW727211 GA Ea0023N19 Gossypium arboreum 7-10 dpa ... 48 2e-04 emb|AV411813|AV411813 AV411813 Lotus japonicus young plants (two... 48 2e-04 emb[X69421]PMPALAAT2 P. miliaceum mRNA for alanine aminotransfer... 48 2e-04 emb|AQ873648|AQ873648 V73B5 mTn-3xHA/lacZ Insertion Library, str... 48 2e-04 40 emb|AW217028|AW217028 EST295742 tomato callus, TAMU Lycopersicon... 48 2e-04 emb|AW093939|AW093939 EST287119 tomato mixed elicitor, BTI Lycop... 47 3e-04 emb|AZ217827|AZ217827 Sheared DNA-81E6.TF Sheared DNA Trypanosom... 47 3e-04 emb|AF049137|AF049137 Dianthus caryophyllus 1-aminocyclopropane-... 47 4e-04 emb|AI773775|AI773775 EST254875 tomato resistant, Cornell Lycope... 39 4e-04 45 emb|AF052832|AF052832 Trypanosoma cruzi CL Brener cosmid 1b21 ch... 47 4e-04 emb|AF049711|AF049711 Petunia x hybrida pollen-specific 1-aminoc... 42 5e-04 emb|AB010102|AB010102 Malus domestica gene for 1-aminocyclopropa... 41 7e-04 gb[U89156]MDU89156 Malus domestica ACC synthase (MdACS-1) gene. ... 41 7e-04 gb|L31347|MAUACCSYN Malus domestica 1-aminocyclopropane-1-carbox... 41 7e-04 50 emb|AB015624|AB015624 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 41 7e-04 emb|AB007449|AB007449 Actinidia deliciosa mRNA for 1-aminocyclop... 42 7e-04

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Database: plantfungal
60 661,018 sequences; 426,114,510 total letters

Searching.....

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Score E.

Sequences producing significant alignments:

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259 e-117

259 e-117

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	gb[M60464]TOBB13G Tobacco beta-1,3-glucanase mRNA, complete cds. 136 1e-86
40	emb AJ000081 CSAJ81 Citrus sinensis mRNA for beta-1,3-glucanase. 113 2e-86
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	emb X54430 NTSP41A Tobacco sp41a mRNA for (1-3)-beta-glucanase. 135 8e-86
	dbj E03985 E03985 DNA encoding beta-1,3-endoglucanase. 167 3e-84
	dbj E02108 E02108 cDNA sequence coding for beta-1,3-endoglucanase. 167 3e-84
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50	gb M80604 TOMB13GLUA Lycopersicon esculentum beta-1,3-glucanase 142 5e-82
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	emb AW035235 AW035235 EST280497 tomato callus, TAMU Lycopersicon 179 5e-77
<b>C O</b>	gb M60462 TOBPRNA Tobacco PRN protein mRNA, 3' end. 117 5e-76
50	emb AW775271 AW775271 EST334336 DSIL Medicago truncatula cDNA cl 163 6e-76
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emb|AW216637|AW216637 EST295351 tomato callus, TAMU Lycopersicon... 164 2e-75 emb|AW032447|AW032447 EST276006 tomato callus, TAMU Lycopersicon... 227 2e-74 emb|AW156463|AW156463 se26e04.y1 Gm-c1015 Glycine max cDNA clone... 256 3e-74 emb|Z68154|GHBGLUCS G.hirsutum mRNA for 1,3-beta-glucanase. 152 le-73 5 gb|M63634|TOBGCBREG Nicotiana plumbaginifolia beta(1,3)-glucanas... 250 9e-73 emb|AW032644|AW032644 EST276203 tomato callus, TAMU Lycopersicon... 189 1e-71 emb|AF230109|AF230109 Populus alba x Populus tremula beta-1,3 gl... 173 3e-71 gb|M20618|TOBGLUBA N.tabacum beta-1,3-glucanase mRNA, clones pGL... 238 3e-71 emb|AI894445|AI894445 EST263900 tomato callus, TAMU Lycopersicon... 231 3e-71 10 emb|AF034107|AF034107 Glycine max beta-1,3-glucanase 2 (SGlu2) g... 142 7e-71 emb|AW031387|AW031387 EST274841 tomato callus, TAMU Lycopersicon... 237 7e-71 emb|AW035366|AW035366 EST280928 tomato callus, TAMU Lycopersicon... 237 7e-71 emb|AI896035|AI896035 EST265478 tomato callus, TAMU Lycopersicon... 237 7e-71 emb|AW031155|AW031155 EST274589 tomato callus, TAMU Lycopersicon... 237 7e-71 15 emb|AW030447|AW030447 EST273702 tomato callus, TAMU Lycopersicon... 236 3e-70 emb|AF034116|AF034116 Glycine max beta-1,3-glucanase 11 (SGlu11)... 124 9e-70 emb|AW030277|AW030277 EST273532 tomato callus, TAMU Lycopersicon... 144 2e-69 emb|AF034110|AF034110 Glycine max beta-1,3-glucanase 5 (SGlu5) g... 122 3e-69 emb|AW035160|AW035160 EST280422 tomato callus, TAMU Lycopersicon... 142 6e-69 20 emb|AW032444|AW032444 EST276003 tomato callus, TAMU Lycopersicon... 142 6e-69 -emb|X74906|LEQB L.esculentum TomQ'b mRNA for beta(1,3)glucanase. emb|AJ011769|CIN011769 Cichorium intybus X Cichorium endivia sub... 97 1e-68 emb|AW030283|AW030283 EST273538 tomato callus, TAMU Lycopersicon... 140 2e-68 gb[M13237]BLYGLUCB Barley beta glucanase mRNA. 25 emb|Z15131|ASBGLUCAN A.sativa mRNA for beta glucanase. 158 9e-68 emblAI896042|AI896042 EST265485 tomato callus, TAMU Lycopersicon... 237 1e-67 gb|M62740|BLYGLB2 Hordeum vulgare 1,3-1,4-beta-D glucan 4-glucan... 158 2e-67 emb|AW031259|AW031259 EST274634 tomato callus, TAMU Lycopersicon... 136 3e-67 emb|AW035840|AW035840 EST281994 tomato callus, TAMU Lycopersicon... 136 3e-67 30 emb|AW032451|AW032451 EST276010 tomato callus, TAMU Lycopersicon... 136 3e-67 emb|AW031353|AW031353 EST274807 tomato callus, TAMU Lycopersicon... 136 3e-67 emb|AI895981|AI895981 EST265424 tomato callus, TAMU Lycopersicon... 136 3e-67 gb[U73709[VVU73709 Vitis vinifera beta-1,3-glucanase mRNA, parti... 244 3e-67 emb|Z22874|TABETGLUB T.aestivum (1,3;1,4) beta glucanase mRNA, c... 160 3e-67 35 emb|AF034114|AF034114 Glycine max beta-1,3-glucanase 9 (SGlu9) p... 119 7e-67 Query= U18993.2 s at 17487 s at /id source genbank /description gb|aac49117.1| (u18993) tryptophan synthase alpha chain [arabidopsis 40 thaliana] /blast\_score 1.00e-158 /ec\_number /family /chip nova /gb link /ncgi (939 letters) Database: plantfungal 45 661,018 sequences; 426,114,510 total letters Searching.....done Score E 50 Sequences producing significant alignments: (bits) Value emb|AW730233|AW730233 GA\_Ea0014B16 Gossypium arboreum 7-10 dpa ... 352 5e-98 emb|AW649960|AW649960 EST328414 tomato germinating seedlings, TA... 308 4e-83 emb|AW621664|AW621664 EST312462 tomato root during/after fruit s... 290 1e-77 55 emb|AW031327|AW031327 EST274781 tomato callus, TAMU Lycopersicon... 282 3e-75 gb[U38804|PPU38804 Porphyra purpurea chloroplast, complete genome. 271 4e-72 emb|AW775853|AW775853 EST334918 DSIL Medicago truncatula cDNA cl... 264 9e-70 emb|Z21642|CHASTRNAA Antithamnion sp. Chloroplast trnK,trnE,trpA... 255 4e-67 gb|BE124387|BE124387 EST393422 GVN Medicago truncatula cDNA clon... 227 9e-59 gb|BE121873|BE121873 894015F07.y1 C. reinhardtii CC-1690, normal... 227 1e-58

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35	http://www3.ncbi.nlm.nih.gov/htbin-	
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•	(441 letters)	
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40	Database: plantfungal	
	661,018 sequences; 426,114,510 total letters	
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Searching......done

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      dbj|D26084|PETZFDB2 Petunia mRNA for zinc-finger DNA binding pro... 70 2e-34
      gb|BE095284|BE095284 00345 leafy spurge Lambda HybriZAP 2.1 two-... 70 2e-34
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      emb|AW781249|AW781249 sk67b08.y1 Gm-c1016 Glycine max cDNA clone... 67 4e-32
      emb|AI988290|AI988290 sc98f10.y1 Gm-c1020 Glycine max cDNA clone... 69 1e-31
      emb|AI487287|AI487287 EST245609 tomato ovary, TAMU Lycopersicon ... 62 3e-30
      emb|AW775559|AW775559 EST334624 DSIL Medicago truncatula cDNA cl... 66 6e-30
      gb|BE123920|BE123920 EST394045 DSIL Medicago truncatula cDNA clo... 66 6e-30
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      dbj|D26085|PETZFDB3 Petunia zinc-finger DNA binding protein gene.
      emb|AW102472|AW102472 sd88f02.y1 Gm-c1009 Glycine max cDNA clone... 70 1e-28
      emb|AI988657|AI988657 sd06b03.yl Gm-c1020 Glycine max cDNA clone... 70 2e-28
      emb|AF053077|AF053077 Nicotiana tabacum osmotic stress-induced z... 66 5e-27
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      emb|AI778714|AI778714 EST259593 tomato susceptible, Cornell Lyco... 62 2e-26
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      emb|AW037956|AW037956 EST279600 tomato mixed elicitor, BTI Lycop... 62 2e-24
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,	gb[U76554]BRU76554 Brassica rapa zinc-finger protein-1 (BR42) mR 63 4e-22
	emb AW625323 AW625323 EST319146 tomato radicle, 5 d post-imbibit 62 4e-22
	emb AW219736 AW219736 EST302218 tomato root during/after fruit s 62 2e-21
	emb AW185023 AW185023 se85f12.yl Gm-c1023 Glycine max cDNA clone 58 5e-2
	emb AW277333 AW277333 sf80all.yl Gm-c1019 Glycine max cDNA clone 69 6e-2
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	emb AW032112 AW032112 EST275566 tomato callus, TAMU Lycopersicon 62 le-20
	emb AI485651 AI485651 EST243972 tomato ovary, TAMU Lycopersicon 62 4e-20
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	emb AW622660 AW622660 EST313460 tomato root during/after fruit s 62 5e-18
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<b>50</b>	emb[X55012]PSUSP43 Pisum sativum mRNA for an unknown seed prote 30 7.6
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Database: plantfungal 661,018 sequences; 426,114,510 total letters

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(bits) Value

Score E

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	emb Y17276 LES17276 Lycopersicon esculentum p69b gene, complete 138 e-130
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25	emb AW287918 AW287918 N100762e rootphos(-) Medicago truncatula c 93 1e-38 emb AW692289 AW692289 NF054B05ST1F1000 Developing stem Medicago 128 2e-38
25	emb AW185724 AW185724 se58g11.y1 Gm-c1019 Glycine max cDNA clone 122 2e-38
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35	Database: plantfungal
	661,018 sequences; 426,114,510 total letters
	Constitution of the Consti
	Searchingdone
40	Saara E
70	Score E Sequences producing significant alignments: (bits) Value
	Sequences producing significant alignments: (bits) Value
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45	emb[Y18932]LES18932 Lycopersicon esculentum p69F gene. 142 e-152
	emb Y17278 LES17278 Lycopersicon esculentum p69d gene, complete 141 e-146
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	emb[X98930]LESBT2 L.esculentum mRNA for serine protease, SBT2. 163 e-142
	emb AJ006786 LES6786 Lycopersicon esculentum p69d gene. 141 e-136
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	emb Y17275 LES17275 Lycopersicon esculentum p69a gene, complete 138 e-126
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dbj|D86598|D86598 Norway spruce mRNA for antifreeze-like protein... 135 6e-77 emb[X85975]AGAG12 A.glutinosa mRNA for subtilisin-like protein. 139 1e-73 gb|BE124129|BE124129 EST394254 DSIL Medicago truncatula cDNA clo... 263 5e-69 emb|AJ006380|LES6380 Lycopersicon esculentum sbt3 gene. 102 3e-68 emb|AJ006376|LES6376 Lycopersicon esculentum mRNA for serine pro... 102 3e-68 emb|AJ006480|LES6480 Lycopersicon esculentum sbt4b gene. 146 2e-66 emb|AJ006481|LES6481 Lycopersicon esculentum sbt4c gene. 138 2e-66 emb|AW267784|AW267784 EST305912 DSIR Medicago truncatula cDNA cl... 128 3e-65 emb|AJ006483|LES6483 Lycopersicon esculentum sbt4e gene. 140 5e-65 emblAJ006377 LES6377 Lycopersicon esculentum mRNA for serine pro... 138 5e-65 10 emb|AW459354|AW459354 sh41a03.yl Gm-c1017 Glycine max cDNA clone... 192 6e-64 emb[AW349252]AW349252 GM210004B21G12R Gm-r1021 Glycine max cDNA ... 144 1eemb|AJ276710|GMA276710 Glycine max mRNA for putative subtilisin ... 124 8e-63 15 emb|AJ012164|CGL012164 Casuarina glauca mRNA for cg12 gene fragm... 114 4e-62 emb|AI730592|AI730592 BNLGHi7352 Six-day Cotton fiber Gossypium ... 216 2e-61 emb|AW728806|AW728806 GA\_Ea0028I22 Gossypium arboreum 7-10 dpa ... 125 5e-60 emb|AI794673|AI794673 sb67d01.yl Gm-c1019 Glycine max cDNA clone... 171 1e-59 emb|AW773858|AW773858 EST332844 KV3 Medicago truncatula cDNA clo... 87 2e-58 20 emb|AW695319|AW695319 NF093H12ST1F1103 Developing stem Medicago ... 105 2e-57 emb|AW299160|AW299160 EST305970 KV2 Medicago truncatula cDNA clo... 135 3e-57 emb|AF201883|AF201883 Gossypioides kirkii subtilisin-like protea... 108 7e-57 emb|AW720622|AW720622 LjNEST22e12rc Lotus japonicus nodule libra... 137 2e-56 emb[AW925917]AW925917 HVSMEg0005N16 Hordeum vulgare pre-anthesis... 157 4e-56 25 emb|AW668484|AW668484 GA\_Ea0014C06 Gossypium arboreum 7-10 dpa ... 188 4e-56 emb|AF139438|AF139438 Gossypium hirsutum (A-subgenome) locus A17... 106 5e-55 emb|AF139437|AF139437 Gossypium herbaceum locus A1751, sequence ... 106 5e-55 emb|AF139441|AF139441 Gossypium robinsonii locus A1751, sequence... 106 7e-55 emb|AF139440|AF139440 Gossypium hirsutum (D-subgenome) locus A17... 106 7e-55 30 emb|AF139439|AF139439 Gossypium raimondii locus A1751, sequence ... 106 7e-55 emb|AW687885|AW687885 NF014E10RT1F1082 Developing root Medicago ... 91 5e-54 emb|AW563397|AW563397 LG1 214 C02.b1 A002 Light Grown 1 (LG1) So... 210 4e-53 emb|AW730974|AW730974 GA Ea0029F08 Gossypium arboreum 7-10 dpa ... 163 8e-53 emb|AW257362|AW257362 EST305499 KV2 Medicago truncatula cDNA clo... 138 4e-52 35 emb|AW221342|AW221342 EST297811 tomato fruit mature green, TAMU ... 154 7e-52 emb|AW221343|AW221343 EST297812 tomato fruit mature green, TAMU ... 154 7e-52 emb|AI441432|AI441432 sa59h01.yl Gm-c1004 Glycine max cDNA clone... 189 1e-51 emb[AW279412]AW279412 sf79c02.yl Gm-c1019 Glycine max cDNA clone... 204 3e-51 emb|AI730369|AI730369 BNLGHi6751 Six-day Cotton fiber Gossypium ... 166 3e-51 40 emb|AW221789|AW221789 EST298600 tomato fruit red ripe, TAMU Lyco... 95 3e-51 emb|AW030462|AW030462 EST273717 tomato callus, TAMU Lycopersicon... 173 4e-51 emb|AQ917453|AQ917453 T233488b Medicago truncatula BAC library M... 134 5e-51 emb|AW559414|AW559414 EST314462 DSIR Medicago truncatula cDNA cl... 152 1e-50 emb|AW697284|AW697284 NF117B12ST1F1096 Developing stem Medicago ... 128 2e-50 45 emb|AW666485|AW666485 GA\_ Ea0005E24 Gossypium arboreum 7-10 dpa ... 137 3e-50 emb|AQ917126|AQ917126 T233155b Medicago truncatula BAC library M... 148 3e-50 emb|AW780798|AW780798 s176c06.y1 Gm-c1027 Glycine max cDNA clone... 161 1e-49 emb|AW221341|AW221341 EST297810 tomato fruit mature green, TAMU ... 154 2e-49 emblAI727340|AI727340 BNLGHi7810 Six-day Cotton fiber Gossypium ... 194 2e-48 50 emb|AW704701|AW704701 sk39d09.y1 Gm-c1028 Glycine max cDNA clone... 124 2e-47 emb|AW101793|AW101793 sd70e04.y1 Gm-c1008 Glycine max cDNA clone... 190 3e-47 emb|AW730930|AW730930 GA\_Ea0029M05 Gossypium arboreum 7-10 dpa ... 122 6e-47 emb|AW219463|AW219463 EST301861 tomato root during/after fruit s... 141 8e-47 emb|AW688392|AW688392 NF006H03ST1F1000 Developing stem Medicago ... 78 1e-46 gb|BE053648|BE053648 GA\_Ea0001B22f Gossypium arboreum 7-10 dpa ... 81 9e-46 55 emblAW108783|AW108783 gate0001B22f Gossypium arboreum 7-10 dpa f... 81 5e-45 emb|AI727162|AI727162 BNLGHi7467 Six-day Cotton fiber Gossypium ... 167 2e-44 emb|AW725183|AW725183 GA\_Ea0015F02 Gossypium arboreum 7-10 dpa ... 158 2e-44 emb|AJ006482|LES6482 Lycopersicon esculentum sbt4d gene. 60 emb|AA660563|AA660563 00449 MtRHE Medicago truncatula cDNA 5' si... 138 4e-44 emb|AW597797|AW597797 sj86c05.y1 Gm-c1034 Glycine max cDNA clone... 157 2e-43

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

35 Searching.....done

## Score E

Sequences producing significant alignments:

(bits) Value

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25 post/entrez/query?db=n&form=6&dopt=g&uid=gb|y11788|/ncgi http://www.ncgr.org/cgi-bin/ff?y11788 (965 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

35 Sequences producing significant alignments:

(bits) Value

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emb|AV414872|AV414872 AV414872 Lotus japonicus young plants (two... 182 2e-55 dbi|D30653|POPPB Populus kitakamiensis mRNA for peroxidase, part... 94 3e-55 emb|AV414074|AV414074 AV414074 Lotus japonicus young plants (two... 215 4e-55 emb|AW032485|AW032485 EST276044 tomato callus, TAMU Lycopersicon... 212 3e-54 5 emb|AW216562|AW216562 EST295276 tomato callus, TAMU Lycopersicon... 210 2e-53 emb|X57564|ARNEUPERO A.rusticana mRNA for neutral peroxidase. emblAI777064|AI777064 EST252031 tomato callus, TAMU Lycopersicon... 206 2e-52 emb|AF155124|AF155124 Gossypium hirsutum bacterial-induced perox... 93 6e-52 emb|AW219314|AW219314 EST301796 tomato root during/after fruit s... 193 4e-51 10 emb|AB042103|AB042103 Asparagus officinalis AspPOX1 mRNA for per... 84 5e-51 emb|AW219926|AW219926 EST302409 tomato root during/after fruit s... 193 8e-51 emb|AW224630|AW224630 EST303073 tomato root, plants pre-anthesis... 193 8e-51 emb|AF149280|AF149280 Phaseolus vulgaris peroxidase 5 precursor ... 81 8e-51 gb|L13654|TOMTPX1A Lycopersicon esculentum peroxidase (TPX1) mRN... 104 3e-50 15 emb|AJ242742|IBA242742 Ipomoea batatas mRNA for peroxidase (pod ... 82 5e-50 emb|Y10470|SOPRXR9 S.oleracea mRNA for peroxidase, clone PC56. emb|AV422753|AV422753 AV422753 Lotus japonicus young plants (two... 195 4e-49 emb|AI773788|AI773788 EST254888 tomato resistant, Cornell Lycope... 194 7e-49 emb|X90692|MSRNAPE1A M.sativa mRNA for peroxidase 1A. 76 1e-48 20 emb[X56011]TAPERO Wheat mRNA for peroxidase. 102 2e-48 emb|X97351|PTPXP4PER P.trichocarpa mRNA for anionic peroxidase P... 90 8e-48 emb|AW666030|AW666030 sk31c04.yl Gm-c1028 Glycine max cDNA clone... 191 9e-48 gb|M37636|ARCPNC1 Arachis hypogaea cationic peroxidase (PNC1) mR... 89 1e-47 emb|AB024437|AB024437 Scutellaria baicalensis mRNA for peroxidas... 122 1e-47 25 gb|M74103|TOBANPER Nicotiana sylvestris anionic peroxidase mRNA,... 100 2e-47 gb|M60729|HRAHRPCC A.rusticana peroxidase isoenzyme C (HPR C) ge... 78 3e-47 emb|AW621675|AW621675 EST312473 tomato root during/after fruit s... 189 3e-47 emb|Y17192|CPY17192 Cucurbita pepo mRNA for peroxidase. emb|AI895427|AI895427 EST264870 tomato callus, TAMU Lycopersicon... 121 7e-47 30 emb|AW035207|AW035207 EST280469 tomato callus, TAMU Lycopersicon... 188 8e-47 gb|L13653|TOMTPX2A Lycopersicon esculentum peroxidase (TPX2) mRN... 101 1e-46 emb|Y10463|SOPRXR2 S.oleracea mRNA for peroxidase, clone PB22. emb|AW224633|AW224633 EST303076 tomato root, plants pre-anthesis... 185 4e-46 gb|L36157|ALFPXDC Medicago sativa peroxidase (pxdC) mRNA, comple... 73 9e-46 35 gb|M91372|CUSPREPERB Cucumis sativus peroxidase mRNA, complete cds. 89 2e-45 emb|X91232|MARNAPRX M.annua mRNA for peroxidase. emb|AF244924|AF244924 Spinacia oleracea peroxidase prx15 precurs... 92 2e-45 emb|Z22920|SPPEROXDS S.polyrrhiza mRNA for peroxidase. 96 2e-45 emb|X90694|MSRNAPE1C M.sativa mRNA for peroxidase 1C. 75 3e-45 gb|M91374|CUSPREPERA Cucumis sativus peroxidase mRNA, complete cds. 109 4e-45 emb[AW035872]AW035872 EST282181 tomato callus, TAMU Lycopersicon... 182 5e-45 emb|Y16776|SOY16776 Spinacia oleracea mRNA for peroxidase, prx10... 95 5e-45 dbi|D42065|TOBCPI40KB Tobacco mRNA for cationic peroxidase isozy... 77 7e-45 dbj|D42064|TOBCPI38KA Tobacco mRNA for cationic peroxidase isozy... 77 1e-44 45 emb|AI772237|AI772237 EST253337 tomato resistant, Cornell Lycope... 180 2e-44 emb|AF149278|AF149278 Phaseolus vulgaris peroxidase 3 precursor ... 84 2e-44 emb|AW035660|AW035660 EST281492 tomato callus, TAMU Lycopersicon... 178 5e-44 emblAF244922|AF244922 Spinacia oleracea peroxidase prx13 precurs... 83 1e-43 emb|X97350|PTPXP3PER P.trichocarpa mRNA for anionic peroxidase P... 83 2e-43 50 gb|L36111|SSNPEROXIB Stylosanthes humilis peroxidase mRNA. 77 3e-43 emb|X58396|HVPEROXI Barley mRNA for peroxidase (EC=1.11.1.7). 90 3e-43 emb|X97348|PTPXP1PER P.trichocarpa mRNA for anionic peroxidase P... 85 4e-43 gb|L36093|BLYPRX Barley peroxidase mRNA, complete cds. emb|AF043234|AF043234 Striga asiatica ferriprotein porphyrin-con... 83 7e-43 55 emb|X90693|MSRNAPE1B M.sativa mRNA for peroxidase 1B. emb|X97349|PTPXP2PER P.trichocarpa mRNA for anionic peroxidase P... 83 2e-42 gb|L24120|LINFLXP Linum usitatissimum peroxidase precursor (FLXP... 88 2e-42 emb|AW705192|AW705192 sk42h02.y1 Gm-c1019 Glycine max cDNA clone... 111 2e-42 emb|AW705312|AW705312 sk59d04.y1 Gm-c1019 Glycine max cDNA clone... 128 2e-42 60 gb|L07554|LINPEROX Linum usitatissimum peroxidase (FLXPER1) mRNA... 92 3e-42 gb|L77080|SSNCAPEA Stylosanthes humilis cationic peroxidase gene... 122 1e-41

emb|AI778513|AI778513 EST259392 tomato susceptible, Cornell Lyco... 170 1e-41 emb|AB027752|AB027752 Nicotiana tabacum mRNA for peroxidase, com... 87 2e-41 emb|AF007211|AF007211 Glycine max peroxidase precursor (GMIPER1)... 72 2e-41 emb|A00741|A00741 A.rusticana synthetic gene (reverse complement... 76 2e-41 emb|A00740|A00740 A.rusticana synthetic gene for peroxidase. dbj|E01651|E01651 cDNA encoding horseradish peroxidase. 76 2e-41 emb|AF244923|AF244923 Spinacia oleracea peroxidase prx14 precurs... 84 3e-41 emb|AF149277|AF149277 Phaseolus vulgaris peroxidase 1 precursor ... 71 3e-41 dbj|D30652|POPPA Populus kitakamiensis mRNA for peroxidase, part... 80 4e-41 10 emb|Y10469|SOPRXR8 S.oleracea mRNA for peroxidase, clone PC55. 111 4e-41 emb|AJ011939|TRE011939 Trifolium repens mRNA for peroxidase. 105 6e-41 emb|Y10465|SOPRXR4 S.oleracea mRNA for peroxidase, clone PC44. 86 6e-41 gb|U12315|CCU12315 Cenchrus ciliaris clone PX18 peroxidase mRNA,... 106 9e-41 dbj|D11337|VIRPRX Vigna angularis mRNA for peroxidase basic isoz... 77 1e-40 15 gb|M91373|CUSPREPER Cucumis sativus peroxidase mRNA, complete cds. 107 1e-40 Query= AF163823.4\_at 18968\_at /id source genbank /description gb|aad45127.1|af163823\_1 (af163823) endoxyloglucan transferase [arabidopsis thaliana] /blast\_score 1.00e-161 /ec\_number /family 20 /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|af163823|/ncgi http://www.ncgr.org/cgi-bin/ff?af163823 (1025 letters) 25 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching.....done 30 Score, E Sequences producing significant alignments: (bits) Value emb|AJ130885|FSY130885 Fagus sylvatica mRNA for xyloglucan endot... 424 e-134 35 emb|X93174|HVXETXEA H.vulgare mRNA for xyloglucan endotransglyco... 418 e-130 gb|L22162|SOYMER5HOM Glycine max brassinosteroid-regulated prote... 384 e-118 emb|AW220372|AW220372 EST302855 tomato root during/after fruit s... 416 e-115 emb|AW030111|AW030111 EST273366 tomato callus, TAMU Lycopersicon... 416 e-115 emb|AW032346|AW032346 EST275800 tomato callus, TAMU Lycopersicon... 415 e-115 40 emb|AW218856|AW218856 EST301338 tomato root during/after fruit s... 413 e-115 emb|AW031139|AW031139 EST274446 tomato callus, TAMU Lycopersicon... 380 e-115 emb|AI725820|AI725820 BNLGHi13219 Six-day Cotton fiber Gossypium... 413 e-114 emb|AI938507|AI938507 sb46e06.yl Gm-c1015 Glycine max cDNA clone... 407 e-112 emb[AW776635]AW776635 EST335700 DSIL Medicago truncatula cDNA cl... 407 e-112 45 emb|AI488729|AI488729 EST247068 tomato ovary, TAMU Lycopersicon ... 406 e-112 emb|AI485531|AI485531 EST243852 tomato ovary, TAMU Lycopersicon ... 402 e-111 emb|AW033905|AW033905 EST277476 tomato callus, TAMU Lycopersicon... 401 e-111 emb|AW216717|AW216717 EST295431 tomato callus, TAMU Lycopersicon... 397 e-110 gb|BE035552|BE035552 MO09A12 MO Mesembryanthemum crystallinum cD... 353 e-109 50 emb[X93175]HVXETXEB H.vulgare mRNA for xyloglucan endotransglyco... 387 e-109 emb|AW651314|AW651314 EST329768 tomato germinating seedlings, TA... 390 e-108 emb|AW218734|AW218734 EST301214 tomato root during/after fruit s... 388 e-107 emb|AI460643|AI460643 sa71e04.yl Gm-c1004 Glycine max cDNA clone... 387 e-107 emb|AI898515|AI898515 EST267958 tomato ovary, TAMU Lycopersicon ... 386 e-106 emb|AI488903|AI488903 EST247242 tomato ovary, TAMU Lycopersicon ... 386 e-106 emb|AW218894|AW218894 EST301376 tomato root during/after fruit s... 386 e-106 emb|AI487536|AI487536 EST245858 tomato ovary, TAMU Lycopersicon ... 379 e-104 emb|AW981262|AW981262 EST392415 DSIL Medicago truncatula cDNA cl... 315 e-104 60 emb[AW033629[AW033629 EST277200 tomato callus, TAMU Lycopersicon... 378 e-104 emb|AW033699|AW033699 EST277270 tomato callus, TAMU Lycopersicon... 376 e-103

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	dbj E06964 E06964 Anti-sense RNA of xyloglucan endotransferase g 310 4e-93
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	dbj E06972 E06972 DNA encoding xyloglucan endotransferase. 319 6e-89
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20 Database: plantfungal

661,018 sequences; 426,114,510 total letters

Searching.....done

Score E
Sequences producing significant alignments: (bits) Value

emb|AJ130885|FSY130885 Fagus sylvatica mRNA for xyloglucan endot... 424 e-134

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Database: plantfungal

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661,018 sequences; 426,114,510 total letters

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Score E

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(1579 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

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Searching.....

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20	dbj D50617 YSCCHRVIN Saccharomyces cerevisiae chromosome VI comp 30 8.3 -dbj D44598 YSCF4121F Saccharomyces cerevisiae chromosome VI phag 30 8.3 dbj D87895 D87895 Aspergillus nidulans chiA gene for chitinase, 30 8.3 emb AQ162420 AQ162420 mgxb0012N11r CUGI Rice Blast BAC Library P 30 8.3 gb BE036961 BE036961 MP09G07 MP Mesembryanthemum crystallinum cD 30 8.3
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UU	emb AI755342 AI755342 EtESTea36d07.y1 Eimeria S5-2 Sporozoite st 33 7.3
	embla C0055061 A C005506 Plasma dium falsinamm ahramasama 12 alama 22 7 2

emb|AZ217295|AZ217295 Sheared DNA-112B7.TF Sheared DNA Trypanoso... 33 7.3 emb|AW287840|AW287840 N100683e rootphos(-) Medicago truncatula c... 33 7.3

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(1743 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

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Searching.....done

Score E

Sequences producing significant alignments:

(bits) Value

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emb|cab45805.1| (al080253) putative protein [arabidopsis thaliana] /blast\_score 7.00e-83 /ec\_number /family /chip nova /gb\_link http://www3.ncbi.nlm.nih.gov/htbin-5 post/entrez/query?db=n&form=6&dopt=g&uid=gb|al080253| /ncgi http://www.ncgr.org/cgi-bin/ff?al080253 (594 letters) Database: plantfungal 10 661,018 sequences; 426,114,510 total letters Score E 15 Sequences producing significant alignments: (bits) Value emb|Z99969|MAZ99969 Musa acuminata mRNA for putative beta-1,3-gl... 62 3e-09 gb|BE033791|BE033791 MF06E10 MF Mesembryanthemum crystallinum cD... 58 5e-08 20 gb|BE034803|BE034803 ML04E10 ML Mesembryanthemum crystallinum cD... 58 5e-08 emb|AI728744|AI728744 BNLGHi11492 Six-day Cotton fiber Gossypium... 35 2e-07 emb|AW394608|AW394608 sh33e11.y1 Gm-c1017 Glycine max cDNA clone... 41 1e-06 emb|AW687017|AW687017 NF005A10RT1F1072 Developing root Medicago ... 40 2e-06 emb|AW685774|AW685774 NF035A03NR1F1000 Nodulated root Medicago t... 40 2e-06 25 emb|AW220014|AW220014 EST302497 tomato root during/after fruit s... 45 2e-06 emb[AW277478]AW277478 sf82e11.y1 Gm-c1019 Glycine max cDNA clone... 37 8e-06 emb|AW728453|AW728453 GA\_ Ea0016L16 Gossypium arboreum 7-10 dpa ... 34 1e-05 emb|AW625648|AW625648 EST319555 tomato radicle, 5 d post-imbibit... 45 1e-05 emb|AW775954|AW775954 EST335019 DSIL Medicago truncatula cDNA cl... 39 2e-05 30 emb|AW299135|AW299135 EST305809 KV2 Medicago truncatula cDNA clo... 47 1e-04 gb|BE033951|BE033951 MG02E05 MG Mesembryanthemum crystallinum cD... 46 2e-04 emb|AW697566|AW697566 ST64E03 Pine TriplEx shoot tip library Pin... 34 8e-04 emb|AI812453|AI812453 11C3 Pine Lambda Zap Xylem library Pinus t... 44 0.001 emb|AW687794|AW687794 NF013E08RT1F1066 Developing root Medicago ... 32 0.003 35 emb|AW423428|AW423428 sh66f08.y1 Gm-c1015 Glycine max cDNA clone... 38 0.003 emb|AW310121|AW310121 sf31d07.x1 Gm-c1028 Glycine max cDNA clone... emb|AW704218|AW704218 sk17c12.y1 Gm-c1028 Glycine max cDNA clone... emb[AW704640]AW704640 sk54f05.y1 Gm-c1019 Glycine max cDNA clone... 36 0.004 emb|AW395252|AW395252 sh45g06.yl Gm-c1017 Glycine max cDNA clone... 36 0.004 40 emb[AW704612]AW704612 sk54c11.y1 Gm-c1019 Glycine max cDNA clone... 36 0.004 emb|AW284126|AW284126 LG1\_262\_A05.g1\_A002 Light Grown 1 (LG1) So... 41 0.008 emb|AW923658|AW923658 DG1\_57\_B06.g1\_A002 Dark Grown 1 (DG1) Sorg... 39 0.040 emb|AW423359|AW423359 sh06g04.y1 Gm-c1016 Glycine max cDNA clone... 38 0.076 emb|AW649685|AW649685 EST328139 tomato germinating seedlings, TA... 28 0.35 45 emb|AA520781|AA520781 TgESTzz64a03.r1 TgME49 invivo Bradyzoite c... 35 0.37 emb|AW620830|AW620830 sj47b03.y1 Gm-c1033 Glycine max cDNA clone... 30 0.48 emb|X65608|SCPDC2G S.cerevisiae PDC2 gene. 35 0.51 emb|Z74377|SCYDR081C S.cerevisiae chromosome IV reading frame OR... 35 0.51 emb|Z46796|SC8554 S.cerevisiae chromosome IV cosmid 8554. 35 0.51 50 emb|X82086|SCCHROIV S.cerevisiae DNA for right arm of chromosome... 35 0.51 gb|L19880|YSCPDC2A Saccharomyces cerevisiae pdc2 gene, complete ... 35 0.51 emb|AW299176|AW299176 EST305986 KV2 Medicago truncatula cDNA clo... 35 0.70 emb|AW310120|AW310120 sf31d06.x1 Gm-c1028 Glycine max cDNA clone... 35 0.70 emb|AW309936|AW309936 sf26g12.x1 Gm-c1028 Glycine max cDNA clone... 35 0.70 55 emb|AW278505|AW278505 sf45c08.yl Gm-c1009 Glycine max cDNA clone... 35 0.70 emb|AW310598|AW310598 sg21h03.x1 Gm-c1024 Glycine max cDNA clone... 34 0.96 emb|AW233878|AW233878 sf31d07.y1 Gm-c1028 Glycine max cDNA clone... 34 0.96 gb|BE125775|BE125775 DG1\_57\_B06.b1\_A002 Dark Grown 1 (DG1) Sorgh... gb|M13237|BLYGLUCB Barley beta glucanase mRNA. 60 gb[M62740]BLYGLB2 Hordeum vulgare 1,3-1,4-beta-D glucan 4-glucan... 34 0.96 emb|AW747074|AW747074 WS1\_65\_A07.g1\_A002 Water-stressed 1 (WS1) ... 34 1.3

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	emb AL157415 LMFL8325 Leishmania major Friedlin chromosome 5 cos 32 3.4
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	emb Z98549 PFSC03014 Plasmodium falciparum DNA *** SEQUENCING IN 31 6.5
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	emb AC009463 AC009463 Trypanosoma brucei chromosome II clone RPC 31 6.5
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      emb|Z12611|STPROINI S.tuberosum proteinase inhibitor I.
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30
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35
                                           Score
      Sequences producing significant alignments:
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      emb|AL132984|SPAC1556 S.pombe chromosome I cosmid c1556.
      emb|Z26877|SCDCHR11 S.cerevisiae (S288C) 36.2kb DNA fragment fro... 408 0.0
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      gb|M94874|YSCSDH1B Saccharomyces cerevisiae succinate dehydrogen... 408 0.0
      emb|Z28148|SCYKL148C S.cerevisiae chromosome XI reading frame OR... 408 0.0
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      dbj|D89263|D89263 Schizosaccharomyces pombe mRNA, partial cds, c... 637 0.0
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      emb|AW687411|AW687411 NF009C11RT1F1085 Developing root Medicago ... 208 2e-85
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(938 letters)

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Database: plantfungal

661,018 sequences; 426,114,510 total letters

Searching.....done

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Score E

Sequences producing significant alignments:

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Ε Score

15 Sequences producing significant alignments:

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(bits) Value

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Database: plantfungal

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	Sequences producing significant alignments: (bits) Value
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1 -	emb AB004558 AB004558 Lycopersicon esculentum mRNA for acid inve 200 e-164
15	emb X69321 DCBFRUCT D.carota (Queen Anne's Lace) Inv*Dc1 gene. 178 e-145
	emb X78424 DCINC1 D.carota (Queen Anne's Lace) Inv*Dc2 gene, 343 161 e-136
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20	gb U87849 CAU87849 Capsicum annuum acid beta-fructosidase mRNA, 200 e-126
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•	
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	emb X75352 DCRNAABF D.carota (Nantaise) mRNA for soluble acid be 203 e-119
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	AN AND AND AND AND AND AND AND AND AND A
15	
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	omori i i i z-i cui i i i cui i cui i cui i cui cui cu
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	emb Z69369 SPAC3F10 S.pombe chromosome I cosmid c3F10. 314 e-119
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emblAW759534|AW759534 sl44f02.yl Gm-c1027 Glycine max cDNA clone... 166 9e-40 emb|AW223508|AW223508 EST300319 tomato fruit red ripe, TAMU Lyco... 165 2e-39 emb|AW219577|AW219577 EST302059 tomato root during/after fruit s... 164 5e-39 emb|AW775340|AW775340 EST334405 DSIL Medicago truncatula cDNA cl... 161 3e-38 emb|Z28328|SCYKR103W S.cerevisiae chromosome XI reading frame OR... 122 6e-38 emb|AW039256|AW039256 EST281513 tomato mixed elicitor, BTI Lycop... 159 2e-37 emb|AJ388890|AJ388890 AJ388890 Medicago truncatula R108 Medicago... 159 2e-37 emb|AQ849029|AQ849029 LMAJFV1\_lm45c02.x1 Leishmania major FV1 ra... 112 2e-37 gb[U55381]LTU55381 Leishmania tropica P-glycoprotein E gene, com... 114 4e-37 10 emb|AW350529|AW350529 GM210009A10F6R Gm-r1021 Glycine max cDNA 3... 157 7e-37 emb|AQ950989|AQ950989 Sheared DNA-52G21.TF Sheared DNA Trypanoso... 154 5e-36 emb|AF034608|AF034608 Candida albicans YOR1 homolog gene, partia... 153 9e-36 emb|AW775084|AW775084 EST334235 KV3 Medicago truncatula cDNA clo... 153 1e-35 emb|AQ911544|AQ911544 LMAJFV1\_lm86e04.y1 Leishmania major FV1 ra... 127 1e-35 15 emb|AW781305|AW781305 sk68b06.yl Gm-c1016 Glycine max cDNA clone... 152 2e-35 emb|AW441948|AW441948 EST311344 tomato fruit red ripe, TAMU Lyco... 114 2e-35 emb|AW350905|AW350905 GM210009B10G8R Gm-r1021 Glycine max cDNA 3... 151 3e-35 emb|AQ849904|AQ849904 LMAJFV1\_lm51d04.x1 Leishmania major FV1 ra... 118 3e-35emb|AB013851|AB013851 Aspergillus oryzae gene for beta-xylosidas... 122 5e-35 emb|AW775168|AW775168 EST334319 KV3 Medicago truncatula cDNA clo... 150 6e-35 emb|AJ278038|BFU278038 Botryotinia fuckeliana BcatrG gene for MR... 148 4e-34 gb[U62929]FNU62929 Filobasidiella neoformans multidrug resistanc... 93 1e-32 emb|AQ946563|AQ946563 Sheared DNA-49C19.TR Sheared DNA Trypanoso... 142 2e-32 25 emb|AQ646228|AQ646228 RPCI93-DpnII-30J14.TV RPCI93-DpnII Trypano... 141 5e-32 gb|U62931|AFU62931 Aspergillus flavus multidrug resistance prote... 88 1e-31 emb|AW756083|AW756083 sl13f12.yl Gm-c1036 Glycine max cDNA clone... 135 2e-30 emblAI676508|AI676508 etmEST0269 EtH1 Eimeria tenella cDNA clone... 135 2e-30 emb|AQ640396|AQ640396 927P1-5E2.TP 927P1 Trypanosoma brucei geno... 105 4e-30 emb|AI782195|AI782195 EST263074 tomato susceptible, Cornell Lyco... 134 4e-30 emb|AW033521|AW033521 EST277092 tomato callus, TAMU Lycopersicon... 134 6e-30 Query= AF084037.3 s at 20246 s at /id source genbank /description gb|aac95354.1| (af084037) receptor-like protein kinase [arabidopsis 35 thaliana] /blast score 0 /ec number /family /chip nova /gb link (2055 letters) Database: plantfungal 40 661,018 sequences; 426,114,510 total letters Searching.....done Score 45 Sequences producing significant alignments: (bits) Value emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 356 e-115 emb[Y14285]BOY14285 Brassica oleracea mRNA for SFR1 protein. emb[X98520]BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 50 gb[M97667]BNASTKR Brassica napus ssp. oleifera serine/threonine ... 312 9e-99 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 313 1e-97 gb|U00443|BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 315 2e-97 gb|M76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 323 3e-97 emb|AB012106|AB012106 Brassica rapa mRNA for SRK45, complete cds. 307 4e-95 55 gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 308 1e-93 emb|Y18260|BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 313 2e-93 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. 312 1e-92 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 310 3e-92 emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 309 7e-92 60 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 266 1e-90 emb[Y14286]BOY14286 Brassica oleracea SFR3 gene, partial. 182 6e-90

dbj|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 260 1e-89 emb|AF088885|AF088885 Nicotiana tabacum receptor-like kinase CHR... 286 2e-89 emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 262 4e-89 dbi|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 258 3e-87 gb|U20948|ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 197 4e-84 emb[Y12530]BOARLKGEN B.oleraceae gene encoding serine/threonine ... 111 1e-83 gb|BE034855|BE034855 ML05C11 ML Mesembryanthemum crystallinum cD... 268 4e-82 emb|AW031255|AW031255 EST274630 tomato callus, TAMU Lycopersicon... 302 6e-81 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 302 8e-81 10 gb U51741 ITU51741 Ipomoea trifida receptor protein kinase 2 (IR... 232 1e-80 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 105 3e-80 gb|BE057261|BE057261 sm99f12.yl Gm-c1015 Glycine max cDNA clone ... 289 4e-77 emb[Y12531]BOBRLKGEN B.oleraceae gene encoding serine/threonine ... 117 7e-75 emb|AW760240|AW760240 sl59g07.yl Gm-c1027 Glycine max cDNA clone... 261 1e-68 15 emb|AJ245480|BNA245480 Brassica napus slg gene for S-locus glyco... 107 2e-68 emb[AB000970]AB000970 Brassica campestris gene for receptor kina... 109 2e-65 emb|Z18921|BOSRKL B.oleracea gene for S-receptor kinase-like pro... 105 3e-65 dbi|D88193|D88193 Brassica rapa DNA for S-receptor kinase, compl... 104 1e-64 emb|AI895838|AI895838 EST265281 tomato callus, TAMU Lycopersicon... 247 2e-64 emb|AB024422|AB024421S2 Brassica oleracea SRK13-b gene, exon 2, ... 105 5e-64 20 emb|AB024420|AB024419S2 Brassica oleracea SRK13 gene, exon 2, 3,... 105 7e-64 emb|AW442344|AW442344 EST311740 tomato fruit red ripe, TAMU Lyco... 190 1e-63 emb|AI486193|AI486193 EST244514 tomato ovary, TAMU Lycopersicon ... 181 5e-63 emb|AW831390|AW831390 sm22a06.yl Gm-c1028 Glycine max cDNA clone... 183 6e-63 25 emb|AW620957|AW620957 sj98a07.y1 Gm-c1023 Glycine max cDNA clone... 202 1e-62 emb|Z30211|BOSRK29G B.oleracea (alboglabra) srk29 gene. 109 2e-62 emb|AB013718|AB013717S2 Brassica rapa gene for SRK46Bra, exon 2,... 104 2e-62 gb|BE034949|BE034949 ML07F03 ML Mesembryanthemum crystallinum cD... 239 7e-62 emb|X79432|BOSRK3 B.oleracea SRK3 gene. 108 8e-62 30 emblAB000971|AB000971 Brassica campestris pseudogene for recepto... 99 7e-61 emb|AI896155|AI896155 EST265598 tomato callus, TAMU Lycopersicon... 158 3e-59 emb|AW203661|AW203661 sf36g06.yl Gm-c1028 Glycine max cDNA clone... 187 le-58 emb|AW309544|AW309544 sf20h08.x1 Gm-c1028 Glycine max cDNA clone... 227 2e-58 emb|AW736407|AW736407 EST332421 KV3 Medicago truncatula cDNA clo... 226 3e-58 35 emblAW033458|AW033458 EST277029 tomato callus, TAMU Lycopersicon... 215 7e-55 emb|AW220676|AW220676 EST297145 tomato fruit mature green, TAMU ... 215 1e-54 emb|AI967314|AI967314 Ljimpest00-017 Ljimp Lambda HybriZap two... 117 3e-54 emb|AW220675|AW220675 EST297144 tomato fruit mature green, TAMU ... 212 9e-54 emb|AW220677|AW220677 EST297146 tomato fruit mature green, TAMU ... 211 2e-53 40 emb|AW666141|AW666141 sk32f11.yl Gm-c1028 Glycine max cDNA clone... 166 5e-53 emb|AW667985|AW667985 GA\_Ea0012C15 Gossypium arboreum 7-10 dpa ... 119 1e-52 emb|AI822907|AI822907 L30-754T3 Ice plant Lambda Uni-Zap XR expr... 119 1e-51 emb|AA738545|AA738545 SbRLK3 Sorghum bicolor cv. TX430 leaf Sorg... 104 le-51 emb|AI822355|AI822355 L0-804T3 Ice plant Lambda Uni-Zap XR expre... 119 4e-51 45 emb|AI899009|AI899009 EST268452 tomato ovary, TAMU Lycopersicon ... 161 7e-51 emb|AI897876|AI897876 EST267319 tomato ovary, TAMU Lycopersicon ... 146 2e-50 emb|AI771857|AI771857 EST252957 tomato ovary, TAMU Lycopersicon ... 157 9e-50 emb|AI488101|AI488101 EST246423 tomato ovary, TAMU Lycopersicon ... 157 9e-50 emb|AA738544|AA738544 SbRLK2 Sorghum bicolor cv. TX430 leaf Sorg... 84 2e-49 emb[Y16999]TCA16999 Theobroma cacao microsatellite DNA, clone mT... 106 2e-49 50 emb|AI486584|AI486584 EST244905 tomato ovary, TAMU Lycopersicon ... 146 6e-49 emb|AI901283|AI901283 sc31d08.y1 Gm-c1014 Glycine max cDNA clone... 152 8e-49 emb|AI898581|AI898581 EST268024 tomato ovary, TAMU Lycopersicon ... 161 8e-49 emb|AW031816|AW031816 EST275270 tomato callus, TAMU Lycopersicon... 119 2e-48 55 emb|AW054349|AW054349 L30-1774T3 Ice plant Lambda Uni-Zap XR exp... 119 3e-48 emb|AI899156|AI899156 EST268599 tomato ovary, TAMU Lycopersicon ... 161 4e-48 emb|AB041503|AB041503 Populus nigra PnPK1 mRNA for protein kinas... 119 5e-48 emb|AW684339|AW684339 NF015G04NR1F1000 Nodulated root Medicago t... 141 1e-47 gb|U51330|TAU51330 Triticum aestivum leaf rust resistance kinase... 77 6e-47 60 emb|AB041504|AB041504 Populus nigra PnPK2 mRNA for protein kinas... 118 8e-47 emb|AI822347|AI822347 L0-796T3 Ice plant Lambda Uni-Zap XR expre... 106 4e-46

emb|X81833|BOSLR31 B.oleracea mRNA for SLR3-1 protein. 85 5e-46 emb|AI938169|AI938169 sc40d07.y1 Gm-c1014 Glycine max cDNA clone... 185 9e-46 emb|Z18884|BOSRKRPC B.oleracea encoding S-receptor kinase relate... 109 1e-45 emb|AI486331|AI486331 EST244652 tomato ovary, TAMU Lycopersicon ... 161 2e-45 emb|AW687233|AW687233 NF007D09RT1F1077 Developing root Medicago ... 99 2e-45 emb|AI896953|AI896953 EST266396 tomato callus, TAMU Lycopersicon... 158 le-44 emb|AF085164|AF085164 Hordeum vulgare receptor-like kinase LRK10... 77 3e-44 emb|AW039406|AW039406 EST281663 tomato mixed elicitor, BTI Lycop... 112 5e-44 emb|AF142596|AF142596 Nicotiana tabacum LRR receptor-like protei... 65 7e-44 10 emb|AW776492|AW776492 EST335557 DSIL Medicago truncatula cDNA cl... 178 1e-43 emb|AW034993|AW034993 EST279222 tomato callus, TAMU Lycopersicon... 166 1e-42 emb|A67797|A67797 Sequence 2 from Patent WO9743427. 62 2e-42 gblU93048|DCU93048 Daucus carota somatic embryogenesis receptor-... 62 2e-42 emb|AW706972|AW706972 sk20a03.yl Gm-c1028 Glycine max cDNA clone... 174 2e-42 15 emb|AW034624|AW034624 EST278308 tomato callus, TAMU Lycopersicon... 174 2e-42 emb|AI895623|AI895623 EST265066 tomato callus, TAMU Lycopersicon... 173 5e-42 emb|AW687052|AW687052 NF005E02RT1F1017 Developing root Medicago ... 170 3e-41 emb|AW775756|AW775756 EST334821 DSIL Medicago truncatula cDNA cl... 137 5e-41 emb|AF085168|AF085168 Triticum aestivum receptor-like protein ki... 68 6e-41 20 emb|AW597214|AW597214 si71g06.yl Gm-c1031 Glycine max cDNA clone... 116 1e-40 emblAI772117|AI772117 EST253217 tomato resistant, Cornell Lycope... 127 2e-40 emb|AF220603|AF220603 Lycopersicon esculentum VFNT Cherry Pto lo... 96 2e-40 gb|U59316|LEU59316 Lycopersicon esculentum serine/threonine prot... 96 2e-40 emb[Y14600]SBRLK1 Sorghum bicolor mRNA for protein serine/threon... 61 4e-40 25 Query= AC002387.237 at 20269 at /id source genbank /description gb|aab82640.1| (ac002387) putative pectinesterase [arabidopsis thaliana] /blast score 0 /ec number /family pectinesterase /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbin-30 post/entrez/query?db=n&form=6&dopt=g&uid=gb|ac002387| /ncgi http://www.ncgr.org/cgi-bin/ff?ac002387 (1533 letters) Database: plantfungal 35 661,018 sequences; 426,114,510 total letters Searching..... E Score 40 Sequences producing significant alignments: (bits) Value gb|U82977|CSU82977 Citrus sinensis pectinesterase mRNA, complete... 483 0.0 gb|U82976|CSU82976 Citrus sinensis pectinesterase mRNA, complete... 218 e-119 gb[U82975]CSU82975 Citrus sinensis pectinesterase (PECS-2.1) gen... 421 e-116 45 emb|AF152172|AF152172 Solanum tuberosum cultivar Desiree pectin ... 208 e-111 emb|AF229849|AF229849 Vigna radiata pectin methylesterase isofor... 208 e-107 gb|U49330|SLU49330 Solanum lycopersicum pectin methylesterase (P... 205 e-106 emb|Z71752|NPPME2MR N.plumbaginifolia mRNA for pectin methyleste... 177 e-104 emb|Z71753|NPPME3MR N.plumbaginifolia mRNA for pectin methyleste... 177 e-103 50 emb|X94443|VRPECMEST V.radiata mRNA for pectinmethylesterase. gb|U82973|CSU82973 Citrus sinensis pectinesterase (PECS-1.1) gen... 218 8e-97 emb|AF056493|AF056493 Pisum sativum pectin methylesterase mRNA, ... 163 3e-95 emb|AF152171|AF152171 Solanum tuberosum cultivar Desiree pectin ... 190 2e-94 emb|AJ249786|NTA249786 Nicotiana tabacum partial mRNA for pectin... 192 4e-94 55 emb|A17011|A17011 tomato fruit pectin esterase with pPel DNA seq... 190 5e-94 emb|A17010|A17010 tomato fruit pectin esterase seq ID no1. 190 9e-94 emb|X74638|LEPEC1 L.esculentum mRNA for pectin esterase. 190 9e-94 emb[X74639]LEPEC2 L.esculentum mRNA for pectin esterase clone. 188 3e-93 gb|U50986|SLU50986 Solanum lycopersicum pectin methylesterase PM,... 188 6e-93

190 le-92

gb|S66607|S66607 Lycopersicon esculentum pectinmethylesterase-li... 190 8e-93

emb|X07910|LEPECES Tomato mRNA for pectin esterase.

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emblA15983|A15983 L.esculentum mRNA for pectin esterase. 190 1e-92 gb[U50985]SLU50985 Solanum lycopersicum pectin methylesterase PM... 185 2e-92 emb|Z71754|NPPME4MR N.plumbaginifolia mRNa for pectin methyleste... 177 2e-92 emb|X95991|PPPECESTR P.persica mRNA for pectin esterase. 161 3e-92 emb|A24196|A24196 L.esculentum pectin esterase clone pPE1. 188 7e-92 emb|AF081457|AF081457 Pisum sativum pectin methylesterase (rcpme... 136 4e-90 emb[X67425]PSPMEAG Pisum sativum pmeA gene for pectinesterase. emb|AW650699|AW650699 EST329153 tomato germinating seedlings, TA... 271 2e-86 emb[AW696177]AW696177 NF103C11ST1F1085 Developing stem Medicago ... 193 4e-83 10 emb|AB029461|AB029461 Salix gilgiana SgPME1 mRNA for pectin meth... 144 1e-82 emb|AW257370|AW257370 EST305507 KV2 Medicago truncatula cDNA clo... 232 7e-81 emb|AI166540|AI166540 xylem.est.362 Poplar xylem Lambda ZAPII li... 167 4e-79 emb|X85216|PVRNAPE P.vulgaris mRNA for pectinesterase. 170 2e-78 gb|U70676|LEU70676 Lycopersicon esculentum pection methylesteras... 188 1e-77 15 gb[U70677]LEU70677 Lycopersicon esculentum (LePME1) gene, partia... 188 4e-77 gb|U70675|LEU70675 Lycopersicon esculentum fruit-specific pectin... 189 1e-76 emb|AW429110|AW429110 EST306566 tomato flower buds 0-3 mm, Corne... 187 1e-75 emb|AW429158|AW429158 EST306614 tomato flower buds 0-3 mm, Corne... 160 3e-74 emb|AW349192|AW349192 GM210004A21F6R Gm-r1021 Glycine max cDNA 3... 129 1e-72 emb|AW930691|AW930691 EST356534 tomato fruit mature green, TAMU ... 168 1e-70 emb|AW559494|AW559494 EST314542 DSIR Medicago truncatula cDNA cl... 141 2e-70 emb|AW774488|AW774488 EST333639 KV3 Medicago truncatula cDNA clo... 141 2e-70 emb[AW687047]AW687047 NF005D06RT1F1057 Developing root Medicago ... 132 6e-70 emb|AW784073|AW784073 NXNV 117 D06 F Nsf Xylem Normal wood Verti... 153 6e-70 25 gb|BE020131|BE020131 sm41e11.yl Gm-c1028 Glycine max cDNA clone ... 246 2e-69 emb|Y07899|CPSPE1 C.papaya mRNA for pectinesterase. emb|AI897776|AI897776 EST267219 tomato ovary, TAMU Lycopersicon ... 198 1e-68 emb|AW220185|AW220185 EST302668 tomato root during/after fruit s... 172 2e-68 emb|AW666622|AW666622 GA\_ Ea0005C20 Gossypium arboreum 7-10 dpa ... 177 1e-67 30 emb|AW760550|AW760550 sl51g07.yl Gm-c1027 Glycine max cDNA clone... 155 2e-67 emb|AW329215|AW329215 N200427e rootphos(-) Medicago truncatula c... 149 3e-67 emb[X68029]PVVPE2 P.vulgaris PvVPE2 mRNA for pectin esterase. 149 2e-66 emb|AW774605|AW774605 EST333756 KV3 Medicago truncatula cDNA clo... 141 4e-66 emb|AW299038|AW299038 EST305712 KV2 Medicago truncatula cDNA clo... 141 4e-66 35 emb|AW649176|AW649176 EST327630 tomato germinating seedlings, TA... 180 7e-65 emb|AI782839|AI782839 EST263718 tomato susceptible, Cornell Lyco... 156 5e-64 gb|U28148|MSU28148 Medicago sativa putative pectinesterase mRNA,... 134 5e-64 emb[X97762]STBPE1 S.tuberosum mRNA BPE1 for pectin methylesterase. 165 1e-63 emb|AI440753|AI440753 sa53f07.yl Gm-c1004 Glycine max cDNA clone... 161 1e-63 40 emb|AW774519|AW774519 EST333670 KV3 Medicago truncatula cDNA clo... 137 3e-63 emb|AW706153|AW706153 sj52e04.yl Gm-c1033 Glycine max cDNA clone... 242 5e-63 emb|AW649290|AW649290 EST327744 tomato germinating seedlings, TA... 196 8e-63 emb|AW257345|AW257345 EST305482 KV2 Medicago truncatula cDNA clo... 167 9e-63 emb|AW616248|AW616248 EST307287 L. hirsutum trichome, Cornell Un... 173 1e-62 45 emb|AW758821|AW758821 NXNV\_091\_A04\_F Nsf Xylem Normal wood Verti... 160\_8e-62 emb|AW398150|AW398150 EST298033 L. pennellii trichome, Cornell U... 173 1e-61 gb|L27101|PETPPE1A Petunia inflata pectinesterase (PPE1) gene, c... 134 2e-61 emb|AI781139|AI781139 EST262018 tomato susceptible, Cornell Lyco... 205 2e-61 emb|AW932254|AW932254 EST358097 tomato fruit mature green, TAMU ... 148 4e-61 50 gb|U82974|CSU82974 Citrus sinensis pectinesterase (PECS-1.2) gen... 216 6e-60 emb|AW221911|AW221911 EST298722 tomato fruit red ripe, TAMU Lyco... 155 6e-60 emb|AW424141|AW424141 sh61d11.y1 Gm-c1015 Glycine max cDNA clone... 146 2e-59 emb|AW289642|AW289642 NXNV003F09F Nsf Xylem Normal wood Vertical... 160 2e-59 emb[AW888107]AW888107 NXNV 129 C06 F Nsf Xylem Normal wood Verti... 141 6e-59 55 emb|AW620942|AW620942 sj95g05.y1 Gm-c1023 Glycine max cDNA clone... 164 le-58 emb|AW154926|AW154926 EST290291 tomato root deficiency, Cornell ... 190 2e-58 emb|AW623570|AW623570 EST321515 tomato flower buds 3-8 mm, Corne... 135 2e-58 emb|AI780635|AI780635 EST261610 tomato susceptible, Cornell Lyco... 196 2e-58 emb|AW616681|AW616681 EST323092 L. hirsutum trichome, Cornell Un... 177 5e-58 60 emb|AW041247|AW041247 EST284111 tomato mixed elicitor, BTI Lycop... 135 7e-58 emb|AW221863|AW221863 EST298674 tomato fruit red ripe, TAMU Lyco... 190 1e-57

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emb|AW774676|AW774676 EST333827 KV3 Medicago truncatula cDNA clo... 113 1e-57 emb|AW287387|AW287387 LG1\_303\_C07.b1\_A002 Light Grown 1 (LG1) So... 222 6e-57 emb|AW930330|AW930330 EST340883 tomato fruit mature green, TAMU ... 122 9e-57 emb|AW458218|AW458218 sh79h10.y1 Gm-c1016 Glycine max cDNA clone... 120 1e-56 gb|L48178|BNAPECT Brassica campestris pectinesterase mRNA, 3' en... 110 1e-56 5 emb|AW616155|AW616155 EST296925 L. hirsutum trichome, Cornell Un... 177 3e-56 emb|AW221834|AW221834 EST298645 tomato fruit red ripe, TAMU Lyco... 155 3e-56 emblAW617508|AW617508 EST323919 L. hirsutum trichome, Cornell Un... 177 3e-56 emb|AW617079|AW617079 EST323490 L. hirsutum trichome, Cornell Un... 177 3e-56 10 emb|AW616977|AW616977 EST323388 L. hirsutum trichome, Cornell Un... 173 9e-56 emb|AW623150|AW623150 EST321095 tomato flower buds 3-8 mm, Corne... 150 1e-55 emb|AW934244|AW934244 EST360087 tomato fruit mature green, TAMU ... 155 2e-55 emb|X68028|PVVPE3 P.vulgaris PvVPE3 mRNA for pectin esterase. emb|AW616290|AW616290 EST307330 L. hirsutum trichome, Cornell Un... 173 4e-55 15 emb|AW617047|AW617047 EST323458 L. hirsutum trichome, Cornell Un... 173 4e-55 emb|AW220280|AW220280 EST302763 tomato root during/after fruit s... 186 6e-55 emb|X97763|STBPE2 S.tuberosum mRNA BPE2 for pectin methylesterase. 156 6e-55 emb|AI441604|AI441604 sa68e03.y1 Gm-c1004 Glycine max cDNA clone... 155 5e-54 emb|AI731654|AI731654 BNLGHi10367 Six-day Cotton fiber Gossypium... 152 1e-53 20

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Database: plantfungal
661,018 sequences; 426,114,510 total letters

Searching......done

35 Score E

Sequences producing significant alignments: (bits) Value

emb[X57187]PVCHITIN P.vulgaris mRNA for chitinase. 238 e-123 gb|U97522|VVU97522 Vitis vinifera class IV endochitinase (VvChi4... 248 e-112 40 emb|X88803|VURNACHI4 V.unguiculata mRNA for chitinase clase 4 (p... 201 e-107 gb|U97521|VVU97521 Vitis vinifera class IV endochitinase (VvChi4... 247 1e-97 dbj|D45183|D45183 Chenopodium amaranticolor mRNA for chitinase, ... 197 8e-96 dbj|D45182|D45182 Chenopodium amaranticolor mRNA for chitinase, ... 136 6e-95 dbj|D45181|D45181 Chenopodium amaranticolor mRNA for chitinase, ... 136 4e-94 45 emb|AF112966|AF112966 Triticum aestivum chitinase IV precursor (... 227 5e-91 gb|U52845|DCU52845 Daucus carota class IV chitinase EP3-1/H5 (EP... 143 5e-91 emb|AI897733|AI897733 EST267176 tomato ovary, TAMU Lycopersicon ... 164 1e-90 gb|U52848|DCU52848 Daucus carota class IV chitinase EP3B/E6 (EP3... 137 3e-90 gb|U52846|DCU52846 Daucus carota class IV chitinase EP3-2/H1 (EP... 142 4e-90 emb|AI897843|AI897843 EST267286 tomato ovary, TAMU Lycopersicon ... 158 2e-89 gb|U52847|DCU52847 Daucus carota class IV chitinase EP3-3/E7 (EP... 133 4e-89 emb[X61488]BNCHITIN B.napus mRNA for chitinase. 181 5e-85 emb|Z46948|SNCHJET15 S.nigra mRNA for chitinase, pathogenesis-re... 166 2e-84 emb|AI897217|AI897217 EST266756 tomato ovary, TAMU Lycopersicon ... 164 2e-84 55 gb|L25826|BEUSP2X Sugar beet chitinase (SP2) mRNA, complete cds. 185 9e-83 emb|X75945|BVCH4RNA B.vulgaris Ch4 mRNA for chitinase. 127 8e-80 emb|A23392|A23392 B.vulgaris mRNA for chitinase 4 (B15). 127 8e-80 emb|AI898279|AI898279 EST267722 tomato ovary, TAMU Lycopersicon ... 126 2e-78 gb|BE034166|BE034166 MG05H02 MG Mesembryanthemum crystallinum cD... 167 4e-78 60 emb|AW691007|AW691007 NF036E09ST1F1000 Developing stem Medicago ... 131 5e-78

emb[AW030814]AW030814 EST274069 tomato callus, TAMU Lycopersicon... 126 1e-77

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emb|AF034566|AF034566 Gossypium hirsutum class I chitinase mRNA,... 108 2e-51 emb|AF043248|AF043248 Solanum tuberosum class I chitinase (ChtC2... 100 3e-51 gb|U83592|MSU83592 Medicago sativa class I chitinase mRNA, compl... 112 4e-51 gb|U83591|MSU83591 Medicago sativa class I chitinase mRNA, compl... 112 4e-51 emb|Z54234|VVCHIT1MR V.vinifera mRNA for chitinase. gb|U02608|STU02608 Solanum tuberosum chitinase (chtB4) mRNA, par... 107 1e-50 emb[X88800[VURNACHI1 V.unguiculata mRNA for chitinase clase 1 (p... 107 2e-50 emb|AF000964|AF000964 Poa pratensis chitinase (Chi1) gene, compl... 100 2e-50 gb|U78888|GHU78888 Gossypium hirsutum class I endochitinase mRNA... 103 4e-50 10 emb|X63899|PSCHITIN P.sativum mRNA for chitinase. 94 le-49 emb|AB015655|AB015655 Cucurbita sp. mRNA for chitinase, complete... 100 7e-49 gb|BE034450|BE034450 MH05B01 MH Mesembryanthemum crystallinum cD... 84 1e-48 gb|U02287|HVU02287 Hordeum vulgare cultivar NK1558 chitinase gen... 111 3e-48 emb|A16119|A16119 Intracellular chitinase mRNA (SEQ ID NO: 2). 15 gblM15173|TOBECH Tobacco (N.tabacum) endochitinase mRNA, partial... 108 6e-48 Query= Y14590.5\_g\_at 20288\_g\_at /id\_source genbank /description emb|caa74930.1| (y14590) class iv chitinase [arabidopsis thaliana] 20 /blast\_score 1.00e-166/ec\_number /family /chip nova/gb\_link /ncgi (825 letters) Database: plantfungal 25 661,018 sequences; 426,114,510 total letters E Score 30 Sequences producing significant alignments: (bits) Value emb|X57187|PVCHITIN P.vulgaris mRNA for chitinase. 238 e-123 gb|U97522|VVU97522 Vitis vinifera class IV endochitinase (VvChi4... 248 e-112 emb|X88803|VURNACHI4 V.unguiculata mRNA for chitinase clase 4 (p... 201 e-107 35 gb|U97521|VVU97521 Vitis vinifera class IV endochitinase (VvChi4... 247 1e-97 dbj|D45183|D45183 Chenopodium amaranticolor mRNA for chitinase, ... 197 8e-96 dbi|D45182|D45182 Chenopodium amaranticolor mRNA for chitinase, ... 136 6e-95 dbi|D45181|D45181 Chenopodium amaranticolor mRNA for chitinase, ... 136 4e-94 emblAF112966|AF112966 Triticum aestivum chitinase IV precursor (... 227 5e-91 40 gb|U52845|DCU52845 Daucus carota class IV chitinase EP3-1/H5 (EP... 143 5e-91 emb|AI897733|AI897733 EST267176 tomato ovary, TAMU Lycopersicon ... 164 1e-90 gb|U52848|DCU52848 Daucus carota class IV chitinase EP3B/E6 (EP3... 137 3e-90 gb|U52846|DCU52846 Daucus carota class IV chitinase EP3-2/H1 (EP... 142 4e-90 emb|AI897843|AI897843 EST267286 tomato ovary, TAMU Lycopersicon ... 158 2e-89 45 gb|U52847|DCU52847 Daucus carota class IV chitinase EP3-3/E7 (EP... 133 4e-89 emb|X61488|BNCHITIN B.napus mRNA for chitinase. 181 5e-85 emb|Z46948|SNCHJET15 S.nigra mRNA for chitinase, pathogenesis-re... 166 2e-84 emb|AI897217|AI897217 EST266756 tomato ovary, TAMU Lycopersicon ... 164 2e-84 gb|L25826|BEUSP2X Sugar beet chitinase (SP2) mRNA, complete cds. 185 9e-83 50 emb[X75945]BVCH4RNA B.vulgaris Ch4 mRNA for chitinase. 127 8e-80 emb|A23392|A23392 B.vulgaris mRNA for chitinase 4 (B15). 127 8e-80 emblAI898279|AI898279 EST267722 tomato ovary, TAMU Lycopersicon ... 126 2e-78 gb|BE034166|BE034166 MG05H02 MG Mesembryanthemum crystallinum cD... 167 4e-78 emb|AW691007|AW691007 NF036E09ST1F1000 Developing stem Medicago ... 131 5e-78 emb|AW030814|AW030814 EST274069 tomato callus, TAMU Lycopersicon... 126 1e-77 emb|AW035013|AW035013 EST279284 tomato callus, TAMU Lycopersicon... 126 2e-77 emb|AI485982|AI485982 EST244303 tomato ovary, TAMU Lycopersicon ... 126 2e-77 gb|BE034428|BE034428 MH04G02 MH Mesembryanthemum crystallinum cD... 170 3e-77 dbj|D45184|D45184 Chenopodium amaranticolor mRNA for chitinase, ... 136 2e-76 gb|BE034975|BE034975 ML07H10 ML Mesembryanthemum crystallinum cD... 156 7e-75 60 emb|AF090336|AF090336 Citrus sinensis chitinase CHI1 (chi1) mRNA... 215 1e-74

gbBE034976BE034976 ML07H11 ML Mesembryanthemum crystallinum cD... 160 1e-74 emb|AI776153|AI776153 EST257241 tomato resistant, Cornell Lycope... 164 1e-74 emb|Z46950|SNCHJET19 S.nigra mRNA chitinase class II, pathogenes... 139 1e-74 emb|AW029733|AW029733 EST272988 tomato callus, TAMU Lycopersicon... 126 2e-74 5 gb|L42467|PIACHI Picea glauca chitinase (chi) mRNA, complete cds. 104 3e-74 emb|AI897657|AI897657 EST267100 tomato ovary, TAMU Lycopersicon ... 156 4e-72 emb[X74919]PVGEC9 P.vulgaris gene for endochitinase. 238 6e-71 emb|AW648023|AW648023 EST326477 tomato germinating seedlings, TA... 126 2e-70 gb|BE034406|BE034406 MH04D10 MH Mesembryanthemum crystallinum cD... 132 2e-68 10 emb|AW033122|AW033122 EST276681 tomato callus, TAMU Lycopersicon... 126 3e-68 gb|BE034497|BE034497 MH05H03 MH Mesembryanthemum crystallinum cD... 114 2e-67 emb|AF112963|AF112963 Triticum aestivum chitinase II precursor (... 154 1e-66 emb|AW684552|AW684552 NF018C03NR1F1000 Nodulated root Medicago t... 122 5e-66 emb|AW922776|AW922776 DG1\_46\_C01.g1\_A002 Dark Grown 1 (DG1) Sorg... 215 5e-65 15 gb|U21848|BNU21848 Brassica napus chitinase class IV (LSC222) mR... 79 2e-64 gb|BE035287|BE035287 MM04H04 MM Mesembryanthemum crystallinum cD... 208 6e-63 emb|AW680953|AW680953 WS1\_9\_A06.b1\_A002 Water-stressed 1 (WS1) S... 224 7e-63 emb|AI729668|AI729668 BNLGHi13889 Six-day Cotton fiber Gossypium... 121 1e-61 emb|AI055037|AI055037 coau0002N18 Cotton Boll Abscission Zone cD... 111 3e-60 20 gbBE034616BE034616 ML04B04 ML Mesembryanthemum crystallinum cD... 101 3e-60 gbBE033502BE033502 ME03F10 ME Mesembryanthemum crystallinum cD... 100 4e-59 emb|AW924422|AW924422 WS1\_69\_C06.b1\_A002 Water-stressed 1 (WS1) ... 171 4e-59 emb|X16939|NTECHITR Nicotiana tabacum mRNA for endochitinase (EC... 108 2e-58 emb|AW922735|AW922735 DG1\_45\_B06.g1\_A002 Dark Grown 1 (DG1) Sorg... 224 6e-58 25 emb|AW746695|AW746695 WS1 54 E02.g1 A002 Water-stressed 1 (WS1) ... 224 6e-58 gb|S44869|S44869 basic chitinase [Nicotiana tabacum=tobacco, cv ... 108 2e-57 gb|BE034481|BE034481 MH05F02 MH Mesembryanthemum crystallinum cD... 208 2e-57 emblAI488583|AI488583 EST246922 tomato ovary, TAMU Lycopersicon ... 126 3e-57 emb|Z78202|PACHI1 Persea americana mRNA for endochitinase. 118 4e-57 30 emb|AW746018|AW746018 WS1\_38\_H11.g1\_A002 Water-stressed 1 (WS1) ... 221 4e-57 emb[X15494|STCHITIN Potato endochitinase gene (EC 3.2.1.14). 111 le-56 emb|X07130|STCHIT Solanum tuberosum mRNA for endochitinase (EC 3... 111 1e-56 emb|AI897391|AI897391 EST266834 tomato ovary, TAMU Lycopersicon ... 126 1e-56 emb|AF000966|AF000966 Poa pratensis chitinase (Chi2) gene, compl... 110 7e-56 35 emb|AW924229|AW924229 WS1 51 H04.b1 A002 Water-stressed 1 (WS1) ... 215 3e-55 emb|AW676775|AW676775 DG1 14\_C09.g1 A002 Dark Grown 1 (DG1) Sorg... 215 3e-55 emb|X14133|STENCHIT Potato mRNA fragment for endochitinase (EC 3... 105 4e-55 emb|X76041|TACHIG T.aestivum (Chinese spring) chi gene for endoc... 109 6e-55 emb|AW745819|AW745819 WS1\_37\_H11.g1\_A002 Water-stressed 1 (WS1) ... 214 7e-55 emb|Z15140|LECHI9 L.esculentum mRNA for chitinase. 40 108 2e-54 emb|AF000965|AF000965 Poa pratensis chitinase (Chi3) pseudogene ... 110 2e-54 gb|M94106|ALCCHINTIA Allium sativum chitinase mRNA, 3' end. 107 5e-54 emb|AF098302|AF098302 Brassica juncea chitinase mRNA, complete cds. 110 6e-54 gb|M13968|PHVCHM P.vulgaris chitinase mRNA, complete cds. 45 gb|U02607|STU02607 Solanum tuberosum chitinase (chtB3) mRNA, par... 105 1e-53 gb|U02605|STU02605 Solanum tuberosum chitinase (chtB1) mRNA, par... 105 1e-53 emb|AF043247|AF043247 Solanum tuberosum class I chitinase (ChtC1... 105 1e-53 gb|U02606|STU02606 Solanum tuberosum chitinase (chtB2) mRNA, par... 104 2e-53 gb|S43926|S43926 CH5B=chitinase [Phaseolus vulgaris=beans, cv Sa... 110 6e-53 50 gb|L37876|PEACHI2I Pisum sativum chitinase class I (chi2) gene, ... 112 9e-53 emb|Y10373|MTCHITIN1 M.truncatula mRNA for chitinase. 112 9e-53 emb|AW029730|AW029730 EST272985 tomato callus, TAMU Lycopersicon... 125 1e-52 gb|BE033398|BE033398 ME01A01 ME Mesembryanthemum crystallinum cD... 205 3e-52 emb|AJ012821|CAR012821 Cicer arietinum mRNA for class I chitinase. 114 1e-51 55 gb|BE034447|BE034447 MH05A08 MH Mesembryanthemum crystallinum cD... 83 1e-51 emb|AF034566|AF034566 Gossypium hirsutum class I chitinase mRNA,... 108 2e-51 emb|AF043248|AF043248 Solanum tuberosum class I chitinase (ChtC2... 100 3e-51 gb[U83592]MSU83592 Medicago sativa class I chitinase mRNA, compl... 112 4e-51 gb[U83591]MSU83591 Medicago sativa class I chitinase mRNA, compl... 112 4e-51 60 emb|Z54234|VVCHIT1MR V.vinifera mRNA for chitinase. 102 7e-51 gb|U02608|STU02608 Solanum tuberosum chitinase (chtB4) mRNA, par... 107 1e-50

emb|X88800|VURNACHI1 V.unguiculata mRNA for chitinase clase 1 (p... 107 2e-50 emb|AF000964|AF000964 Poa pratensis chitinase (Chi1) gene, compl... 100 2e-50 gb[U78888]GHU78888 Gossypium hirsutum class I endochitinase mRNA... 103 4e-50 emb|X63899|PSCHITIN P.sativum mRNA for chitinase. 94 le-49 emb|AB015655|AB015655 Cucurbita sp. mRNA for chitinase, complete... 100 7e-49 gbBE034450BE034450 MH05B01 MH Mesembryanthemum crystallinum cD... 84 1e-48 gb[U02287]HVU02287 Hordeum vulgare cultivar NK1558 chitinase gen... 111 3e-48 emb|A16119|A16119 Intracellular chitinase mRNA (SEQ ID NO: 2). gb[M15173]TOBECH Tobacco (N.tabacum) endochitinase mRNA, partial... 108 6e-48 10 Query= M92353.4 s at 20291 s at /id source genbank /description gb|aaa32738.1| (m92353) anthranilate synthase alpha subunit [arabidopsis thaliana] /blast score 0 /ec number /family /chip nova 15 /gb link /ncgi (1788 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 20 Searching..... Score Sequences producing significant alignments: (bits) Value 25 gb|L34344|RTAANTSYNB Ruta graveolens anthranilate synthase alpha... 772 0.0 gb|L34343|RTAANTSYNA Ruta graveolens anthranilate synthase alpha... 467 0.0 emb|AF079168|AF079168 Nicotiana tabacum feedback-insensitive ant... 365 0.0 emb|AW931942|AW931942 EST357785 tomato fruit mature green, TAMU ... 356 3e-97 30 emb|AW218352|AW218352 EST303535 tomato radicle, 5 d post-imbibit... 210 1e-73 emb|AL031966|SPCC1442 S.pombe chromosome III cosmid c1442. 209 6e-69 emb|AW651095|AW651095 EST329549 tomato germinating seedlings, TA... 222 2e-64 dbj|D89256|D89256 Schizosaccharomyces pombe mRNA, partial cds, c... 205 5e-64 emb[AW982499]AW982499 HVSMEg0003G22f Hordeum vulgare pre-anthesi... 215 2e-58 35 emb|AW460005|AW460005 si07d11.yl Gm-c1029 Glycine max cDNA clone... 116 1e-57 gb[U18839]SCE9747 Saccharomyces cerevisiae chromosome V cosmids ... 123 4e-48 emb|X68327|SCTRP2 S.cerevisiae TRP2 gene for anthranilate syntha... 123 4e-48 emb|AW719463|AW719463 LjNEST5b1r Lotus japonicus nodule library,... 184 1e-45 emb|AI736775|AI736775 sb33d01.y1 Gm-c1012 Glycine max cDNA clone... 104 2e-40 40 gb|K01388|YSCTRP2 Yeast (S.cerevisiae) TRP2 gene coding for anth... 122 4e-38 emb|AL032684|SPBP8B7 S.pombe chromosome II p1 p8B7. 95 6e-25 emb|AW509018|AW509018 si39b01.yl Gm-r1030 Glycine max cDNA clone... 113 5e-24 gb[T14852]T14852 crs299 lambdaZAPST Ricinus communis cDNA clone ... 107 2e-22 emb|AW223881|AW223881 EST300692 tomato fruit red ripe, TAMU Lyco... 103 4e-21 emb|AF119554|AF119554 Plasmodium falciparum para-aminobenzoic ac... 67 6e-10 45 emb|AL111470|CNS019CM Botrytis cinerea strain T4 cDNA library un... 45 2e-05 emb|AI329873|AI329873 b9g02ne.r1 Neurospora crassa evening cDNA ... 48 3e-04 emb|AW224247|AW224247 EST300974 tomato fruit red ripe, TAMU Lyco... 46 7e-04 emb|AF149719|AF149719 Aspergillus fumigatus para aminobenzoic ac... 42 0.012 50 emb|AQ448372|AQ448372 mgxb0020C12f CUGI Rice Blast BAC Library P... 39 0.15 emb|AQ324360|AQ324360 mgxb0018K01r CUGI Rice Blast BAC Library P... 39 0.15 emb|AQ160089|AQ160089 mgxb0003G09r CUGI Rice Blast BAC Library P... 39 0.15 emb|AW599019|AW599019 gb01b03.y1 Moss EST library PPN Physcomitr... 37 0.38 emb|AW599000|AW599000 ga99h03.y1 Moss EST library PPN Physcomitr... 37 0.38 55 emb|AW678847|AW678847 WS1\_1\_A04.g1\_A002 Water-stressed 1 (WS1) S... 35 0.88 emb|AW680390|AW680390 WS1\_52 D12.g1\_A002 Water-stressed 1 (WS1) ... 35 0.89 emb|AW678385|AW678385 WS1\_15\_H06.g1\_A002 Water-stressed 1 (WS1) ... 35 0.89 emb|AW747146|AW747146 WS1\_66\_E07.b1\_A002 Water-stressed 1 (WS1) ... 35 0.89 emb|AW678361|AW678361 WS1\_15\_H06.b1\_A002 Water-stressed 1 (WS1) ... 35 0.90 60 emb|AW745749|AW745749 WS1\_37\_D12.b1\_A002 Water-stressed 1 (WS1) ... 35 0.90 emb|AW747427|AW747427 WS1 68 B09.b1 A002 Water-stressed 1 (WS1) ... 35 0.90

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      emb|AW744836|AW744836 LG1_384_E07.g1_A002 Light Grown 1 (LG1) So... 35 0.90
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      emb|AW746170|AW746170 WS1 39 B05.g1 A002 Water-stressed 1 (WS1) ... 35 0.90
      emblAW922317lAW922317 DG1 17 E06.g1 A002 Dark Grown 1 (DG1) Sorg... 35 0.91
      emb|AQ648582|AQ648582 RPCI93-EcoRI-1M22.TP RPCI93-EcoRI Trypanos... 36 0.99
      emb|AQ643551|AQ643551 RPCI93-EcoRI-3124.TJ RPCI93-EcoRI Trypanos... 36 0.99
      emb|AI443370|AI443370 sa31b05.x1 Gm-c1004 Glycine max cDNA clone... 35 1.4
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      emb|AW101313|AW101313 sd77d08.yl Gm-c1009 Glycine max cDNA clone... 35 1.4
      emb|AW678030|AW678030 WS1 12 B10.g1 A002 Water-stressed 1 (WS1) ... 35 1.4
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Database: plantfungal

15 661,018 sequences; 426,114,510 total letters

Searching.....done

Score F

20 Sequences producing significant alignments:

(bits) Value

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Database: plantfungal

661,018 sequences; 426,114,510 total letters

10

5

Searching......done

Score E

Sequences producing significant alignments: (bits) Value 15

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 261 e-116

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35	emb AA680502 AA680502 T3691 Bloodstream form of serodeme ILTat1 34 4.0
<i></i>	emb AW691484 AW691484 NF041F05ST1F1000 Developing stem Medicago 34 4.0
: .	emb AL356815 NCB24H17 Neurospora crassa DNA linkage group II BAC 30 4.2
	emb AF181702 AF181702 Leptosphaeria contecta 18S ribosomal RNA g 34 5.5
40	emb AW145916 AW145916 ga35g12.yl Moss EST library PPN Physcomitr 34 5.5
	emb AW617253 AW617253 EST323664 L. hirsutum trichome, Cornell Un 34 5.5
	emb AW615864 AW615864 EST325362 tomato flower buds 0-3 mm, Corne 34 5.5
•	emb AW691785 AW691785 NF044B05ST1F1000 Developing stem Medicago 34 5.5
	dbj D38256 YSCSCT1 Yeast gene for suppressor of ctr mutation. 34 5.5
45	gb U26175 LDU26175 Leishmania donovani lipophosphoglycan biosynt 34 5.5
	emb AB029613 AB029613 Aspergillus terreus TOP2 gene for type II 34 5.5
	emb AW696259 AW696259 NF104F08ST1F1074 Developing stem Medicago 34 5.5
	emb AC004710 AC004710 Plasmodium falciparum chromosome 12, *** S 33 7.6
<b>5</b> 0	emb AV421315 AV421315 AV421315 Lotus japonicus young plants (two 33 7.6
50	emb AW599293 AW599293 gb13a06.yl Moss EST library PPN Physcomitr 33 7.6
	emb AC005507 AC005507 Plasmodium falciparum chromosome 12 clone 33 7.6
	gb U16862 PFU16862 Plasmodium falciparum HSP70-like protein gene 33 7.6
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55	emb AW694856 AW694856 NF080G09ST1F1071 Developing stem Medicago 33 7.6
	emb AV408901 AV408901 AV408901 Lotus japonicus young plants (two 33 7.6
	emb AV412721 AV412721 AV412721 Lotus japonicus young plants (two 33 7.6
	emb AI759851 AI759851 sb65b08.yl Gm-c1017 Glycine max cDNA clone 33 7.6
60	emb AZ124240 AZ124240 T223104b Medicago truncatula BAC library M 33 7.6
60	emb Y08926 PFAARP1PR P.falciparum mRNA for AARP1 protein, partial. 33 7.6
	emblAV410224lAV410224 AV410224 Lotus japonicus voung plants (two 33 7 6

emblAO849926|AO849926 LMAJFV1 lm51f05.x1 Leishmania major FV1 ra... 33 7.6 emblAL034557PFMAL4P1 Plasmodium falciparum chromosome 4 strain ... 27 8.3 emblAO944606|AO944606 Sheared DNA-48P15.TR Sheared DNA Trypanoso... 28 8.4 emb|AF099095|AF099095 Samanea saman pulvinus inward-rectifying c... 26 9.8 Query= AC005896.161\_g\_at 20620\_g\_at /id\_source genbank /description gb[aac98070.1] (ac005896) putative c2h2-type zinc finger protein [arabidopsis thaliana] /blast score 8.00e-92 /ec number /family /chip nova /gb link /ncgi 10 (864 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 15 Score E Sequences producing significant alignments: (bits) Value 20 emb|AB006600|AB006600 Petunia x hybrida mRNA for ZPT2-13, comple... 86 1e-30 emb|AB000451|AB000451 Petunia hybrida mRNA for PEThy; ZPT2-5, com... 74 5e-29 emb|AW685937|AW685937 NF031H10NR1F1000 Nodulated root Medicago L.. 76 9e-29 emb|AB006599|AB006599 Petunia x hybrida mRNA for ZPT2-12, comple... 77 1e-28 emb|AB006601|AB006601 Petunia x hybrida mRNA for ZPT2-14, comple... 72 2e-26 25 emb|AI898309|AI898309 EST267752 tomato ovary, TAMU Lycopersicon ... 68 3e-26 emb|AI485362|AI485362 EST243683 tomato ovary, TAMU Lycopersicon ... emb|AI483886|AI483886 EST249757 tomato ovary, TAMU Lycopersicon... emblAW030869lAW030869 EST274159 tomato callus, TAMU Lycopersicon... 68 5e-24 emb|AI966235|AI966235 sc35g05.yl Gm-c1014 Glycine max cDNA clone... 72 3e-23 30 emb|AB006603|AB006603 Petunia x hybrida mRNA for ZPT2-8, complet... 65 1e-22 emb|AI486627|AI486627 EST244948 tomato ovary, TAMU Lycopersicon ... 61 1e-22 emb|AI485555|AI485555 EST243876 tomato ovary, TAMU Lycopersicon ... emb|AI488341|AI488341 EST246663 tomato ovary, TAMU Lycopersicon ... emb|AI486228|AI486228 EST244549 tomato ovary, TAMU Lycopersicon ... 35 emb|AI959966|AI959966 sc35g05.x1 Gm-c1014 Glycine max cDNA clone... emb|AI771123|AI771123 EST252223 tomato ovary, TAMU Lycopersicon ... emb|AI897376|AI897376 EST266819 tomato ovary, TAMU Lycopersicon ... 68 5e-22 emb|AI894711|AI894711 EST264154 tomato callus, TAMU Lycopersicon... 68 6e-22 emb|AV422177|AV422177 AV422177 Lotus japonicus young plants (two... 103 2e-21 emb|AB006602|AB006602 Petunia x hybrida mRNA for ZPT2-7, complet... 66 6e-21 emb|AI485123|AI485123 EST243427 tomato ovary, TAMU Lycopersicon ... 68 6e-21 emb|AW216442|AW216442 EST295072 tomato callus, TAMU Lycopersicon... 68 8e-21 emb|AW687462|AW687462 NF009H03RT1F1031 Developing root Medicago ... 70 2e-20 emb|AI771824|AI771824 EST252924 tomato ovary, TAMU Lycopersicon ... 68 2e-20 45 emblAW034640|AW034640 EST278324 tomato callus, TAMU Lycopersicon... 68 7e-20 emb[AW033868]AW033868 EST277439 tomato callus, TAMU Lycopersicon... 72 1e-19. emb|AI487993|AI487993 EST246315 tomato ovary, TAMU Lycopersicon ... 68 le-18 emb|AW684455|AW684455 NF017B06NR1F1000 Nodulated root Medicago t... 68 2e-18 emb|X60700|PHEPF1 P.hybrida EPF1 gene for DNA-binding protein. 50 emb|AB006604|AB006604 Petunia x hybrida mRNA for ZPT2-9, complet... 59 5e-18 emb|AI489727|AI489727 EST248066 tomato ovary, TAMU Lycopersicon ... 50 1e-16 emb|AW034622|AW034622 EST278306 tomato callus, TAMU Lycopersicon... 87 2e-16 emb|AB035132|AB035132 Petunia x hybrida gene for C2H2 zinc-finge... 57. 1e-15 emb[AB006597]AB006597 Petunia x hybrida mRNA for ZPT2-10, comple... 57 1e-15 55 emb|AB035133|AB035133 Petunia x hybrida gene for C2H2 zinc-finge... 56 2e-15 emb|AB006605|AB006605 Petunia x hybrida mRNA for ZPT3-3, complet... 56 2e-15 emb|AW777036|AW777036 M111167e DSIR Medicago truncatula cDNA clo... 74 2e-15 emb|AI488445|AI488445 EST246784 tomato ovary, TAMU Lycopersicon ... 50 4e-15 emb|AF119050|AF119050 Datisca glomerata zinc-finger protein 1 (z... 53 7e-15 60 emb|AB000455|AB000455 Petunia hybrida mRNA for PEThy;ZPT4-1, com... 55 9e-15 emb|AI988657|AI988657 sd06b03.yl Gm-c1020 Glycine max cDNA clone... 51 le-14

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•	dbi[D16416 WHTWZF1B Wheat mRNA for WZF1, complete cds. 53 2e-14
•	dbj[D26084]PETZFDB2 Petunia mRNA for zinc-finger DNA binding pro 52 3e-14
10	emb Y18788 MSY18788 Medicago sativa mRNA for putative TFIIIA (or 51 3e-14
10	emb AW560934 AW560934 EST315982 DSIR Medicago truncatula cDNA cl 51 3e-14
	emb AI988290 AI988290 sc98f10.y1 Gm-c1020 Glycine max cDNA clone 52 3e-14
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	dbj D26083 PETZFDB1 Petunia hybrida gene for zinc-finger DNA bin 52 6e-14
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15	emb AW729218 AW729218 GA_Ea0024G18 Gossypium arboreum 7-10 dpa 51 1e-13
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	dbj D26086 PETZFP4 Petunia zinc-finger protein gene. 49 3e-12
	emb AW616587 AW616587 EST322998 L. hirsutum trichome, Cornell Un 47 9e-12
	emb AW924420 AW924420 WS1_69_C08.b1_A002 Water-stressed 1 (WS1) 50 9e-12
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40	emb AI487287 AI487287 EST245609 tomato ovary, TAMU Lycopersicon 46 1e-11
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45	emb X87374 PSZINCFIN P.sativum putative zinc finger protein. 64 3e-11
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	emb AI966679 AI966679 sc55al1.yl Gm-c1015 Glycine max cDNA clone 52 1e-10
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	• • • • • • • • • • • • • • • • • • • •
55	
<i>JJ</i>	emb AB006598 AB006598 Petunia x hybrida mRNA for ZPT2-11, comple 64 7e-10
	emb AW219736 AW219736 EST302218 tomato root during/after fruit s 46 7e-10
	emb AW924443 AW924443 WS1_69_C08.g1_A002 Water-stressed 1 (WS1) 48 9e-10
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Query= AL049658.102\_at 20625\_at /id\_source genbank /description emb|cab41131.1| (al049658) hypothetical protein [arabidopsis thaliana]

5

/blast score 0 /ec number /family /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|al049658| /ncgi http://www.ncgr.org/cgi-bin/ff?al049658 (1890 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

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Score

Sequences producing significant alignments: (bits) Value 15 emb|AW559251|AW559251 EST306087 DSIR Medicago truncatula cDNA cl... 169 3e-49 emb[AW032676]AW032676 EST276235 tomato callus, TAMU Lycopersicon... 181 1e-44 emb|AI778594|AI778594 EST259473 tomato susceptible, Cornell Lyco... 181 1e-44 emb|AW774727|AW774727 EST333878 KV3 Medicago truncatula cDNA clo... 169 6e-41 emb|AV423915|AV423915 AV423915 Lotus japonicus young plants (two... 163 3e-39' 20 emb|AW559836|AW559836 EST314884 DSIR Medicago truncatula cDNA cl... 131 6e-37 emb|AW349142|AW349142 GM210003B22H5R Gm-r1021 Glycine max cDNA 3... 107 3eemb|AW560074|AW560074 EST315122 DSIR Medicago truncatula cDNA cl... 84 6e-23 emb|AI855891|AI855891 sc30h08.y1 Gm-c1014 Glycine max cDNA clone... 100 4e-21 25 emblAW560073|AW560073 EST315121 DSIR Medicago truncatula cDNA cl... 77 5e-21 emb[AW225676]AW225676 ST70B12 Pine TriplEx shoot tip library Pin... 91 3e-17 emb|AW870069|AW870069 NXNV 123 G03 F Nsf Xylem Normal wood Verti... 68 3e-17 emb|AW587665|AW587665 ST63B10 Pine TriplEx shoot tip library Pin... 87 4e-17 emb|AW011208|AW011208 ST18A05 Pine TriplEx shoot tip library Pin... 89 1e-16 30 emb|AW438038|AW438038 ST83G07 Pine TriplEx shoot tip library Pin... 84 5e-15 emb|AV417662|AV417662 AV417662 Lotus japonicus young plants (two... 54 7e-15 emb[AI920196]AI920196 1726 Pine Lambda Zap Xylem library Pinus t... 54 2e-14 emb|AW981483|AW981483 EST392636 DSIL Medicago truncatula cDNA cl... 70 5e-13 emblAW689240|AW689240 NF017A06ST1F1000 Developing stem Medicago ... 76 8e-13 35 emb|AW684289|AW684289 NF015A11NR1F1000 Nodulated root Medicago t... 70 1e-12 emblAW933515|AW933515 EST359274 tomato fruit mature green, TAMU ... 48 3e-12 emb|AW685948|AW685948 NF036G07NR1F1000 Nodulated root Medicago t... 58 8e-12 emb|AW687095|AW687095 NF005H11RT1F1095 Developing root Medicago ... 72 1e-11 emb|AW686129|AW686129 NF033H12NR1F1000 Nodulated root Medicago t... 47 le-l1 40 emb|AI894750|AI894750 EST264193 tomato callus, TAMU Lycopersicon... 59 3e-11 emb|AW698315|AW698315 NXNV 071 G01 F Nsf Xylem Normal wood Verti... 71 3e-11 emb|AW290408|AW290408 NXNV019H08F Nsf Xylem Normal wood Vertical... 67 4c-11 emblAW065112IAW065112 ST39G09 Pine TriplEx shoot tip library Pin... 51 5e-11 emblAW560897[AW560897 EST315945 DSIR Medicago truncatula cDNA cl... 55 7e-11 45 emb|AW257406|AW257406 EST305543 KV2 Medicago truncatula cDNA clo... 55 7e-11 emb|AW217058|AW217058 EST295772 tomato callus, TAMU Lycopersicon... 51 5e-10 emb|AV407304|AV407304 AV407304 Lotus japonicus young plants (two... 65 2e-09 gb|BE049795|BE049795 NXNV 144 C08 F Nsf Xylem Normal wood Vertic... 53 7e-09 emb|AW746848|AW746848 WS1 55 B02.g1 A002 Water-stressed 1 (WS1) ... 57 5e-07 emb|AW888098|AW888098 NXNV 108 F09 F Nsf Xylem Normal wood Verti... 56 1e-06 50 emblAW032082|AW032082 EST275536 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AW032864|AW032864 EST276423 tomato callus, TAMU Lycopersicon... 46 2e-06 emb|AW736866|AW736866 NXNV 083 H05\_F Nsf Xylem Normal wood Verti... 52 2e-05 emb|AW775153|AW775153 EST334304 KV3 Medicago truncatula cDNA clo... 51 4e-05 55 emb|AV411498|AV411498 AV411498 Lotus japonicus young plants (two... 47 5e-04 emb|AW934555|AW934555 EST353447 tomato flower buds 0-3 mm, Corne... 36 9e-04 emb|AW010126|AW010126 ST02C06 Pine TriplEx shoot tip library Pin... 46 0.001 emb|AW031593|AW031593 EST275047 tomato callus, TAMU Lycopersicon... 31 0.001 emb|AW746715|AW746715 WS1 55 B02.b1 A002 Water-stressed 1 (WS1) ... 39 0.001 emb|AW720513|AW720513 LjNEST18e1r Lotus japonicus nodule library... 46 0.001 60 emb|AW773847|AW773847 EST332833 KV3 Medicago truncatula cDNA clo... 35 0.004

	emb AZ051219 AZ051219 Gm_UMb001_062_F16R UMN Soybean BAC Library 38 0.006
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,	emb AW687408 AW687408 NF009C08RT1F1065 Developing root Medicago 35 0.019
	• • •
	_ : =
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	emb AW559393 AW559393 EST314441 DSIR Medicago truncatula cDNA cl 37 0.56
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40	emb AW033298 AW033298 EST276869 tomato callus, TAMU Lycopersicon 35 2.0
40	gb[L36822]SSNPAL Stylosanthes humilis phenylalanine ammonia-lyas 35 2.0
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45	emb AW685287 AW685287 NF027B11NR1F1000 Nodulated root Medicago t 31 3.1
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	from this gene." /blast_score 9.00e-78 /ec_number /family /chip nova
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---WO 02/22675 - PCT/US01/28506

http://www.ncgr.org/cgi-bin/ff?x91919 (762 letters)

Database: plantfungal 5 661,018 sequences; 426,114,510 total letters Searching......done Score E 10 Sequences producing significant alignments: (bits) Value emb|AJ224519|CAR224519 Cicer arietinum mRNA for LEA protein (clo... 148 6e-35 emb[X15348]BNLEA76 Brassica napus LEA76 mRNA for late embryogene... 135 4e-31 emb|AF117884|AF117884 Glycine max seed maturation protein PM30 (... 88 1e-30 15 emb|AJ224518|CAR224518 Cicer arietimum mRNA for LEA protein (clo... 126 2e-28 emb|AW472097|AW472097 si20a05.yl Gm-c1029 Glycine max cDNA clone... 86 9e-27 emb|AW395529|AW395529 sg72c09.yl Gm-c1007 Glycine max cDNA clone... 88 7e-24 emb|AW507599|AW507599 si53h08.yl Gm-r1030 Glycine max cDNA clone... 88 7e-24 emb|AW318205|AW318205 sg62d04.y1 Gm-c1007 Glycine max cDNA clone... 88 7e-24 20 emb|AW568476|AW568476 si59c06.yl Gm-r1030 Glycine max cDNA clone... 88 7e-24 emb|AW567816|AW567816-si66b10.yl-Gm-r1030 Glycine max cDNA clone... 88 7e-24 emb|AW706800|AW706800 sk03e03.y1 Gm-c1023 Glycine max cDNA clone... 88 1e-23 emb|AW509384|AW509384 si22d10.yl Gm-c1029 Glycine max cDNA clone... 86 2e-23 emb|AW746690|AW746690 WS1\_54\_F07.g1\_A002 Water-stressed 1 (WS1) ... 106 2e-22 25 emb|AW096396|AW096396 EST289576 tomato mixed elicitor, BTI Lycop... 103 1e-21 emb[X78205]HVHVA1 H.vulgare (Himalaya) HVA1 gene. 77 1e-20 emb|X13498|HVABAIP Barley pHVA1 mRNA for an ABA-inducible protein. 77 1e-20 gb[M36000]BLYABA Barley abscisic acid (ABA) mRNA, complete cds. emb|AW680076|AW680076 WS1\_3\_C03.g1\_A002 Water-stressed 1 (WS1) S... 100 2e-20 30 emb|AW397921|AW397921 sg69h06.y1 Gm-c1007 Glycine max cDNA clone... 57 3e-20 emb|AW747095|AW747095 WS1\_65\_C09.g1\_A002 Water-stressed 1 (WS1) ... gb|BE034388|BE034388 MH04B06 MH Mesembryanthemum crystallinum cD... 73 1e-18 emb[X13201|GHLEA7 Cotton set 5A Lea gene for seed protein D-7. emb|X15086|GHLEA29R Cotton set 5A Lea mRNA for seed protein D-29. 35 emb[X56882]TA3LEA Wheat mRNA for a group 3 late embryogenesis ab... 75 1e-18 emb|AW746385|AW746385 WS1\_49\_G03.g1\_A002 Water-stressed 1 (WS1) ... 93 2e-18 emb|AW680063|AW680063 WS1\_3\_B02.g1\_A002 Water-stressed 1 (WS1) S... 91 1e-17 emb|AF255052|AF255052 Triticum aestivum cold-responsive LEA/RAB-... 85 6e-17 emb|AF139915|AF139915 Triticum aestivum ABA-inducible protein WR... 85 6e-17 40 emb|AW164114|AW164114 Ljirnpest20-575-b9 Ljirnp Lambda HybriZap ... 88 7e-17 emb[Y10779]SSY10779 S.stapfianus pSD.42 mRNA. 86 3e-16 emb|AW569002|AW569002 si62c03.y1 Gm-r1030 Glycine max cDNA clone... emblAW508284|AW508284 si52b01.yl Gm-r1030 Glycine max cDNA clone... 62 4e-16 gb|BE034389|BE034389 MH04B07 MH Mesembryanthemum crystallinum cD... 72 7e-16 emb|AW681055|AW681055 WS1 8 F06.b1 A002 Water-stressed 1 (WS1) S... 75 7e-13 emb|AW746839|AW746839 WS1 55 D06.g1 A002 Water-stressed 1 (WS1) ... 75 7e-13 emb|AW678335|AW678335 WS1\_14\_F02.g1\_A002 Water-stressed 1 (WS1) ... 75 7e-13 emb|AW678291|AW678291 WS1 14 D10.g1 A002 Water-stressed 1 (WS1) ... 74 1e-12 emb|AW679525|AW679525 WS1 29 F04.g1 A002 Water-stressed 1 (WS1) ... 74 1e-12 50 emb|Z18891|BPBP8GEN Betula pendula BP8 gene. emb|AW432699|AW432699 sh85b12.y1 Gm-c1016 Glycine max cDNA clone... 74 1e-12 emb|AW678194|AW678194 WS1\_13\_E06.g1\_A002 Water-stressed 1 (WS1) ... 72 5e-12 emb|AW459847|AW459847 sh96a04.y1 Gm-c1016 Glycine max cDNA clone... 70 2e-11 emb[X13203|GHLEA29 Cotton set 5A Lea gene for seed protein D-29. 55 emb|Z49713|PMDORORF6 P.menziesii mRNA (open reading frame) (DF77B). 59 3e-08 emb|Z49712|PMDORORF5 P.menziesii mRNA (open reading frame) (DF77A). 59 3e-08 emb|AJ225460|AJ225460 AJ225460 Abscisic acid-treated protonemata... 59 5e-08 emb[X89041]RFLASPPRO R.fluitans mRNA for landform specific protein. 58 6e-08 gb[M80664|SOYLEAB Soybean late embryogenesis abundant (LEA) prot... 58 9e-08

emb|AF166485 |AF166485 Glycine max maturation protein pPM32 (PM32... 57 2e-07 gb|M19388|COTSPG G.hirsutum (cotton) storage protein (late embry... 57 2e-07

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60 Database: plantfungal

(885 letters)

661,018 sequences; 426,114,510 total letters

Searching....

Score 5 (bits) Value Sequences producing significant alignments: emb|AI486681|AI486681 EST245003 tomato ovary, TAMU Lycopersicon ... 414 e-115 emb|AI488706|AI488706 EST247045 tomato ovary, TAMU Lycopersicon ... 373 e-102 emb|AW185116|AW185116 se87a08.yl Gm-c1023 Glycine max cDNA clone... 364 e-100 10 emb|AW776370|AW776370 EST335435 DSIL Medicago truncatula cDNA cl... 316 1e-85 emb|AW441466|AW441466 EST310862 tomato fruit red ripe, TAMU Lyco... 311 4e-84 emb|AW775237|AW775237 EST331959 GVN Medicago truncatula cDNA clo... 278 4e-74 emb|AW564397|AW564397 LG1 292 F08.b1 A002 Light Grown 1 (LG1) So... 275 2e-73 emb|AW648604|AW648604 EST327154 tomato germinating seedlings, TA... 275 3e-73 15 emb|AW309188|AW309188 sg05d07.y1 Gm-c1019 Glycine max cDNA clone... 274 8e-73 emb|AW329561|AW329561 N200812e rootphos(-) Medicago truncatula c... 204 3e-69 emblAI897460|AI897460 EST266903 tomato ovary, TAMU Lycopersicon ... 158 7e-67 emb|AI775112|AI775112 EST256212 tomato resistant, Cornell Lycope... 248 4e-65 emb|AI898201|AI898201 EST267644 tomato ovary, TAMU Lycopersicon ... 118 1e-51 20 emb|AW011189|AW011189 ST17G07 Pine TriplEx shoot tip library Pin... 172 4e-48 emb|AW622515|AW622515 EST313315 tomato root during/after fruit s... 189 3e-47 emb|AW933718|AW933718 EST359561 tomato fruit mature green, TAMU ... 188 7e-47 emb[AI897919]AI897919 EST267362 tomato ovary, TAMU Lycopersicon ... 102 6e-45 emb|AW623431|AW623431 EST321376 tomato flower buds 3-8 mm, Corne... 179 2e-44 25 emb|A1967352|A1967352 Ljimpest01-065-f5 Ljimp Lambda HybriZap ... 167 5e-44 emb|AW496876|AW496876 ga49e04.yl Moss EST library PPU Physcomitr... 175 5e-43 emb|AW616498|AW616498 EST322909 L. hirsutum trichome, Cornell Un... 174 1e-42 emb|AW687128|AW687128 NF006C08RT1F1065 Developing root Medicago ... 156 1e-41 emb|AW191300|AW191300 T113619e KV2 Medicago truncatula cDNA clon... 167 1e-40 30 emb|AW257178|AW257178 EST305315 KV2 Medicago truncatula cDNA clo... 144 7e-34 emb|AW699388|AW699388 gb07g07.y1 Moss EST library PPN Physcomitr... 139 2e-32 emb|AV421449|AV421449 AV421449 Lotus japonicus young plants (two... 132 4e-30 emblAW626193lAW626193 EST320100 tomato radicle, 5 d post-imbibit... 132 5e-30 emb|AL109608|SPCC23B6 S.pombe chromosome III cosmid c23B6. 35 emblAV426790IAV426790 AV426790 Lotus japonicus young plants (two... 123 2e-27 emb|AI778049|AI778049 EST258928 tomato susceptible, Cornell Lyco... 116 2e-25 emb|AW626687|AW626687 NXNV068E07 Nsf Xylem Normal wood Vertical ... 114 1e-24 emb|AW929355|AW929355 EST338143 tomato flower buds 8 mm to pre-a... 106 2e-22 emb|AW719259|AW719259 LjNEST1h10r Lotus japonicus nodule library... 105 4e-22 40 emb|Z69381|SCCXIV39K S.cerevisiae 38,855 bp segment of chromosom... emb|Z71507|SCYNL231C S.cerevisiae chromosome XIV reading frame O... emb|AB001995|AB001995 Schizosaccharomyces pombe gene for Tellp, ... 82 6e-17 emb|AI442227|AI442227 sa49c11.yl Gm-c1004 Glycine max cDNA clone... 85 8e-16 emb|AI725484|AI725484 BNLGHi12268 Six-day Cotton fiber Gossypium... 54 2e-15 45. emb|X92494|SCBNI1 S.cerevisiae BNI1, N0647, APL1, N0665, N0670, ... 46 5e-13 emb|Z71540|SCYNL264C S.cerevisiae chromosome XIV reading frame O... emb|AI731920|AI731920 BNLGHi11386 Six-day Cotton fiber Gossypium... 44 2e-12 emb|AI729569|AI729569 BNLGHi13678 Six-day Cotton fiber Gossypium... emb|A1728685|A1728685 BNLGHill318 Six-day Cotton fiber Gossypium... 50 emb|AI725979|AI725979 BNLGHi13788 Six-day Cotton fiber Gossypium... emb|AJ273110|AJ273110 AJ273110 Metarhizium anisopliae ARSEF 2575... 52 6e-09 emb|AQ655616|AQ655616 Sheared DNA-2J3.TR Sheared DNA Trypanosoma... 61 1e-08 emb|AQ501933|AQ501933 V11E10 mTn-3xHA/lacZ Insertion Library Sac... 60 2e-08 55 7e-08 emb[X97320]CGSEC14GN C.glabrata SEC14 gene. 55 emb|Z49259|SC9582X S.cerevisiae chromosome XIII cosmid 9582. 55 9e-08 emb|X15483|SCSEC14G Yeast SEC14 gene for cytosolic factor. 55 9e-08 emb|AQ023632|AQ023632 CpGR0082A Cryptosporidium parvum genomic r... 55 7e-07 emb|AQ449556|AQ449556 500001H03.x1 CplOWAM13mp18gDNA1 Cryptospor... 55 7e-07 60 emb|Z28091|SCYKL091C S.cerevisiae chromosome XI reading frame OR... 50 8e-07 emb|A86344|A86344 Sequence 1003 from Patent EP0866129. 54 le-06

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	(2633 letters)
55	Database: plantfungal 661,018 sequences; 426,114,510 total letters
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60	Score E Sequences producing significant alignments: (bits) Value

---WO 02/22675 - PCT/US01/28506

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post/entrez/query?db=n&form=6&dopt=g&uid=gb|al033545|/ncgi

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http://www.ncgr.org/cgi-bin/ff?a1033545 (1140 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

Ε Score

10 Sequences producing significant alignments:

(bits) Value emb|AI507807|AI507807 sb12h09.yl Gm-c1004 Glycine max cDNA clone... 52 7e-08 emb|AW443491|AW443491 EST308421 tomato mixed elicitor, BTI Lycop... 48 2e-07 emb|AW719418|AW719418 LjNEST4c12r Lotus japonicus nodule library... 50 2e-07 gb|BE020862|BE020862 sm53c05.y1 Gm-c1028 Glycine max cDNA clone ... 50 2e-07 emb|AI777049|AI777049 EST252016 tomato callus, TAMU Lycopersicon... 51 2e-07 emb|AW032807|AW032807 EST276366 tomato callus, TAMU Lycopersicon... 51 2e-07 emb|AI776903|AI776903 EST258003 tomato resistant, Cornell Lycope... 47 3e-07 emb|AW092951|AW092951 EST286131 tomato mixed elicitor, BTI Lycop... 47 3e-07 emb|AW203388|AW203388 sf29f06.yl Gm-c1028 Glycine max cDNA clone... 50 3e-07 emb|AW597294|AW597294-si71c07.yl Gm-c1031 Glycine max cDNA clone... 50 3e-07 emb|AW622484|AW622484 EST313272 tomato root during/after fruit s... 47 3e-07 emb|AW707234|AW707234 sk22d10.yl Gm-c1028 Glycine max cDNA clone... 50 3e-07 emb[AW568382]AW568382 si70b03.yl Gm-c1031 Glycine max cDNA clone... 50 3e-07 emb|AW568746|AW568746 si72e11.yl Gm-c1031 Glycine max cDNA clone... 50 3e-07 emb|AV422321|AV422321 AV422321 Lotus japonicus young plants (two... 50 3e-07 emb|AW830420|AW830420 sm26e07.yl Gm-c1028 Glycine max cDNA clone... 50 3e-07 emb|AW650542|AW650542 EST328996 tomato germinating seedlings, TA... 47 3e-07 emb|AJ441446|AJ441446 sa86a06.y1 Gm-c1004 Glycine max cDNA clone... emb[AW234379]AW234379 sf24e05.y1 Gm-c1028 Glycine max cDNA clone... 50 3e-07 emb|AI437875|AI437875 sa40g10.y1 Gm-c1004 Glycine max cDNA clone... 50 3e-07 emb|AV416097|AV416097 AV416097 Lotus japonicus young plants (two... emb|AV413898|AV413898 AV413898 Lotus japonicus young plants (two... 50 3e-07 emb|AI441173|AI441173 sa52c02.y1 Gm-c1004 Glycine max cDNA clone... 50 3e-07 emb|AV415427|AV415427 AV415427 Lotus japonicus young plants (two... 50 3e-07 emb[AI772831]AI772831 EST253931 tomato resistant, Cornell Lycope... 47 3e-07 gb|BE020429|BE020429 sm40f03.y1 Gm-c1028 Glycine max cDNA clone ... 50 3e-07 emb|AI443594|AI443594 sa42b01.yl Gm-c1004 Glycine max cDNA clone... 50 3e-07 emb|AW569805|AW569805 si81g01.y1 Gm-c1031 Glycine max cDNA clone... 50 3e-07 emb|AW093202|AW093202 EST286382 tomato mixed elicitor, BTI Lycop... 46 4e-07 emb|AW906384|AW906384 EST342506 potato-stolon, Cornell Universit... 46 6e-07 emb|AW597346|AW597346 si91f06.yl Gm-c1031 Glycine max cDNA clone... 50 6e-07 emb|AV423170|AV423170 AV423170 Lotus japonicus young plants (two... 50 6e-07 emb|AW351202|AW351202 GM210010B20H11R Gm-r1021 Glycine max cDNA ... 48 8e-07 emb|AW649092|AW649092 EST327546 tomato germinating seedlings, TA... 46 8e-07 emb[AV424931]AV424931 AV424931 Lotus japonicus young plants (two... 48 8e-07 emb[AW350777]AW350777 GM210009B10D3R Gm-r1021 Glycine max cDNA 3... 48 1e-06 emb|AW597304|AW597304 si71d07.yl Gm-c1031 Glycine max cDNA clone... 48 1e-06 emb|AW706307|AW706307 sj54f10.y1 Gm-c1033 Glycine max cDNA clone... 48 1e-06 emb|AW133241|AW133241 se16b08.y1 Gm-c1013 Glycine max cDNA clone... 48 1e-06 emb|AW568935|AW568935 si73c12.y1 Gm-c1031 Glycine max cDNA clone... 48 1e-06 gb|BE020648|BE020648 sm51g03.y1 Gm-c1028 Glycine max cDNA clone ... 48 1e-06 emb|AW567667|AW567667 si77d01.yl Gm-c1031 Glycine max cDNA clone... 48 1e-06 emb|AI495459|AI495459 sa98c07.yl Gm-c1004 Glycine max cDNA clone... 48 1e-06 emb|AW234280|AW234280 sf23c12.yl Gm-c1028 Glycine max cDNA clone... 48 1e-06 emb|AW348755|AW348755 GM210003A22D8R Gm-r1021 Glycine max cDNA 3... 47 2e-06 emb|AW309755|AW309755 sf24d05.x1 Gm-c1028 Glycine max cDNA clone... 47 2e-06 emb|AW703937|AW703937 sk25h11.yl Gm-c1028 Glycine max cDNA clone... 47 2e-06

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PCT/US01/28506 -----WO 02/22675

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60 Database: plantfungal 661,018 sequences; 426,114,510 total letters

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	Score E
5	Sequences producing significant alignments: (bits) Value
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	emb AQ654450 AQ654450 Sheared DNA-25M9.TR Sheared DNA Trypanosom 35 1.9 emb AQ906953 AQ906953 GSSTc02997 Trypanosome cruzi random genomi 35 2.5
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40	gb/BE052627/BE052627 GA_Ea0032J13f Gossypium arboreum 7-10 dpa 34 2.6
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	embly 5280 RTCG APT cruzi can core for algorithm de 2 shochet 22 5 0

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	gb U38804 PPU38804 Porphyra purpurea chloroplast, complete genome. 32 9.4
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50	/chip nova /gb link
	http://www3.ncbi.nlm.nih.gov/htbin-
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	http://www.ncgr.org/cgi-bin/ff?aj011674
35	(2196 letters)
<i></i>	(2190 letters)
	Database: plantfungal
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	Score E
:	Sequences producing significant alignments: (bits) Value
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50	emb AB008191 AB008191 Brassica rapa mRNA for SRK29, complete cds. 333 e-104
50	emb Y18259 BOY18259 Brassica oleracea mRNA for SRK5 protein, par 332 e-104
	emb AB024416 AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 328 e-103
•	gb M76647 BNASKR6A Brassica oleracea receptor protein kinase (SK 329 e-100
	emb AB032473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 321 1e-99
	emb AB012106 AB012106 Brassica rapa mRNA for SRK45, complete cds. 316 2e-98
55	gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 323 2e-98
	gb U00443 BNU00443 Brassica napus cultivar T2 S-receptor kinase 324 4e-98
	emb[Y14286]BOY14286 Brassica oleracea SFR3 gene, partial. 195 4e-97
	gb L08607 BNASRECKIN Brassica napus S-receptor kinase mRNA, comp 312 4e-95
	emb AB013720 AB013720 Brassica oleracea mRNA for SRK23Bol, parti 314 4e-95
60	emb AW620957 AW620957 sj98a07.yl Gm-c1023 Glycine max cDNA clone 285 3e-9
	emb AF088885 AF088885 Nicotiana tabacum receptor-like kinase CHR 306 6e-94
	omor is coccost is coccost incoming anacum receptor-like kingse Cirk 300 06-94

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gb|U20948|ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 221 1e-93 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 270 3e-93 gb[U51741]ITU51741 Ipomoea trifida receptor protein kinase 2 (IR... 264 2e-90 gb|BE034855|BE034855 ML05C11 ML Mesembryanthemum crystallinum cD... 289 2e-90 emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 275 7e-90 dbj|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 274 9e-89 emb[Y12530]BOARLKGEN B.oleraceae gene encoding serine/threonine ... 127 6e-88 dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 272 6e-88 gb[BE057261]BE057261 sm99f12.yl Gm-c1015 Glycine max cDNA clone ... 313 3e-84 10 emb|AJ245480|BNA245480 Brassica napus slg gene for S-locus glyco... 122 2e-82 emb|AW031255|AW031255 EST274630 tomato callus, TAMU Lycopersicon... 302 9e-81 emb[Y12531]BOBRLKGEN B.oleraceae gene encoding serine/threonine ... 128 3e-78 emb|AW760240|AW760240 s159g07.yl Gm-c1027 Glycine max cDNA clone... 283 4e-75 emb|AW831390|AW831390 sm22a06.y1 Gm-c1028 Glycine max cDNA clone... 213 3e-73 emblAB000970lAB000970 Brassica campestris gene for receptor kina... 127 6e-73 15 gb|BE034949|BE034949 ML07F03 ML Mesembryanthemum crystallinum cD... 266 3e-71 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 118 1e-70 emb|Z18921|BOSRKL B.oleracea gene for S-receptor kinase-like pro... 114 4e-69 emb|AI895838|AI895838 EST265281 tomato callus, TAMU Lycopersicon... 261 2e-68 20 emb|AI486193|AI486193 EST244514 tomato ovary, TAMU Lycopersicon ... 182 4e-68 emb|AW442344|AW442344 EST311740 tomato fruit red ripe, TAMU Lyco... 210 8e-68 emb|AB024422|AB024421S2 Brassica oleracea SRK13-b gene, exon 2, ... 110 9e-68 emb|AB000971|AB000971 Brassica campestris pseudogene for recepto... 111 2e-67 emb|AB024420|AB024419S2 Brassica oleracea SRK13 gene, exon 2, 3,... 109 2e-67 25 emb|AI896155|AI896155 EST265598 tomato callus, TAMU Lycopersicon... 186 3e-67 emb|AB013718|AB013717S2 Brassica rapa gene for SRK46Bra, exon 2,... 109 3e-66 emb|Z30211|BOSRK29G B.oleracea (alboglabra) srk29 gene. 120 2e-65 dbj|D88193|D88193 Brassica rapa DNA for S-receptor kinase, compl... 120 3e-65 emb|X79432|BOSRK3 B.oleracea SRK3 gene. 116 le-64 30 emb|AW203661|AW203661 sf36g06.y1 Gm-c1028 Glycine max cDNA clone... 208 2e-63 emb|AW736407|AW736407 EST332421 KV3 Medicago truncatula cDNA clo... 241 2e-62 emb|AI967314|AI967314 Ljirnpest00-017 Ljirnp Lambda HybriZap two... 134 1e-60 emb|AI897876|AI897876 EST267319 tomato ovary, TAMU Lycopersicon ... 171 3e-60 emb|AW220676|AW220676 EST297145 tomato fruit mature green, TAMU ... 231 2e-59 35 emb|AW220675|AW220675 EST297144 tomato fruit mature green, TAMU ... 231 2e-59 emb|AI486584|AI486584 EST244905 tomato ovary, TAMU Lycopersicon ... 171 3e-59 emblAW031816|AW031816 EST275270 tomato callus, TAMU Lycopersicon... 136 1e-58 emb|AW220677|AW220677 EST297146 tomato fruit mature green, TAMU ... 227 2e-58 emb|AW309544|AW309544 sf20h08.x1 Gm-c1028 Glycine max cDNA clone... 167 4e-57 40 emb|AW666141|AW666141 sk32f11.y1 Gm-c1028 Glycine max cDNA clone... 176 7e-56 emb[AW033458[AW033458 EST277029 tomato callus, TAMU Lycopersicon... 219 8e-56 emb|AI901283|AI901283 sc31d08.yl Gm-c1014 Glycine max cDNA clone... 175 9e-56 emblY16999|TCA16999 Theobroma cacao microsatellite DNA, clone mT... 131 1e-55 emb|AI822907|AI822907 L30-754T3 Ice plant Lambda Uni-Zap XR expr... 117 1e-53 45 emb|AI822355|AI822355 LO-804T3 Ice plant Lambda Uni-Zap XR expre... 116 4e-53 emb|AW667985|AW667985 GA\_Ea0012C15 Gossypium arboreum 7-10 dpa ... 111 7e-53 emb|AW684339|AW684339 NF015G04NR1F1000 Nodulated root Medicago t... 140 9e-53 emb|AB041503|AB041503 Populus nigra PnPK1 mRNA for protein kinas... 78 4e-52 emb|A1899009|A1899009 EST268452 tomato ovary, TAMU Lycopersicon ... 160 5e-52 50 emb|AW034624|AW034624 EST278308 tomato callus, TAMU Lycopersicon... 205 1e-51 emblAI771857|AI771857 EST252957 tomato ovary, TAMU Lycopersicon ... 156 6e-51 emb|AI488101|AI488101 EST246423 tomato ovary, TAMU Lycopersicon ... 156 6e-51 emb|AI898581|AI898581 EST268024 tomato ovary, TAMU Lycopersicon ... 160 6e-51 emblAW039406|AW039406 EST281663 tomato mixed elicitor, BTI Lycop... 115 1e-50 55 emb|AI896953|AI896953 EST266396 tomato callus, TAMU Lycopersicon... 183 1e-50 emb|AI899156|AI899156 EST268599 tomato ovary, TAMU Lycopersicon ... 160 3e-50 emb|Z18884|BOSRKRPC B.oleracea encoding S-receptor kinase relate... 125 2e-49 emb|AI895623|AI895623 EST265066 tomato callus, TAMU Lycopersicon... 196 5e-49 emb|AW054349|AW054349 L30-1774T3 Ice plant Lambda Uni-Zap XR exp... 116 6e-49 60 emb|AW687233|AW687233 NF007D09RT1F1077 Developing root Medicago ... 100 8e-49 emb|AF142596|AF142596 Nicotiana tabacum LRR receptor-like protei... 74 4e-48

emb|AI822347|AI822347 L0-796T3 Ice plant Lambda Uni-Zap XR expre... 116 5e-47 emb[AW776492]AW776492 EST335557 DSIL Medicago truncatula cDNA cl... 189 8e-47 emb|AI486331|AI486331 EST244652 tomato ovary, TAMU Lycopersicon ... 160 9e-47 emb|AW706972|AW706972 sk20a03.yl Gm-c1028 Glycine max cDNA clone... 188 1e-46 5 gb[U51330]TAU51330 Triticum aestivum leaf rust resistance kinase... 78 1e-46 emb|AI967315|AI967315 Ljirnpest00-018 Ljirnp Lambda HybriZap two... 142 2e-46 emb|AW279355|AW279355 sf65g10.yl Gm-c1013 Glycine max cDNA clone... 111 4e-46 emblAI729170|AI729170 BNLGHi12834 Six-day Cotton fiber Gossypium... 130 5e-46 emb|AI775997|AI775997 EST257097 tomato resistant, Cornell Lycope... 186 5e-46 10 emb|AI930642|AI930642 sb37f05.yl Gm-c1013 Glycine max cDNA clone... 182 1e-44 emb|AF085164|AF085164 Hordeum vulgare receptor-like kinase LRK10... 78 1e-44 emb|AI731504|AI731504 BNLGHi9991 Six-day Cotton fiber Gossypium ... 67 3e-44 emb|AW394449|AW394449 sh05d09.yl Gm-c1016 Glycine max cDNA clone... 119 4e-44 emb[AI938169]AI938169 sc40d07.y1 Gm-c1014 Glycine max cDNA clone... 179 6e-44 15 emb|Z18861|BOSRKRPA B.oleracea encoding S-receptor kinase relate... 114 7e-44 emb|AB041504|AB041504 Populus nigra PnPK2 mRNA for protein kinas... 76 1e-43 emb|AF197947|AF197947 Glycine max receptor protein kinase-like p... 125 1e-43 emb[Y14600]SBRLK1 Sorghum bicolor mRNA for protein serine/threon... 74 1e-43 emb|AW687052|AW687052 NF005E02RT1F1017 Developing root Medicago ... 177 2e-43 20 emb|AI055189|AI055189 coau0003E19 Cotton Boll Abscission Zone cD... 107 4e-43 \_\_\_gb|BE058691|BE058691 sn19e05.yl Gm-c1016 Glycine max cDNA clone ... 92 4e-43 emb|Z18883|BOSRKRPB B.oleracea encoding S-receptor kinase relate... 176 4e-43

Query= AC002392.162 at 12307 at /id source genbank /description 25 gb|aad12037.1| (ac002392) putative receptor-like protein kinase [arabidopsis thaliana] /blast\_score 0 /ec\_mmber /family kinase /chip nova/gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|ac002392| /ncgi 30 http://www.ncgr.org/cgi-bin/ff?ac002392 (2631 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

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Score E

Sequences producing significant alignments: (bits) Value emb|AW774582|AW774582 EST333733 KV3 Medicago truncatula cDNA clo... 204 5e-74 emb|AI729170|AI729170 BNLGHi12834 Six-day Cotton fiber Gossypium... 245 1e-63 emb|Z73295|CRPK1 C.roseus mRNA for receptor-like protein kinase. 146 5e-56 emb|AF220602|AF220602 Lycopersicon pimpinellifolium Rio Grande 7... 112 7e-54 gb U59317 LPU59317 Lycopersicon pimpinellifolium serine/threonin... 108 7e-54 gb U13923 LEU13923 Lycopersicon pimpinellifolium serine/threonin... 108 7e-54 emb|AW222552|AW222552 EST299363 tomato fruit red ripe, TAMU Lyco... 210 3e-53 gb|BE053916|BE053916 GA Ea0031D03f Gossypium arboreum 7-10 dpa ... 209 8e-53 emb|AF220603|AF220603 Lycopersicon esculentum VFNT Cherry Pto lo... 112 4e-52 emb|AW200786|AW200786 se93e06.yl Gm-c1027 Glycine max cDNA clone... 206 5e-52 emb|AW729859|AW729859 GA\_Ea0026H04 Gossypium arboreum 7-10 dpa ... 206 7e-52 emb|AW774672|AW774672 EST333823 KV3 Medicago truncatula cDNA clo... 82 7e-51 emb|AW667985|AW667985 GA\_Ea0012C15 Gossypium arboreum 7-10 dpa ... 158 2e-50 gb|U59316|LEU59316 Lycopersicon esculentum serine/threonine prot... 111 9e-50 gb[U59318]LEU59318 Lycopersicon esculentum serine/threonine prot... 102 3e-49 emb|AF121450|AF121450 Capsicum annuum protein kinase homolog C11... 104 1e-48 emb|AF108892|AF108892 AF108892 Capsicum annuum root 1st-branched... 194 3e-48 emb|AI967314|AI967314 Ljirnpest00-017 Ljirnp Lambda HybriZap two... 129 5e-48

gb|U59315|LPU59315 Lycopersicon pimpinellifolium serine/threonin... 103 1e-47 60 gb [U02271 LEU02271 Lycopersicon pimpinellifolium Rio Grande-PtoR... 103 1e-47 emb|AF197947|AF197947 Glycine max receptor protein kinase-like p... 185 3e-47

emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 113 5e-47 emb|AF197946|AF197946 Glycine max receptor protein kinase-like p... 182 2e-46 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 112 2e-46 emb|AW773915|AW773915 EST332901 KV3 Medicago truncatula cDNA clo... 82 3e-46 emb|AF053127|AF053127 Malus domestica leucine-rich receptor-like... 130 4e-46 dbj[D38564]BOLRPKB Brassica campestris mRNA for receptor protein... 110 4e-46 emb|AB012106|AB012106 Brassica rapa mRNA for SRK45, complete cds. 111 5e-46 dbilD38563|BOLRPKA Brassica campestris mRNA for receptor protein... 109 6e-46 emb|AW982539|AW982539 HVSMEg0003I16f Hordeum vulgare pre-anthesi... 146 6e-46 10 emb|AI736063|AI736063 sb22d04.yl Gm-c1007 Glycine max cDNA clone... 147 6e-46 emb|AW256717|AW256717 EST304854 KV2 Medicago truncatula cDNA clo... 82 7e-46 dbj|D31737|TOBPSTK Tobacco mRNA for protein-serine/threonine kin... 94 2e-45 dbj[E05046|E05046 DNA encoding ZmPK1 homologue protein in tobacco. 94 2e-45 emb|AW979740|AW979740 EST341365 tomato root deficiency, Cornell ... 184 2e-45 15 emb|AI895365|AI895365 EST264808 tomato callus, TAMU Lycopersicon... 184 2e-45 emb[Y14600]SBRLK1 Sorghum bicolor mRNA for protein serine/threon... 82 3e-45 gb|U28007|LEU28007 Lycopersicon esculentum Pto kinase interactor... 127 1e-44 emb|A67797|A67797 Sequence 2 from Patent WO9743427. 79 2e-44 gb|U93048|DCU93048 Daucus carota somatic embryogenesis receptor-... 79 2e-44 20 emb|AF121451|AF121451 Capsicum annuum protein kinase homolog C11... 105 5e-44 emb|AB041503|AB041503 Populus nigra PnPK1 mRNA for protein kinas... 133 6e-44 emb|AW621923|AW621923 EST312721 tomato root during/after fruit s... 81 2e-43 emb|AA738545|AA738545 SbRLK3 Sorghum bicolor cv. TX430 leaf Sorg... 78 3e-43 gb[U20948]ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 99 1e-42 25 emb|AF121449|AF121449 Capsicum annuum protein kinase homolog C15... 99 1e-42 emb|AW219672|AW219672 EST302154 tomato root during/after fruit s... 125 1e-42 emb[AW030530]AW030530 EST273785 tomato callus, TAMU Lycopersicon... 162 1e-42 emb|AI898917|AI898917 EST268360 tomato ovary, TAMU Lycopersicon ... 122 1e-42 emb|AI489287|AI489287 EST247626 tomato ovary, TAMU Lycopersicon ... 122 2e-42 30 emb|AI730776|AI730776 BNLGHi7867 Six-day Cotton fiber Gossypium ... 151 2e-42 emb|AA738544|AA738544 SbRLK2 Sorghum bicolor cv. TX430 leaf Sorg... 78 2e-42 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 115 6e-42 gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 110 9e-42 emb|AB041504|AB041504 Populus nigra PnPK2 mRNA for protein kinas... 132 9e-42 35 gb[M97667]BNASTKR Brassica napus ssp. oleifera serine/threonine ... 115 2e-41 emb|AW036763|AW036763 EST252152 tomato ovary, TAMU Lycopersicon ... 164 2e-41 emb|AW036762|AW036762 EST252151 tomato ovary, TAMU Lycopersicon ... 164 2e-41 emb|AW668493|AW668493 GA\_Ea0014C20 Gossypium arboreum 7-10 dpa ... 127 4e-41 gb|U00443|BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 110 4e-41 40 emb|AW776704|AW776704 EST335769 DSIL Medicago truncatula cDNA cl... 113 2e-40 emb|AF131222|AF131222 Lophopyrum elongatum protein serine/threon... 97 2e-40 emb|AI484701|AI484701 EST242962 tomato ovary, TAMU Lycopersicon ... 133 2e-40 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 111 3e-40 emb[AW031816]AW031816 EST275270 tomato callus, TAMU Lycopersicon... 105 3e-40 emb|AW615998|AW615998 EST325448 tomato flower buds 0-3 mm, Corne... 117 3e-40 emb|AW218745|AW218745 EST301225 tomato root during/after fruit s... 120 5e-40 emb|AI731501|AI731501 BNLGHi9983 Six-day Cotton fiber Gossypium ... 98 6e-40 gb|M76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 108 9e-40 emb|AF121448|AF121448 Capsicum annuum protein kinase homolog C11... 95 2e-39 50 emb|AI771280|AI771280 EST252296 tomato ovary, TAMU Lycopersicon ... 165 2e-39 emb|AI486547|AI486547 EST244868 tomato ovary, TAMU Lycopersicon ... 123 2e-39 emblAW094468|AW094468 EST287648 tomato mixed elicitor, BTI Lycop... 156 2e-39 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. 108 3e-39 emb|AI771841|AI771841 EST252941 tomato ovary, TAMU Lycopersicon ... 164 4e-39 55 emb|AI485090|AI485090 EST243394 tomato ovary, TAMU Lycopersicon ... 164 4e-39 emb|A1731504|A1731504 BNLGHi9991 Six-day Cotton fiber Gossypium ... 78 4e-39 emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 111 4e-39 emb|Y18260|BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 107 6e-39 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 107 6e-39 60 emb|AW267961|AW267961 EST306183 DSIR Medicago truncatula cDNA cl... 94 7e-39 emb[Y18259]BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 107 1e-38

emb|AI486766|AI486766 EST245088 tomato ovary, TAMU Lycopersicon ... 89 1e-38 emb|AJ485862|AJ485862 EST244183 tomato ovary, TAMU Lycopersicon ... 162 1e-38 emb|AI487456|AI487456 EST245778 tomato ovary, TAMU Lycopersicon ... 123 2e-38 gb|BE060160|BE060160 HVSMEg0011B12f Hordeum vulgare pre-anthesis... 75 2e-38 emb|X98520|BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 107 3e-38 emb|AF142596|AF142596 Nicotiana tabacum LRR receptor-like protei... 78 3e-38 emb|AW617255|AW617255 EST323666 L. hirsutum trichome, Cornell Un... 148 4e-38 gb[U51741]ITU51741 Ipomoea trifida receptor protein kinase 2 (IR... 71 4e-38 emb|AW774790|AW774790 EST333941 KV3 Medicago truncatula cDNA clo... 95 5e-38 10 emb[Y14286]BOY14286 Brassica oleracea SFR3 gene, partial. 105 5e-38 emb|AI898390|AI898390 EST267833 tomato ovary, TAMU Lycopersicon ... 121 1e-37 gb|BE020963|BE020963 sm54a06.y1 Gm-c1028 Glycine max cDNA clone ... 125 3e-37 emblAI730535lAI730535 BNLGHi7007 Six-day Cotton fiber Gossypium ... 73 5e-37 15 emb|AF085167|AF085167 Hordeum vulgare receptor-like kinase ARK1A... 81 7e-37 emb|AW687267|AW687267 NF007G07RT1F1055 Developing root Medicago ... 140 8e-37 emb|AW648736|AW648736 EST327106 tomato germinating seedlings, TA... 98 1e-36 emb|AW650851|AW650851 EST329305 tomato germinating seedlings, TA... 126 2e-36 20 Query= AB023448.2 s at 12332 s at /id source genbank /description

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

30 Score

35

40

45

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Sequences producing significant alignments:

(bits) Value

E

emblAF135130|AF135130 Arabis holboellii from Denmark class I chi... 438 0.0 emblX16939INTECHITR Nicotiana tabacum mRNA for endochitinase (EC... 491 e-158 gblM15173|TOBECH Tobacco (N.tabacum) endochitinase mRNA, partial... 494 e-158 gb|S44869|S44869 basic chitinase [Nicotiana tabacum=tobacco, cv ... 490 e-158 emb|A16119|A16119 Intracellular chitinase mRNA (SEQ ID NO: 2). 493 e-158 emb[X15494]STCHITIN Potato endochitinase gene (EC 3.2.1.14). 541 e-153 emb[X07130|STCHIT Solanum tuberosum mRNA for endochitinase (EC 3... 541 e-153 emb[Y10373]MTCHITIN1 M.truncatula mRNA for chitinase. 482 e-151 gb|U02606|STU02606 Solanum tuberosum chitinase (chtB2) mRNA, par... 442 e-150 gb[U02605|STU02605 Solanum tuberosum chitinase (chtB1) mRNA, par... 443 e-150 emb|Z15140|LECHI9 L.esculentum mRNA for chitinase. 442 e-149 gb|U02607|STU02607 Solanum tuberosum chitinase (chtB3) mRNA, par... 440 e-148 emb|Z78202|PACHI1 Persea americana mRNA for endochitinase. 467 e-148 gb[U83592[MSU83592 Medicago sativa class I chitinase mRNA, compl... 310 e-146 gb[U83591]MSU83591 Medicago sativa class I chitinase mRNA, compl... 310 e-146 gb|L37876|PEACH12I Pisum sativum chitinase class I (chi2) gene, ... 309 e-146 emb[AF000966]AF000966 Poa pratensis chitinase (Chi2) gene, compl... 472 e-146 emb|Z54234|VVCHIT1MR V.vinifera mRNA for chitinase. gb|M13968|PHVCHM P.vulgaris chitinase mRNA, complete cds. 286 e-145 emb|AJ012821|CAR012821 Cicer arietinum mRNA for class I chitinase. 305 e-144

emb|X76041|TACHIG T.aestivum (Chinese spring) chi gene for endoc... 466 e-144 gb|U78888|GHU78888 Gossypium hirsutum class I endochitinase mRNA... 332 e-144 emb|AF034566|AF034566 Gossypium hirsutum class I chitinase mRNA,... 333 e-144 emb|AF000965|AF000965 Poa pratensis chitinase (Chi3) pseudogene ... 469 e-144 emb|X14133|STENCHIT Potato mRNA fragment for endochitinase (EC 3... 435 e-143 gb|S43926|S43926 CH5B=chitinase [Phaseolus vulgaris=beans, cv Sa... 284 e-143 emb|X88800|VURNACHI1 V.unguiculata mRNA for chitinase clase 1 (p... 296 e-143

60 gb|U02287|HVU02287 Hordeum vulgare cultivar NK1558 chitinase gen... 459 e-141 gb|M94106|ALCCHINTIA Allium sativum chitinase mRNA, 3' end. 455 e-140

	golivisatios attivum chitinase mkna, 3' end. 448 e-140
	emb AB015655 AB015655 Cucurbita sp. mRNA for chitinase, complete 286 e-139
	emb AF000964 AF000964 Poa pratensis chitinase (Chi1) gene, compl 408 e-139
	gb L34211 BLYCHI33A Hordeum vulgare chitinase (CHI33) gene, comp 448 e-138
5	gb[U02608 STU02608 Solanum tuberosum chitinase (chtB4) mRNA, par 408 e-137
	emb[X63899]PSCHITIN P.sativum mRNA for chitinase. 270 e-135
	emb AF061805 AF061805 Elaeagnus umbellata acidic chitinase mRNA, 430 e-134
	emb AF202731 AF202731 Glycine max endochitinase homolog (Chn1) m 273 e-134
10	gb L34210 BLYCHI26A Hordeum vulgare chitinase (CHI26) gene, comp 475 e-133
10	emb A37990 A37990 Sequence 9 from Patent EP0616035. 475 e-133
	gb[M62904]BLYCHI H.vulgare L. 26kD chitinase mRNA, complete cds. 475 e-133
	emb AF098302 AF098302 Brassica juncea chitinase mRNA, complete cds. 265 e-132
	gb U30324 TCU30324 Theobroma cacao class I chitinase gene, compl 238 e-131
	emb X95610 CSHITTB C.sativa mRNA for chitinase lb. 259 e-130
15	gb U48687 CSU48687 Castanea sativa endochitinase mRNA, complete 259 e-130
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	emb AF135137 AF135137 Arabis fecunda country USA class I chitina 459 e-128
	emb AF061806 AF061806 Elaeagnus umbellata basic chitinase mRNA, 297 e-128
	emb AB023464 AB023464 Arabis gemmifera ChiB gene for basic endoc 455 e-127
20	emb AF135152 AF135152 Arabis parishii country USA class I chitin 455 e-127
20	cinopar 133132 par 133132 Arabis parisini country USA class I chiun 435 e-127
	emb AF135141 AF135141 Arabis gunnisoniana class I chitinase gene 455 e-127
	emb AF135135 AF135135-Arabis drummondii class I chitinase gene, 455 e-127
	emb AF135143 AF135143 Arabis lemmonii country USA class I chitin 452 e-126
	emb X67693 STMREN S.tuberosum mRNA for endochitinase. 452 e-126
25	emb AF135132 AF135132 Arabis gunnisoniana from USA class I chiti 452 e-126
	emb AF135144 AF135144 Arabis lemmonii country USA class I chitin 451 e-126
	emb AF135140 AF135140 Arabis glabra country USA class I chitinas 449 e-125
	gb[U30465]LEU30465 Lycopersicon esculentum class II chitinase (C 447 e-125
	emb AF135153 AF135153 Arabis parishii country USA class I chitin 445 e-124
30	gb L22032 ULMCHITIN Ulmus americana chitinase (pHS2) mRNA, compl 295 e-124
	emb AF135148 AF135148 Arabis lyallii class I chitinase gene, par 444 e-124
	emb AF135151 AF135151 Arabis microphylla country USA class I chi 444 e-124
	emb AF135150 AF135150 Arabis microphylla country USA class I chi 443 e-124
	emb AF135147 AF135147 Arabis lignifera country USA class I chiti 443 e-124
35	emb AF135136 AF135136 Arabis fecunda country USA class I chitina 443 e-124
55	emb AF135145 AF135145 Arabis lignifera country USA class I chiti 443 e-123
	emb AF135146 AF135146 Arabis lignifera country USA class I chiti 441 e-123
	emb AF135149 AF135149 Arabis microphylla class I chitinase gene, 441 e-123
40	emb AF043247 AF043247 Solanum tuberosum class I chitinase (ChtC1 431 e-122
40	emb AF043248 AF043248 Solanum tuberosum class I chitinase (ChtC2 431 e-122
	emb Z15138 LECHI14 L.esculentum mRNA for chitinase (partial). 439 e-122
	emb AF135142 AF135142 Halimolobos perplexa var. perplexa class I 435 e-121
	emb AF135134 AF135134 Arabis blepharophylla class I chitinase ge 434 e-121
	gb[U01660]U01660 Populus trichocarpa x Populus deltoides acidic 207 e-121
45	emb AF135138 AF135138 Arabis glabra country USA class I chitinas 431 e-120
	emb AF135133 AF135133 Arabis blepharophylla country USA class I 423 e-118
	gb M95835 BNACH25A Brassica napus (clone BnCh25) endochitinase g 422 e-117
	emb AW034530 AW034530 EST278146 tomato callus, TAMU Lycopersicon 418 e-116
	emb AW560048 AW560048 EST315096 DSIR Medicago truncatula cDNA cl 350 e-112
50	emb AW687771 AW687771 NF013C08RT1F1065 Developing root Medicago 311 e-11
-	emb AW033115 AW033115 EST276674 tomato callus, TAMU Lycopersicon 391 e-108
	emb AW034645 AW034645 EST278376 tomato callus, TAMU Lycopersicon 366 e-106
	emb AF082713 AF082713 AF082713 Capsicum annuum leaf mRNA Capsicu 364 e-106
<i></i>	emb AW738053 AW738053 EST339480 tomato flower buds, anthesis, Co 379 e-104
55	emb AF141373 AF141373 Petroselinum crispum chitinase precursor ( 237 e-104
	emb Z70032 CSACHIT2 C.sinensis mRNA for class II acidic chitinase. 197 e-104
	emb AW030745 AW030745 EST274000 tomato callus, TAMU Lycopersicon 360 e-104
	emb AF141374 AF141374 Petroselinum crispum chitinase precursor ( 237 e-104
	emb AW033034 AW033034 EST276593 tomato callus, TAMU Lycopersicon 367 e-103
60	emb AW216454 AW216454 EST295084 tomato callus, TAMU Lycopersicon 373 e-102
	embla W267781   A W267781   FST305909 DSTR Medicago truncatula cDNA cl. 216 e 102

emb|AW033757|AW033757 EST277328 tomato callus, TAMU Lycopersicon... 285 e-101 emb|AW037673|AW037673 EST279276 tomato mixed elicitor, BTI Lycop... 281 e-100 emb|AW216787|AW216787 EST295501 tomato callus, TAMU Lycopersicon... 364 e-100 emb|AI771248|AI771248 EST252264 tomato ovary, TAMU Lycopersicon... 363 1e-99 emb|AW031102|AW031102 EST274409 tomato callus, TAMU Lycopersicon... 275 1e-98 emb|X15349|HVENDCHT Barley (H.vulgare) mRNA for endochitinase. 359 2e-98 emb|AW622028|AW622028 EST312826 tomato root during/after fruit s... 269 9e-97 emb|Z15139|LECHI17 L.esculentum mRNA for chitinase. 242 2e-96 emb|AW032161|AW032161 EST275615 tomato callus, TAMU Lycopersicon... 334 2e-96 emb|AW560177|AW560177 EST315225 DSIR Medicago truncatula cDNA cl... 295 1e-95

Query= AL021637.176\_s\_at 12341\_s\_at /id\_source genbank /description emb|caa16619.1| (al021637) vacuolar sorting receptor-like protein [arabidopsis thaliana] /blast\_score 0 /ec\_number /family /chip nova /gb\_link /ncgi
(1881 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

20

60

15

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\_Searching......done

Score E.

Sequences producing significant alignments:

(bits) Value

25 gb[U79958]PSU79958 Pisum sativum BP-80 vacuolar sorting receptor... 810 0.0 emb|AB006809|AB006809 Cucurbita sp. mRNA for PV72, complete cds. 780 0.0 emb|AW267745|AW267745 EST305873 DSIR Medicago truncatula cDNA cl... 453 e-126 emb|AW931583|AW931583 EST357426 tomato fruit mature green, TAMU ... 308 e-112 30 emb|AW309187|AW309187 sg05d06.yl Gm-c1019 Glycine max cDNA clone... 405 e-112 gb[BE054150]BE054150 GA Ea0034H17f Gossypium arboreum 7-10 dpa ... 398 e-110 emb|AW622833|AW622833 EST306903 tomato flower buds 3-8 mm, Corne... 387 e-106 emb|AW689392|AW689392 NF018F12ST1F1000 Developing stem Medicago ... 371 e-102 emb|AW737948|AW737948 EST339375 tomato flower buds, anthesis, Co... 342 3e-94 35 emb|AW774434|AW774434 EST333585 KV3 Medicago truncatula cDNA clo... 263 2e-90 emb|AW932529|AW932529 EST358372 tomato fruit mature green, TAMU ... 326 3e-88 emb|AW615949|AW615949 EST325315 tomato flower buds 0-3 mm, Corne... 216 1e-87 emb|AI728635|AI728635 BNLGHi11276 Six-day Cotton fiber Gossypium... 281 8e-84 emb|AI782787|AI782787 EST263666 tomato susceptible, Cornell Lyco... 307 2e-82 40 emb|AI443067|AI443067 sa47a01.y1 Gm-c1004 Glycine max cDNA clone... 305 6e-82 emb|AW747297|AW747297 WS1\_67\_G06.b1\_A002 Water-stressed 1 (WS1) ... 292\_8e-78 emb|AI727826|AI727826 BNLGHi9195 Six-day Cotton fiber Gossypium ... 280 1e-77 emb|AI484571|AI484571 EST242801 tomato ovary, TAMU Lycopersicon ... 284 1e-75 emb|AW685785|AW685785 NF030C07NR1F1000 Nodulated root Medicago t... 166 1e-71 45 emb|AV406766|AV406766 AV406766 Lotus japonicus young plants (two... 260 2e-68 emb|AW509740|AW509740 ga63h11.yl Moss EST library PPU Physcomitr... 232 1e-65 emb|AV428420|AV428420 AV428420 Lotus japonicus young plants (two... 249 4e-65 emb|AW695542|AW695542 NF096C05ST1F1037 Developing stem Medicago ... 124 1e-58 emb|AW289687|AW289687 NXNV004E04F Nsf Xylem Normal wood Vertical... 226 4e-58 50 emb|AW064744|AW064744 ST35C06 Pine TriplEx shoot tip library Pin... 198 2e-56 emb|AW309191|AW309191 sg05d10.yl Gm-c1019 Glycine max cDNA clone... 171 3e-52 emb|AW096632|AW096632 EST289812 tomato mixed elicitor, BTI Lycop... 165 7e-52 emb|AW568619|AW568619 si60a11.yl Gm-r1030 Glycine max cDNA clone... 205 1e-51 emb|AF209910|AF209910 Prunus dulcis vacuolar sorting receptor pr... 160 5e-51 55 emb|AW623959|AW623959 EST321904 tomato flower buds 3-8 mm, Come... 75 1e-50 emb|AW397829|AW397829 sg68h03.yl Gm-c1007 Glycine max cDNA clone... 201 2e-50 emb|AW256542|AW256542 EST304679 KV2 Medicago truncatula cDNA clo... 151 2e-50

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emb|AA660289|AA660289 00158 MtRHE Medicago truncatula cDNA 5', m... 107 9e-44 emb|AW056624|AW056624 ST53G07 Pine TriplEx shoot tip library Pin... 178 1e-43

emblAW690002|AW690002 NF026G04ST1F1000 Developing stem Medicago ... 109 2e-43 emb|AW290400|AW290400 NXNV019G06F Nsf Xylem Normal wood Vertical... 177 3e-43 emb|AI162330|AI162330 A016P01U Hybrid aspen plasmid library Popu... 131 2e-39 emb|AI161766|AI161766 A006P54U Hybrid aspen plasmid library Popu... 93 3e-38 emb|AW737369|AW737369 EST338892 tomato flower buds, anthesis, Co... 149 5e-35 gb|BE125908|BE125908 DG1 59 E01.b1 A002 Dark Grown I (DG1) Sorgh... 149 5e-35 emb|AW201441|AW201441 sf03b09.yl Gm-c1027 Glycine max cDNA clone... 113 1e-34 emblAW398931 AW398931 EST309431 L. pennellii trichome, Cornell U... 95 2e-34 gb|BE049814|BE049814 NXNV 144 F04 F Nsf Xylem Normal wood Vertic... 143 6e-33 10 emb|AW126100|AW126100 N100297e rootphos(-) Medicago truncatula c... 100 5e-30 emb|AW508719|AW508719 si35f03.y1 Gm-r1030 Glycine max cDNA clone... 125 1e-27 gb|L38113|L38113 BNAF0628E Mustard flower buds Brassica rapa cDN... 117 4e-26 emb[AW317388]AW317388 sg48g10.y1 Gm-c1025 Glycine max cDNA clone... 80 1e-25 emb|AW706755|AW706755 sk02f10.yl Gm-c1023 Glycine max cDNA clone... 118 2e-25 15 emb|AA660955|AA660955 00852 MtRHE Medicago truncatula cDNA 5' si... 74 7e-25 emb|AW616461|AW616461 EST322872 L. hirsutum trichome, Cornell Un... 69 2e-24 emb|AW348825|AW348825 GM210003B11E6R Gm-r1021 Glycine max cDNA 3... 68 8e-24 emb|AW932524|AW932524 EST358367 tomato fruit mature green, TAMU ... 80 1e-23 emb|AW774190|AW774190 EST333273 KV3 Medicago truncatula cDNA clo... 101 2e-20 20 emb|AW125944|AW125944 N100139e rootphos(-) Medicago truncatula c... 66 6e-20 emb|AW093844|AW093844 EST287024 tomato mixed elicitor, BTI Lycop... 96 7e-19 emb|AW906386|AW906386 EST342508 potato stolon, Cornell Universit... 67 2e-16 gb|BE060808|BE060808 HVSMEg0013H06f Hordeum vulgare pre-anthesis... 87 4e-16 emblAW620693|AW620693 sj08d10.yl Gm-c1032 Glycine max cDNA clone... 76 1e-15 25 emblAW256398|AW256398 EST304465 KV2 Medicago truncatula cDNA clo... 74 6e-14 emb|AW037563|AW037563 EST278890 tomato mixed elicitor, BTI Lycop... 79 8e-14 emb|AW691052|AW691052 NF037B05ST1F1000 Developing stem Medicago ... 72 4e-13 emb|AW686287|AW686287 NF040A08NR1F1000 Nodulated root Medicago t... 77 4e-13 emb|AW685952|AW685952 NF036G09NR1F1000 Nodulated root Medicago t... 46 2e-11 30 emb|AW760128|AW760128 s158d09.y1 Gm-c1027 Glycine max cDNA clone... 58 1e-10 emb|AW119909|AW119909 sd54d08.yl Gm-c1016 Glycine max cDNA clone... 66 8e-10 emb|AI794754|AI794754 sb68f11.y1 Gm-c1019 Glycine max cDNA clone... 64 4e-09 emb|AW774852|AW774852 EST334003 KV3 Medicago truncatula cDNA clo... 64 4e-09 emb|AW127457|AW127457 M110648 DSIL Medicago truncatula cDNA clon... 58 3e-07 35 emb|AW443352|AW443352 EST308282 tomato mixed elicitor, BTI Lycop... 56 1e-06 emb|AW747372|AW747372 WS1 67\_G06.g1 A002 Water-stressed 1 (WS1) ... 55 2e-06 emb|AI939286|AI939286 sc69h02.y1 Gm-c1016 Glycine max cDNA clone... 46 0.001 emb|AF198615|AF198615 Neospora caninum microneme protein Nc-P38 ... 43 0.009 emb|AI822258|AI822258 L0-701T3 Ice plant Lambda Uni-Zap XR expre... 36 0.019 40 gbBE035779BE035779 MO16H05 MO Mesembryanthemum crystallinum cD... 36 0.025 emb|AI822869|AI822869 L30-712T3 Ice plant Lambda Uni-Zap XR expr... 36 0.025 emb|AI822754|AI822754 LO-1287T3 Ice plant Lambda Uni-Zap XR expr... 36 0.025 emb|AI756736|AI756736 EtESTea24h07.yl Eimeria S5-2 Sporozoite st... 34 0.035 gb[M36941]BLYHORDCA Hordeum vulgare C-hordein gene, complete cds. 45 emb|AU036628|AU036628 Schizosaccharomyces pombe genomic clone ha... 40 0.083 emb|AA948752|AA948752 LO-271M13R Ice plant Lambda Uni-Zap XR exp... 35 0.086 emb|AW760788|AW760788 sl35a03.yl Gm-c1027 Glycine max cDNA clone... 39 0.11 emb|AW830924|AW830924 sm19c06.yl Gm-c1027 Glycine max cDNA clone... 39 0.11 emblAQ008266lAQ008266 CpG0480B CpIOWAgDNA1 Cryptosporidium parvu... 39 0.16 50 emblAW696326|AW696326 NF106G11ST1F1087 Developing stem Medicago ... 38 0.21 emb|AI974517|AI974517 T110467e KV0 Medicago truncatula cDNA clon... 38 0.21 emb|AV408651|AV408651 AV408651 Lotus japonicus young plants (two... 38 0.21 emb|AQ651638|AQ651638 Sheared DNA-7G23.TF Sheared DNA Trypanosom... 37 0.40 emb|AQ942781|AQ942781 Sheared DNA-42B17.TF Sheared DNA Trypanoso... 37 0.56 55 gb|BE036036|BE036036 MO18H02 MO Mesembryanthemum crystallinum cD... 36 0.76 gb|C95899|C95899 C95899 Marchantia polymorpha immature sex organ... 36 1.0 emblAA966307 V8c01a1.rl Aspergillus nidulans 24hr asexu... 36 1.0 emb|AI759219|AI759219 EtESTea26c02.yl Eimeria S5-2 Sporozoite st... 28 1.3 gb[U11583[YSCH9196 Saccharomyces cerevisiae chromosome VIII cosm... 35 1.4 60 emb|AQ639257|AQ639257 927P1-10E12.TV 927P1 Trypanosoma brucei ge... 35 1.4 emb|AQ943504|AQ943504 Sheared DNA-43F22.TF Sheared DNA Trypanoso... 35 1.4

--WO-02/22675 -PCT/US01/28506

emb|AW623282|AW623282 EST321227 tomato flower buds 3-8 mm, Corne... dbj|D85226|D85226 Brassica campestris DNA for S glycoprotein, pa... 28 1.6 emb|AF001525|AF001525 Musa acuminata ripening-associated protein... 35 2.0

Query= X84728.6 s at 12349 s at /id source genbank/description gb|aaa17993.1| (m91192) phenylalanine ammonia-lyase [trifolium subterraneum] /blast score 0 /ec number /family /chip nova /gb link

(1962 letters)

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

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Sequences producing significant alignments:

(bits) Value

gb|L11747|POPPALGA Populus tricocarpa X Populus deltoides (hybri... 354 0.0 emb|X99997|BFPAL B.finlaysoniana mRNA for phenylalanine ammonia-... 352 0.0 emb[X58180]MSPAL M.sativa PAL mRNA for phenylalanine ammonia-lyase. 351 0.0 gb|U43338|CLU43338 Citrus limon phenylalanine ammonia-lyase (pal... 350 0.0 gb|L36822|SSNPAL Stylosanthes humilis phenylalanine ammonia-lyas... 350 0.0 emb|AF036948|AF036948 Prunus avium phenylalanine ammonia-lyase (... 349 0.0 emb|AJ250836|CAR250836 Cicer arietinum mRNA for phenylalanine am... 349 0.0 emb|X17462|PCPAL4 P.crispum RNA for PAL4, phenylalanine ammonia-... 349 0.0 emb[X81158]PCPAL2 P.crispum mRNA for phenylalanine ammonia-lyase 2. 349 0.0 emb|X81159|PCPAL3 P.crispum mRNA for phenylalanine ammonia-lyase 3. 348 0.0 dbj[D83075]D83075 Lithospermum erythrorhizon mRNA for phenylalan... 348 0.0 emb|AF237955|AF237955 Rubus idaeus phenylalanine ammonia-lyase 2... 348 0.0 emb|AJ002221|DLJ002221 Digitalis lanata mRNA for phenylalanine a... 347 0.0 gb[M29232[IPBPAL Sweet potato phenylalanine ammonia-lyase mRNA (... 345 0.0 dbj|D26596|CMEPAL Camellia sinensis mRNA for phenylalanine ammon... 344 0.0 dbj|D17467|TOBPAL1 Tobacco mRNA for phenylalanine ammonia-lyase,... 344 0.0 emb[X78269]NTPHEAL N.tabacum (Samsun NN) mRNA for phenylalanine ... 343 0.0 dbj|D83076|D83076 Lithospermum erythrorhizon mRNA for phenylalan... 343 0.0 dbj|D78640|IPBPALA Ipomoea batatas mRNA for phenylalanine ammoni... 342 0.0 emb[Y12461]HAPHAMLY Helianthus annuus mRNA for phenylalanine amm... 341 0.0 emb|AB042520|AB042520 Catharanthus roseus mRNA for phenylalanine... 339 0.0 emb|AF237954|AF237954 Rubus idaeus phenylalamine ammonia-lyase 1... 330 0.0 dbj[D10001]PEAPAL P.sativum mRNA for phenylalanine ammonia-lyase... 349 0.0 emb|X63103|STPAL1 S.tuberosum PAL-1 gene for phenylalanine ammon... 353 0.0 emblAJ238754|CCL238754 Citrus clementina X Citrus reticulata mRN... 350 0.0 emblAJ238753|CCL238753 Citrus clementina X Citrus reticulata mRN... 353 0.0 dbj|D10003|PEAPAL2 P.sativum gene for phenylalanine ammonia-lyas... 349 0.0 dbj E04042 E04042 cDNA sequence coding for pea phenylalanine amm... 349 0.0 emb|AF165998|AF165998 Vigna unguiculata phenylalanine ammonia-ly... 355 0.0 gb[U16130]PAU16130 Persea americana phenylalanine ammonia lyase ... 331 0.0 dbj[D30656]POPPALA Populus kitakamiensis gene for phenylalanine ... 356 0.0 gb[M84466[TOBTPA1A Tobacco phenylalanine ammonialyase (tpa1) gen... 352 0.0 emb|AB008200|AB008200 Nicotiana tabacum palB gene for phenylalan... 352 0.0 dbj[D85850]D85850 Daucus carota gDcPAL1 gene for phenylalanine a... 347 0.0 emb|X52953|DMPAL1 Glycine max PAL1 gene for phenylalanine ammoni... 355 0.0 emb|X99705|TAPALGEN1 T.aestivum PAL gene. 342 0.0

350 0.0

gb[M91192]TFRPAL1X Trifolium subterraneum phenylalanine ammonia-... 345 0.0 gb|M90692|TOMPAL5A Lycopersicon esculentum phenylalanine ammonia... 347 0.0 emb|AB008199|AB008199 Nicotiana tabacum palA gene for phenylalan... 344 0.0

emb[X16772]PCPAL1EX2 P.crispum PAL-1 gene for phenylalanine ammo... 350 0.0

dbj|D43803|POPPALG4B Populus kitakamiensis gene for phenylalanin... 336 0.0 gb|M83314|TOMPHEAMLY Lycopersicon esculentum phenylalanine ammon... 353 0.0

emb|Y07654|PCPAL1 P.crispum pal1 gene.

	emb[X76130 CMPAL C.melo (cantaloupe) pal mRNA. 318 0.0 dbj[D30657 POPPALB Populus kitakamiensis gene for phenylalanine 333 0.0
	dbj D43802 POPPALG2BA Populus kitakamiensis gene for phenylalani 336 0.0
_	emb AF081215 AF081215 Capsicum chinense phenylalanine ammonia-ly 346 0.0
5	dbj D10002 PEAPAL1 P.sativum gene for phenylalanine ammonia-lyas 349 0.0
	emb Z49147 HVPAL7RM H.vulgare partial PAL mRNA for phenylalanine 342 0.0
	emb[X63104]STPAL2 S.tuberosum PAL-2 gene for phenylalanine ammon 353 0.0
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10	gb[M11939]PHVPAL Phaseolus vulgaris L. phenylalanine ammonia-lya 354 0.0
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	emb Z49146 HVPAL3MR H.vulgare partial PAL mRNA for phenylalanine 311 0.0
	emb X75967 VVPAL V.vinifera PAL mRNA for phenylalanine ammonia l 350 0.0
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	emb AF167487 AF167487 Eucalyptus globulus phenylalanine ammonia 334 e-178
	emb[X99725]TAPALGEN2 T.aestivum PAL gene, coding region. 300 e-137
	emb AW218834 AW218834 EST301314 tomato root during/after fruit s 347 e-134
20	emb AW031612 AW031612 EST275066 tomato callus, TAMU Lycopersicon 327 e-132
	emb AJ289609 BPE289609 Betula pendula partial pal gene for pheny 205 e-128
	emb AJ278116 BPE278116 Betula pendula partial pall gene for phen 205 e-128
	emb AW219303 AW219303 EST301785 tomato root during/after fruit s 339 e-126
	emb AW726548 AW726548 GA_Ea0022A01 Gossypium arboreum 7-10 dpa 323 e-124
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	gb BE035366 BE035366 MM06H04 MM Mesembryanthemum crystallinum cD 235 e-11:
	emb AW776946 AW776946 EST336011 DSIL Medicago truncatula cDNA cl 280 e-114
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30	emb AF218453 AF218453 Coffea arabica clone 369.1.6r phenylalanin 247 e-105
	emb AW216505 AW216505 EST295219 tomato callus, TAMU Lycopersicon 209 e-104
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0.5	emb AW220322 AW220322 EST302805 tomato root during/after fruit s 255 2e-98
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	emb AW781748 AW781748 sl90e11.yl Gm-c1037 Glycine max cDNA clone 326 3e-88
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	emb A1894514 A1894514 EST263957 tomato callus, TAMU Lycopersicon 294 1e-78
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	Joh link http://www.2 nchi nlm nih gov/hthin

/gb\_link http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb|al022347| /ncgi ~ http://www.ncgr.org/cgi-bin/ff?al022347 (1554 letters) 60

Database: plantfungal 661,018 sequences; 426,114,510 total letters

5 Searching.....done

Score E Sequences producing significant alignments: (bits) Value 10 emb[AF078082]AF078082 Phaseolus vulgaris receptor-like protein k... 251 1e-65 gb|BE034949|BE034949 ML07F03 ML Mesembryanthemum crystallinum cD... 231 8e-60 emb|AW031255|AW031255 EST274630 tomato callus, TAMU Lycopersicon... 231 1e-59 emb[X98520]BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 230 2e-59 emb|Y18260|BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 227 1e-58 15 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 226 4e-58 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. 225 7e-58 emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 225 7e-58 gb|U00443|BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 224 1e-57 gb[BE034855]BE034855 ML05C11 ML Mesembryanthemum crystallinum cD... 186 1e-57 20 emb|AB012106|AB012106 Brassica rapa mRNA for SRK45, complete cds. 222 6e-57 gblM76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 221 8e-57 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 221 2e-56 gb|BE057261|BE057261 sm99f12.yl Gm-c1015 Glycine max cDNA clone ... 220 3e-56 emb[Y14285]BOY14285 Brassica oleracea mRNA for SFR1 protein. 219 6e-56 25 gb|U51741|ITU51741 Ipomoea trifida receptor protein kinase 2 (IR... 218 8e-56 gb[M97667]BNASTKR Brassica napus ssp. oleifera serine/threonine ... 216 3e-55 gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 215 5e-55 emb|AF088885|AF088885 Nicotiana tabacum receptor-like kinase CHR... 213 3e-54 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 211 1e-53 30 emb[Y12531]BOBRLKGEN B.oleraceae gene encoding serine/threonine ... 116 6e-53 emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 207 2e-52 emb|Y14286|BOY14286 Brassica oleracea SFR3 gene, partial. dbj D38564 BOLRPKB Brassica campestris mRNA for receptor protein... 204 1e-51 emb|AW736407|AW736407 EST332421 KV3 Medicago truncatula cDNA clo... 198 4e-51 35 dbj[D38563]BOLRPKA Brassica campestris mRNA for receptor protein... 202 7e-51 gb|U20948|ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 201 1e-50 emb|Al896155|Al896155 EST265598 tomato callus, TAMU Lycopersicon... 166 1e-50 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 196 2e-49 emb|AW760240|AW760240 sl59g07.yl Gm-c1027 Glycine max cDNA clone... 196 4e-49 · 40 emb|AW033458|AW033458 EST277029 tomato callus, TAMU Lycopersicon... 196 4e-49 emb|AB024420|AB024419S2 Brassica oleracea SRK13 gene, exon 2, 3,... 114 4e-48 emb|AJ245480|BNA245480 Brassica napus slg gene for S-locus glyco... 113 7e-48 emb[Y12530]BOARLKGEN B.oleraceae gene encoding serine/threonine ... 115 1e-47 emb|Z18921|BOSRKL B.oleracea gene for S-receptor kinase-like pro... 111 2e-47 45 emb|AB024422|AB024421S2 Brassica oleracea SRK13-b gene, exon 2, ... 112 3e-47 emb|AB000970|AB000970 Brassica campestris gene for receptor kina... 111 3e-47 emb[X79432]BOSRK3 B.oleracea SRK3 gene. 107 6e-47 emb|Z30211|BOSRK29G B.oleracea (alboglabra) srk29 gene. 108 6e-47 emb|AB013718|AB013717S2 Brassica rapa gene for SRK46Bra, exon 2,... 111 2e-46 50 emb|AW684339|AW684339 NF015G04NR1F1000 Nodulated root Medicago t... 120 3e-46 emb|AB000971|AB000971 Brassica campestris pseudogene for recepto... 105 8e-46 emb|A1895838|A1895838 EST265281 tomato callus, TAMU Lycopersicon... 184 2e-45 emb|AW039406|AW039406 EST281663 tomato mixed elicitor, BTI Lycop... 107 3e-45 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 106 4e-45 55 emblAW034624|AW034624 EST278308 tomato callus, TAMU Lycopersicon... 182 6e-45 dbj|D88193|D88193 Brassica rapa DNA for S-receptor kinase, compl... 109 9e-45 emb|Z18884|BOSRKRPC B.oleracea encoding S-receptor kinase relate... 116 1e-43 emb|AI775997|AI775997 EST257097 tomato resistant, Cornell Lycope... 177 2e-43 emb|A1895623|A1895623 EST265066 tomato callus, TAMU Lycopersicon... 177 3e-43

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Searching......done

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Score E

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

15

## Score E

Sequences producing significant alignments: (bits) Value emb|AW756217|AW756217 sl17e08.yl Gm-c1036 Glycine max cDNA clone... 96 1e-20 20 emb|AW561018|AW561018 EST316066 DSIR Medicago truncatula cDNA cl... 95 2e-20 gb[L47866]L47866 BNAF1507 Mustard flower buds Brassica rapa cDNA... 94 6e-20 emb|X94986|MEBGLA M.esculenta bglA gene. 94 6e-20 emb|AW257154|AW257154 EST305291 KV2 Medicago truncatula cDNA clo... 93 8e-20 gb[U39228[PAU39228 Prunus avium beta-glucosidase mRNA, partial cds. 93 8e-20 25 emb|AI729894|AI729894 BNLGHi5504 Six-day Cotton fiber Gossypium ... 93 8e-20 gb|U95298|MEU95298 Manihot esculenta linamarase (pLIN-GEN) gene,... 93 8e-20 gb|S35175|S35175 linamarase=beta-glucosidase [Manihot esculenta=... 92 2e-19 emb|AF072736|AF072736 Pinus contorta beta-glucosidase mRNA, comp... 92 2e-19 emb|AW568866|AW568866 si73c03.yl Gm-c1031 Glycine max cDNA clone... 91 4e-19 30 gb|U50201|PSU50201 Prunus serotina prunasin hydrolase precursor ... 90 5e-19 emb|AW560545|AW560545 EST315593 DSIR Medicago truncatula cDNA cl... 90 7e-19 emb|AI900802|AI900802 sb93e10.y1 Gm-c1017 Glycine max cDNA clone... 89 2e-18 gb|BE058816|BE058816 sn21c08.yl Gm-c1016 Glycine max cDNA clone ... 89 2e-18 emb|AW597982|AW597982 sj40c05.yl Gm-c1008 Glycine max cDNA clone... 89 2e-18 35 emb|AW395210|AW395210 sh45c06.y1 Gm-c1017 Glycine max cDNA clone... 89 2e-18 emb|AW570004|AW570004 si85e02.yl Gm-c1031 Glycine max cDNA clone... 89 2e-18 emb|AW733524|AW733524 sk74g01.yl Gm-c1016 Glycine max cDNA clone... 89 2e-18 emb|AW423986|AW423986 sh59a09.yl Gm-c1015 Glycine max cDNA clone... emb|AW423387|AW423387 sh65b01.yl Gm-c1015 Glycine max cDNA clone... 89 2e-18 40 emb|AW597368|AW597368 si91h08.yl Gm-c1031 Glycine max cDNA clone... 88 3e-18 emb[AW569966]AW569966 si83h05.yl Gm-c1031 Glycine max cDNA clone... 88 3e-18 emb[AW569751]AW569751 si79h11.yl Gm-c1031 Glycine max cDNA clone... 88 3e-18 emb|AW569883|AW569883 si82g07.y1 Gm-c1031 Glycine max cDNA clone... 88 3e-18 emb|AW317654|AW317654 sg55e02.y1 Gm-c1025 Glycine max cDNA clone... 87 5e-18 45 emb|AW350857|AW350857 GM210009B10F4R Gm-r1021 Glycine max cDNA 3... 87 5e-18 emb|AW309715|AW309715 sf23f08.x1 Gm-c1028 Glycine max cDNA clone... 87 7e-18 emb|AF221526|AF221526 Prunus serotina prunasin hydrolase isoform... 86 9e-18 emb|AB003089|AB003089 Polygonum tinctorium mRNA for beta-glucosi... 86 9e-18 emb|AW279169|AW279169 sf67c06.yl Gm-c1013 Glycine max cDNA clone... 85 2e-17 50 gb|U72154|BNU72154 Brassica nigra beta-glucosidase (psr3.1) mRNA... 85 2e-17 emb|AW394514|AW394514 sh32a04.yl Gm-c1017 Glycine max cDNA clone... 84 3e-17 emb[AW202129]AW202129 sf12c05.yl Gm-c1027 Glycine max cDNA clone... 84 3e-17 gb|U26025|PSU26025 Prunus serotina amygdalin hydrolase isoform A... 84 5e-17 dbi|D83177|CSAF26G Costus speciosus mRNA for furostanol glycosid... 83 9e-17 55 emb|AI774009|AI774009 EST255109 tomato resistant, Cornell Lycope... 63 2e-16 emb|AF143377|AF143377 AF143377 Pisum sativum library (Fristensky... 82 2e-16 emb|AI772446|AI772446 EST253546 tomato resistant, Cornell Lycope... 63 3e-16 emb|AW651266|AW651266 EST329720 tomato germinating seedlings, TA... 63 3e-16 emb|AW317658|AW317658 sg55f02.y1 Gm-c1025 Glycine max cDNA clone... 81 3e-16 60 emb|AW042637|AW042637 ST24C01 Pine TriplEx shoot tip library Pin... 60 4e-16

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emb|AW132549|AW132549 se05f01.yl Gm-c1013 Glycine max cDNA clone... 80 6e-16 gb[L41869]BLYBGQ6 Hordeum vulgare L. beta-glucosidase (BGQ60) ge... 65 6e-16 emb|AB003110|AB003110 Trichoderma reesei bgl2 gene for bete-gluc... 80 8e-16 emb|AW760442|AW760442 sl50d01.yl Gm-c1027 Glycine max cDNA clone... 79 1e-15 emb|AJ005950|AJ005950 AJ005950 chickpea mRNA CAP-4 Cicer arietin... 79 1e-15 emb|X56734|TRBG361 Trifolium repens mRNA for non-cyanogenic beta... 78 2e-15 emb|AW201515|AW201515 sf04b02.yl Gm-c1027 Glycine max cDNA clone... 78 4e-15 emb[AA415086]AA415086 Mg0020 RCW Lambda Zap Express Library Pyri... 78 4e-15 emb|AI941073|AI941073 sb84a09.yl Gm-c1010 Glycine max cDNA clone... 63 5e-15 10 emb|AW311377|AW311377 sg38b12.yl Gm-c1025 Glycine max cDNA clone... 63 5e-15 emb|AW677220|AW677220 DG1 6 D11.g1 A002 Dark Grown 1 (DG1) Sorgh... 77 7e-15 gb|U33817|SBU33817 Sorghum bicolor dhurrinase mRNA, nuclear gene... 77 7e-15 emb|AW744827|AW744827 LG1 384\_D09.g1\_A002 Light Grown 1 (LG1) So... 77 7e-15 emb|AW923243|AW923243 DG1\_50\_B11.g1\_A002 Dark Grown 1 (DG1) Sorg... 77 7e-15 15 emb|AW923628|AW923628 DG1 56 E12.g1 A002 Dark Grown 1 (DG1) Sorg... emb|AW922651|AW922651 DG1 20 B09.g1\_A002 Dark Grown 1 (DG1) Sorg... 77 7e-15 emb|AW286529|AW286529 LG1\_334\_A07.g1\_A002 Light Grown 1 (LG1) So... 77 7e-15' emb|AI974587|AI974587 T113037e KV2 Medicago truncatula cDNA clon... 65 9e-15 emb|AW676749|AW676749 DG1\_14\_B06.g1\_A002 Dark Grown 1 (DG1) Sorg... 76 1e-14 20 emb|AB003109|AB003109 Humicola grisea var. thermoidea bgl4 gene ... 76 1e-14 emb|AI723853|AI723853 RHIZ1\_28\_C08.yl\_A001 Rhizome1 Sorghum hale... 76 1e-14 emb|AI724790|AI724790 RHIZ1\_9\_C08.y2\_A001 Rhizomel Sorghum halep... 76 1e-14 emb|AW685200|AW685200 NF027E09NR1F1000 Nodulated root Medicago t... 65 1e-14 gb|BE124555|BE124555 EST393590 GVN Medicago truncatula cDNA clon... 65 le-14 25 emb|AW774242|AW774242 EST333393 KV3 Medicago truncatula cDNA clo... 65 1e-14 emblAW286277|AW286277 LG1 329 G04.gl A002 Light Grown 1 (LG1) So... 76 1e-14 emb|AW286262|AW286262 LG1\_329\_E04.g1\_A002 Light Grown 1 (LG1) So... 76 1e-14 emb|AW284398|AW284398 LG1\_289\_B04.g1\_A002 Light Grown 1 (LG1) So... 76 1e-14 emb|AW283021|AW283021 LG1\_301\_G01.g1\_A002 Light Grown 1 (LG1) So... 76 1e-14 30 emb|AW284016|AW284016 LG1\_264\_G03.g1\_A002 Light Grown 1 (LG1) So... 76 1e-14 emb|AW282563|AW282563 LG1\_312\_A09.g1\_A002 Light Grown 1 (LG1) So... 76 1e-14 emb|AW282564|AW282564 LG1\_312\_A08.g1\_A002 Light Grown 1 (LG1) So... 76 1e-14 emb|AW565734|AW565734 LG1 348 H08.g1 A002 Light Grown 1 (LG1) So... emb[X82577]BNBGL B.napus mRNA for beta-glucosidase. 35 emb|AF149311|AF149311 Rauvolfia serpentina raucaffricine-O-beta-... 75 3e-14 gb[BE021518[BE021518 sm59e05.yl Gm-c1028 Glycine max cDNA clone ... 74 4e-14 emb|AF082991|AF082991 Avena sativa beta-D-glucosidase beta subun... 73 9e-14 emb|AW423704|AW423704 sh50f06.yl Gm-c1017 Glycine max cDNA clone... 73 1e-13 emb|X78433|ASLBDG A.sativa L. mRNA for beta-D-glucosidase. 40 emb|AF163097|AF163097 Dalbergia cochinchinensis dalcochinin 8'-O... 62 2e-13 emb|AW733253|AW733253 sk70d08.yl Gm-c1016 Glycine max cDNA clone... 71 3e-13 emb|AW922447|AW922447 DG1\_19\_F09.g1\_A002 Dark Grown 1 (DG1) Sorg... 60 5e-13 emb|AW671821|AW671821 LG1 351 F12.g1 A002 Light Grown 1 (LG1) So... 59 9e-13 gb[BE059820]BE059820 sn37e08.y1 Gm-c1016 Glycine max cDNA clone ... 68 3e-12 45 emblAJ133406|DLA133406 Digitalis lanata mRNA for cardenolide 16-... 68 4e-12 emb|AW773684|AW773684 EST332670 KV3 Medicago truncatula cDNA clo... 61 6e-12 emb[AW099840]AW099840 sd17b01.y2 Gm-c1012 Glycine max cDNA clone... 60 6e-12 emb X67838 BNDNAM B. napus DNA for myrosinase. 66 le-11 emb|AW923660|AW923660 DG1\_57\_B08.g1\_A002 Dark Grown 1 (DG1) Sorg... 66 1e-11 50 emb|AW284341|AW284341 LG1 275 C12.gl A002 Light Grown 1 (LG1) So... 66 1e-11 emb[X79080]BNMYRMC B.napus mRNA for myrosinase MC. emb|Z21977|BNMYR1A B.napus Myr1.Bn1 gene encoding myrosinase, th... 66 2e-11 emb|X56733|TRBG104 T.repens mRNA for cyanogenic beta-glucosidase... 52 2e-11 emb|AF016864|AF016864 Orpinomyces sp. PC-2 beta-glucosidase (bgl... 64 5e-11 55 emb[X59881]SAMYRMB1 S.alba thioglucoside glucohydrolase (myrosin... 63 7e-11 emb[AW927073]AW927073 HVSMEg0009H02 Hordeum vulgare pre-anthesis... 57 1e-10 gb[L11258[BNAMYROS Brassica campestris myrosinase gene, complete...

Query= AF024504.13\_at 12630\_at /id\_source genbank /description 60 gb|aaf18681.1|af024504\_11 (af024504) unknown protein [arabidopsis thaliana] /blast\_score 1.00e-129 /ec\_number /family /chip nova

/gb\_link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|af024504| /ncgi http://www.ncgr.org/cgi-bin/ff?af024504 (1374 letters)

5

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

10

Score E

Sequences producing significant alignments:

(bits) Value

emb|AW216758|AW216758 EST295472 tomato callus, TAMU Lycopersicon... 119 3e-34 15 emb|AW219044|AW219044 EST301526 tomato root during/after fruit s... 108 7e-33 emblAW031258|AW031258 EST274633 tomato callus, TAMU Lycopersicon... 108 7e-33 emb[AI772760]AI772760 EST253860 tomato resistant, Cornell Lycope... 108 7e-33 emb|AI486533|AI486533 EST244854 tomato ovary, TAMU Lycopersicon ... 71 5e-29 emb[AI898645]AI898645 EST268088 tomato ovary, TAMU Lycopersicon ... 71 6e-28 20 emb|AW686482|AW686482 NF041H08NR1F1000 Nodulated root Medicago t... 112 8e-28 emb|AW704059|AW704059 sk27d01.yl Gm-c1028 Glycine max cDNA clone... 119 2e-27 emb|AW350257|AW350257 GM210007B20B11R Gm-r1021 Glycine max cDNA ... 122 5e-27 emb|AW759958|AW759958 s156c04.y1 Gm-c1027 Glycine max cDNA clone... 115 7e-25 emb|AW126130|AW126130 N100327e rootphos(-) Medicago truncatula c... 115 1e-24 25 emblAI776363|AI776363 EST257463 tomato resistant, Cornell Lycope... 114 1e-24 emb|AI731720|AI731720 BNLGHi10579 Six-day Cotton fiber Gossypium... 112 5e-24 emb|AW034441|AW034441 EST278012 tomato callus, TAMU Lycopersicon... 103 5e-24 emb|AI896076|AI896076 EST265519 tomato callus, TAMU Lycopersicon... 102 1e-23 emb|AI898707|AI898707 EST268150 tomato ovary, TAMU Lycopersicon ... 71 2e-22 30 emb|AW310471|AW310471 sf36e11.x1 Gm-c1028 Glycine max cDNA clone... 105 6e-22 emb|AW219445|AW219445 EST302023 tomato root during/after fruit s... 105 8e-22 emb|AW648110|AW648110 EST326564 tomato germinating seedlings, TA... 105 1e-21 emb|AW831572|AW831572 sm34h08.y1 Gm-c1028 Glycine max cDNA clone... 100 2e-20 gb|BE124541|BE124541 EST393576 GVN Medicago truncatula cDNA clon... 89 8e-20 35 emb|AW671843|AW671843 LG1\_352\_B09.b1\_A002 Light Grown 1 (LG1) So... 64 2e-18 emb|AW564955|AW564955 LG1\_313\_H09.b1\_A002 Light Grown 1 (LG1) So... 64 2e-18 emb|AW329431|AW329431 N200666e rootphos(-) Medicago truncatula c... 93 4e-18 emb|AW929644|AW929644 EST338432 tomato flower buds 8 mm to pre-a... 73 7e-18 emb|AW907159|AW907159 EST343282 potato stolon, Cornell Universit... 60 2e-14 40 emb|AW565825|AW565825 LG1\_352\_B09.g1\_A002 Light Grown 1 (LG1) So... 57 8e-14 emb|AW625736|AW625736 EST319643 tomato radicle, 5 d post-imbibit... 56 8e-14 emb|AI812987|AI812987 2D4 Pine Lambda Zap Xylem library Pinus ta... 52 4e-13 gblBE060164bE060164 HVSMEg0011B20f Hordeum vulgare pre-anthesis... 76 5e-13 emb|AW217067|AW217067 EST295781 tomato callus, TAMU Lycopersicon... 74 2e-12 45 emb[AW398760]AW398760 EST309260 L. pennellii trichome, Cornell U... 54 2e-12 emb|AW694156|AW694156 NF072H06ST1F1059 Developing stem Medicago ... 58 3e-12 emb|AW773860|AW773860 EST332846 KV3 Medicago truncatula cDNA clo... 58 3e-12 emb|AW394635|AW394635 sh33h09.yl Gm-c1017 Glycine max cDNA clone... 55 3e-12 emb|AW203644|AW203644 sf36e11.yl Gm-c1028 Glycine max cDNA clone... 69 4e-12 50 emb|AI728748|AI728748 BNLGHi11497 Six-day Cotton fiber Gossypium... 57 4e-12 emb|AW697758|AW697758 Str1-D8 Sugar Beet germination cDNA librar... 46 5e-11 emb|AW030302|AW030302 EST273557 tomato callus, TAMU Lycopersicon... 53 7e-11 emb|AW982668|AW982668 HVSMEg0003O06f Hordeum vulgare pre-anthesi... 48 2e-10 emb|AW287116|AW287116 LG1 265 G01.b2 A002 Light Grown 1 (LG1) So... 64 2e-09 55 emb|AW734790|AW734790 sk88g11.y1 Gm-c1035 Glycine max cDNA clone... 42 5e-09 emb|AT000776|AT000776 AT000776 Brassica rapa guard cell Brassica... 62 7e-09 emb|A1775378|A1775378 EST256478 tomato resistant, Cornell Lycope... 42 1e-08 emblAW775488|AW775488 EST334553 DSIL Medicago truncatula cDNA cl... 58 2e-07 emb|AW928719|AW928719 EST337507 tomato flower buds 8 mm to pre-a... 39 2e-07 60 emb|AW282482|AW282482 LG1 313\_B10.g1\_A002 Light Grown 1 (LG1) So... 57 3e-07 emb|AW225609|AW225609 T210061e KV0 Medicago truncatula cDNA clon... 40 4e-07

---PCT/US01/28506 -WO 02/22675

	emb AT000645 AT000645 AT000645 Brassica rapa guard cell Brassica 56 6e-07 emb AW731593 AW731593 GA_Ea0030P10 Gossypium arboreum 7-10 dpa 37 1e-06
•	emb AW282432 AW282432 LG1_313_H09.g1_A002 Light Grown 1 (LG1) So 55 2e-06 emb AI736841 kJ36841 sb34c09.y1 Gm-c1012 Glycine max cDNA clone 50 5e-05
5	emb AW154916 AW154916 EST290281 tomato root deficiency, Cornell 40 6e-05
	emb AI054948 AI054948 coau0002J14 Cotton Boll Abscission Zone cD 49 7e-05 emb AI487433 AI487433 EST245755 tomato ovary, TAMU Lycopersicon 44 1e-04
	emb A1490477 A1490477 EST248846 tomato ovary, TAMU Lycopersicon 44 1e-04
•	emb AI895257 AI895257 EST264700 tomato callus, TAMU Lycopersicon 47 4e-04
10	emb AW564044 AW564044 LG1_281_A09.b1_A002 Light Grown 1 (LG1) So 47 4e-04
	emb AW565728 AW565728 LG1_348_E11.g1_A002 Light Grown 1 (LG1) So 47 4e-04
	emb AW922753 AW922753 DG1_45_C05.g1_A002 Dark Grown 1 (DG1) Sorg 47 4e-04
	emb AW746301 AW746301 WS1_40_G05.g1_A002 Water-stressed 1 (WS1) 47 4e-04 emb AW033344 AW033344 EST276915 tomato callus, TAMU Lycopersicon 40 5e-04
15	emb AI486814 AI486814 EST245136 tomato ovary, TAMU Lycopersicon 46 5e-04
	emb AI486642 AI486642 EST244963 tomato ovary, TAMU Lycopersicon 44 6e-04
	emb AW774292 AW774292 EST333443 KV3 Medicago truncatula cDNA clo 46 7e-04
	emb AW472252 AW472252 si21g12.yl Gm-c1029 Glycine max cDNA clone 46 7e-04
20	emb AW705448 AW705448 sk49c01.yl Gm-c1019 Glycine max cDNA clone 35 0.001
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	emb AW011001 AW011001 ST15E02 Pine TriplEx shoot tip library Pin 45 0.002
	emb AI563253 AI563253 EST00377 watermelon lambda zap library Cit 44 0.002
	emb AW568784 AW568784 si61a06.yl Gm-r1030 Glycine max cDNA clone 44 0.002
25	emb AI485669 AI485669 EST243990 tomato ovary, TAMU Lycopersicon 44 0.003
	emb AW154917 AW154917 EST290282 tomato root deficiency, Cornell 44 0.003
	emb AI895825 AI895825 EST265268 tomato callus, TAMU Lycopersicon 44 0.003 emb AF109661 AF109661 AF109661 Capsicum annuum root susceptible 42 0.004
	emb AF109661 AF109661 AF109661 Capsicum annuum root susceptible 42 0.004 emb AI487481 AI487481 EST245803 tomato ovary, TAMU Lycopersicon 41 0.023
30	emb AV411141 AV411141 AV411141 Lotus japonicus young plants (two 41 0.023
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	http://www.ncgr.org/cgi-bin/ff?ac006920
60	(1512 letters)
	•

Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 5 Score Sequences producing significant alignments: (bits) Value emb|AF223643|AF223643 Pisum sativum xyloglucan fucosyltransferas... 311 e-147 10 emb|AW719438|AW719438 LjNEST4f10r Lotus japonicus nodule library... 164 1e-64 emb|AA660464|AA660464 00349 MtRHE Medicago truncatula cDNA 5', m... 128 6e-38 emb|AW693268|AW693268 NF064A03ST1F1000 Developing stem Medicago ... 109 5e-32 emb|AW508697|AW508697 si35c12.yl Gm-r1030 Glycine max cDNA clone... 77 6e-31 emb|AW689377|AW689377 NF018E07ST1F1000 Developing stem Medicago ... 78 2e-30 15 gb|BE058474|BE058474 sn16f08.y1 Gm-c1016 Glycine max cDNA clone ... 102 6e-21 emb|AW704030|AW704030 sk27a04.yl Gm-c1028 Glycine max cDNA clone... 64 5e-20 emblAU089974|AU089974 AU089974 Hordeum vulgare subsp. vulgare Up... 45 1e-19 emb|AW720399|AW720399 LjNEST22c8r Lotus japonicus nodule library... 83 1e-18 emb|AU089968|AU089968 AU089968 Hordeum vulgare subsp. vulgare Up... 45 5e-17 20 emb|AV417193|AV417193 AV417193 Lotus japonicus young plants (two... 88 2e-16 emb[AQ855554]AQ855554 CpG1646A CpIOWAgDNA1 Cryptosporidium parvu... 35 1.1 emb|Z37536|LTND9 L.tarentolae ND9 mRNA encoding putative NADH de... 35 2.1 emb|AW934637|AW934637 EST353529 tomato flower buds 0-3 mm, Corne... 35 2.1 emb|AF050740|AF050740 Plasmodium falciparum variant-specific sur... 35 2.1 25 emb|AI329848|AI329848 b8h09ne.rl Neurospora crassa evening cDNA ... 35 2.1 emb|AW035275|AW035275 EST280638 tomato callus, TAMU Lycopersicon... 35 2.1 emb|AW933190|AW933190 EST359033 tomato fruit mature green, TAMU ... 35 2.1 emb|AL109832|SPAC630 S.pombe chromosome I cosmid c630. 34 2.9 emb|AC004709|AC004709 Plasmodium falciparum chromosome 12, \*\*\* S... 34 4.0 30 emb|A85491|A85491 Sequence 150 from Patent EP0866129. emb|AC004688|AC004688 Plasmodium falciparum chromosome 12 clone ... 34 4.0 emb[AA786891]AA786891 m7a05a1.rl Aspergillus nidulans 24hr asexu... 34 4.0 gb[BE124954]BE124954 EST393989 GVN Medicago truncatula cDNA clon... 27 4.7 emb|AL356172|NCB23L21 Neurospora crassa DNA linkage group II BAC... 33 5.5 35 emb|AW617216|AW617216 EST323627 L. hirsutum trichome, Cornell Un... 33 5.5 emb|AL160493|LMFLCHR26 Leishmania major Friedlin assembled chrom... 33 7.6 emb|AF056622|AF056622 Vitis vinifera putative Cu/Zn superoxide d... 33 7.6 40 Query= AC004077.149\_s\_at 12989\_s\_at /id\_source genbank /description gb|aac26690.1| (ac004077) putative cytochrome p450 [arabidopsis thaliana] /blast\_score 0 /ec\_number /family /chip nova /gb\_link /ncgi (1488 letters) 45 Database: plantfungal 661,018 sequences; 426,114,510 total letters 50 Sequences producing significant alignments: (bits) Value emblAW203670|AW203670 sf36h04.yl Gm-c1028 Glycine max cDNA clone... 334 1e-90 55 emb|AW268012|AW268012 EST306234 DSIR Medicago truncatula cDNA cl... 246 2e-83 emb|AW041000|AW041000 EST283864 tomato mixed elicitor, BTI Lycop... 307 1e-82 emb|AW472433|AW472433 si25b07.yl Gm-c1029 Glycine max cDNA clone... 306 3e-82 emb|AW560118|AW560118 EST315166 DSIR Medicago truncatula cDNA cl... 166 3e-64 emb|AW394771|AW394771 sh35b02.yl Gm-c1017 Glycine max cDNA clone... 213 3e-54 60 emb|AW622568|AW622568 EST313368 tomato root during/after fruit s... 181 5e-51

emb|AW737712|AW737712 EST339139 tomato flower buds, anthesis, Co... 162 1e-45

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	Common Massillinin Miliplica A

	emb Z67751 SC38KCXVI S.cerevisiae DNA (chromosome XVI; 38 kb). 87 8e-28
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30	emb AF106079 AF106079 Chlamydomonas reinhardtii dynein heavy cha 34 1.8
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25	gb M76671 TOMEXTENB L.esculentum extensin (class II) gene, compl 33 4.6
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Database: plantfungal

## 661,018 sequences; 426,114,510 total letters

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(1761 letters)

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15 Database: plantfungal 661,018 sequences; 426,114,510 total letters

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20 Score

Sequences producing significant alignments: (bits) Value

emb[AV411445]AV411445 AV411445 Lotus japonicus young plants (two... 198 1e-49 emb|AV415235|AV415235 AV415235 Lotus japonicus young plants (two... 190 3e-47 emb|AV415282|AV415282 AV415282 Lotus japonicus young plants (two... 190 3e-47 emb|AW906097|AW906097 EST342218 potato stolon, Cornell Universit... 162 8e-39 emb|AV413830|AV413830 AV413830 Lotus japonicus young plants (two... 161 1e-38 emblAV419573|AV419573 AV419573 Lotus japonicus young plants (two... 82 5e-36 emblAA660119lAA660119 EST00006 watermelon lambda zap express lib... 148 7e-36 emb|AI507855|AI507855 sa88a07.y1 Gm-c1004 Glycine max cDNA clone... 91 2e-35 emb|AW160273|AW160273 EST290131 L. pennellii trichome, Cornell U... 146 4e-34 emb|AV428532|AV428532 AV428532 Lotus japonicus young plants (two... 80 3e-30 emb|AV424557|AV424557 AV424557 Lotus japonicus young plants (two... 128 1e-28 emb|AW668007|AW668007 GA Ea0012E04 Gossypium arboreum 7-10 dpa ... 117 2e-25 emblAW922981|AW922981 DG1\_48\_B03.b1\_A002 Dark Grown 1 (DG1) Sorg... 105 8e-22 emb|AW432282|AW432282 sh71f09.yl Gm-c1015 Glycine max cDNA clone... 100 3e-20 emb|AW621610|AW621610 EST312408 tomato root during/after fruit s... 100 5e-20 emb|AV419981|AV419981 AV419981 Lotus japonicus young plants (two... 95 2e-18 emb|AI937949|AI937949 sc06a10.yl Gm-c1012 Glycine max cDNA clone... 88 3e-16 emblAV406642|AV406642 AV406642 Lotus japonicus young plants (two... 67 6e-10 emb|AW497095|AW497095 ga53g06.yl Moss EST library PPU Physcomitr... 42 3e-06 emb|AW289850|AW289850 NXNV006F04F Nsf Xylem Normal wood Vertical... 40 4e-05 emb|AW279534|AW279534 sf90g12.yl Gm-c1019 Glycine max cDNA clone... 46 7e-04 emb|AQ911817|AQ911817 LMAJFV1 ln03b09.yl Leishmania major FV1 ra... 46 7e-04 emb|AW689523|AW689523 NF021G01ST1F1000 Developing stem Medicago ... 39 0.007 emb|AW218923|AW218923 EST301405 tomato root during/after fruit s... 41 0.030 emb|AW695963|AW695963 NF101A10ST1F1072 Developing stem Medicago ... 40 0.041 emb|AW931531|AW931531 EST357374 tomato fruit mature green, TAMU ... 40 0.041 emb|AW691108|AW691108 NF041C09ST1F1000 Developing stem Medicago ... 39 0.14 emb|AA791402|AA791402 LO-136M13R Ice plant Lambda Uni-Zap XR exp... 38 0.20 emb|AI001408|AI001408 L0-330M13R Ice plant Lambda Uni-Zap XR exp... 38 0.20 emb|AI731457|AI731457 BNLGHi9807 Six-day Cotton fiber Gossypium ... 38 0.27 emb|AQ161167|AQ161167 mgxb0007101r CUGI Rice Blast BAC Library P... 35 1.3 emb|AW690030|AW690030 NF027E08ST1F1000 Developing stem Medicago ... 35 1.8 gb|BE124825|BE124825 EST393860 GVN Medicago truncatula cDNA clon... 35 1.8 emb|AW684862|AW684862 NF022D06NR1F1000 Nodulated root Medicago t... 35 1.8 emb|AW686248|AW686248 NF039F10NR1F1000 Nodulated root Medicago t... 35 1.8 emb|AW696000|AW696000 NF100H02ST1F1026 Developing stem Medicago ... 35 1.8 emb|AW686364|AW686364 NF037B05NR1F1000 Nodulated root Medicago t... 35 1.8 emb|AW926002|AW926002 HVSMEg0006A15 Hordeum vulgare pre-anthesis... 35 2.5 emb|AA660722|AA660722 00613 MtRHE Medicago truncatula cDNA 5', m... 34 3.5

emb|AI054829|AI054829 coau0002E06 Cotton Boll Abscission Zone cD... 34 3.5 emb|AW719283|AW719283 LjNEST2b8r Lotus japonicus nodule library,... 34 3.5 emb[AW677540]AW677540 DG1 8 H03.b1 A002 Dark Grown 1 (DG1) Sorgh... emb|X78996|CSTETFP C.sativus mRNA for tetrafunctional protein. emb|AQ654502|AQ654502 Sheared DNA-25C14.TR Sheared DNA Trypanoso... 33 6.6 emb|AW981113|AW981113 EST392266 GVN Medicago truncatula cDNA clo... 33 6.6 gb|L40389|YSAERG11X Candida glabrata ERG11 gene, complete cds. emb[X79365]CSMFPB C.sativus MFP-b mRNA. gb|U05612|CSU05612 Colysis sintenensis chloroplast large subunit... 28 6.8 emb|X02433|MIPSCOX2 Pea mitochondrial gene for cytochrome oxidas... 24 6.9 emb|AQ654503|AQ654503 Sheared DNA-25C15.TR Sheared DNA Trypanoso... 33 9.0 emb|AI163319|AI163319 A039P66U Hybrid aspen plasmid library Popu... 33 9.0 emb|AQ641986|AQ641986 RPCI93-EcoRI-6N23.TJ RPCI93-EcoRI Trypanos... 33 9.0 emb|AL034558|PFMAL3P2 Plasmodium falciparum MAL3P2, complete seq... 33 9.0 15 emb|AQ649633|AQ649633 Sheared DNA-17K4.TF Sheared DNA Trypanosom... 33 9.0 emb|AI728749|AI728749 BNLGHill499 Six-day Cotton fiber Gossypium... 33 9.0 emblAQ940463|AQ940463 Sheared DNA-33A23.TR Sheared DNA Trypanoso... 33 9.0 emb|AQ647492|AQ647492 RPCI93-EcoRI-6G10.TJ RPCI93-EcoRI Trypanos... 33 9.0 emb|AW443117|AW443117 EST308047 tomato mixed elicitor, BTI Lycop... 33 9.0 20 emb|AA231646|AA231646 CDO328.F cDNA from oat Avena sativa cDNA c... 33 9.0 Query= AF000657.28 at 15415 at /id\_source genbank/description gb|aab72169.1| (af000657) hypothetical protein [arabidopsis thaliana] /blast\_score 9.00e-96 /ec number /family /chip nova /gb link 25 http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|af000657| /ncgi http://www.ncgr.org/cgi-bin/ff?af000657 (564 letters) 30 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching 35 Score E Sequences producing significant alignments: (bits) Value emb|AW622665|AW622665 EST313465 tomato root during/after fruit s... 166 8e-42 emb|AW625889|AW625889 EST319892 tomato radicle, 5 d post-imbibit... 166 1e-41 40 emb|AI776839|AI776839 EST257939 tomato resistant, Cornell Lycope... 166 2e-40 emb|A1898331|A1898331 EST267774 tomato ovary, TAMU Lycopersicon ... 166 2e-40 emb|AI772887|AI772887 EST253987 tomato resistant, Cornell Lycope... 166 2e-40 emb|AI782639|AI782639 EST263518 tomato susceptible, Cornell Lyco... 161 2e-40 emb[AW035141]AW035141 EST280403 tomato callus, TAMU Lycopersicon... 162 3e-40 45 emb|AW559773|AW559773 EST314821 DSIR Medicago truncatula cDNA cl... 136 4e-40 emb|AI488935|AI488935 EST247274 tomato ovary, TAMU Lycopersicon ... 162 1e-39 emb|AW626015|AW626015 EST319922 tomato radicle, 5 d post-imbibit... 161 4e-39 emb|AW035118|AW035118 EST280380 tomato callus, TAMU Lycopersicon... 159 1e-38 emb|AI442841|AI442841 sa27e02.x1 Gm-c1004 Glycine max cDNA clone... 131 2e-38 50 gb|BE057238|BE057238 sm99d06.yl Gm-c1015 Glycine max cDNA clone ... 133 4e-37 gb|BE021888|BE021888 sm63g06.yl Gm-c1028 Glycine max cDNA clone ... 151 4e-36 emblAW782173|AW782173 sm02d12.yl Gm-c1027 Glycine max cDNA clone... 151 4e-36 emb|AW981240|AW981240 EST392330 DSIL Medicago truncatula cDNA cl... 127 5e-36 emb|AW624844|AW624844 EST313673 tomato radicle, 5 d post-imbibit... 150 6e-36 55 emb|AW348098|AW348098 GM210001A11F3R Gm-r1021 Glycine max cDNA 3... 121 1e-35 emb|AW348956|AW348956 GM210004A12C2R Gm-r1021 Glycine max cDNA 3... 127 2egb|BE058374|BE058374 sn15b11.yl Gm-c1016 Glycine max cDNA clone ... 127 2e-35 emb|AW570563|AW570563 sj63f09.yl Gm-c1033 Glycine max cDNA clone... 127 2e-35 60 emb|AW568265|AW568265 si69f02.yl Gm-r1030 Glycine max cDNA clone... 127 2e-35

emb|AW666011|AW666011 sk31a06.yl Gm-c1028 Glycine max cDNA clone... 127 2e-35

emb|AW306271|AW306271 se48e06.y1 Gm-c1017 Glycine max cDNA clone... 125 6e-35 emb|AI772066|AI772066 EST253166 tomato resistant, Cornell Lycope... 146 1e-34 emb|AW395160|AW395160 sh40f10.y1 Gm-c1017 Glycine max cDNA clone... 124 1e-34 emb|AW306309|AW306309 se49a06.yl Gm-c1017 Glycine max cDNA clone... 124 1e-34 gb|BE022272|BE022272 sm73a07.yl Gm-c1028 Glycine max cDNA clone ... 146 1e-34 gb|BE021996|BE021996 sm65c01.yl Gm-c1028 Glycine max cDNA clone ... 146 1e-34 emb|AW706543|AW706543 sj57h02.y1 Gm-c1033 Glycine max cDNA clone... 146 1e-34 emb|AW620987|AW620987 sj50d09.y1 Gm-c1033 Glycine max cDNA clone... 146 1e-34 emb|AW349990|AW349990 GM210006B20G11R Gm-r1021 Glycine max cDNA ... 146 1e-10 emb|AW706119|AW706119 sj52a11.yl Gm-c1033 Glycine max cDNA clone... 146 1e-34 emb|AW311107|AW311107 sg33a02.y1 Gm-c1025 Glycine max cDNA clone... 124 2e-34 emb|AW200806|AW200806 se93g05.y1 Gm-c1027 Glycine max cDNA clone... 123 3e-34 emb|AW830450|AW830450 sm27b08.y1 Gm-c1028 Glycine max cDNA clone... 145 3e-34 15 emb|AW458429|AW458429 sh09b05.y1 Gm-c1016 Glycine max cDNA clone... 127 4e-34 emb|AW559537|AW559537 EST314585 DSIR Medicago truncatula cDNA cl... 79 5e-34 emb|AW559584|AW559584 EST314632 DSIR Medicago truncatula cDNA cl... 79 5e-34 emb|AI939020|AI939020 sc64e06.yl Gm-c1016 Glycine max cDNA clone... 144 7e-34 emb|AW234019|AW234019 sf33b10.y1 Gm-c1028 Glycine max cDNA clone... 143 1e-33 20 emb|AW011055|AW011055 ST16B07 Pine TriplEx shoot tip library Pin... 142 2e-33 emb|AW687043|AW687043 NF005D02RT1F1016 Developing root Medicago ... 133 2e-33 gb[BE022312]BE022312 sm73c09.y1 Gm-c1028 Glycine max cDNA clone ... 142 3e-33 emb|AW620826|AW620826 sj47a07.y1 Gm-c1033 Glycine max cDNA clone... 119 3e-33 emb|AW666225|AW666225 sk34a04.y1 Gm-c1028 Glycine max cDNA clone... 140 9e-33 25 emb|AI494896|AI494896 sb06g10.yl Gm-c1004 Glycine max cDNA clone... 140 9e-33 emb|AI443387|AI443387 sa31d11.x1 Gm-c1004 Glycine max cDNA clone... 139 1e-32 emb|AW203753|AW203753 sf37h02.y1 Gm-c1028 Glycine max cDNA clone... 139 2e-32 emb|AW559725|AW559725 EST314717 DSIR Medicago truncatula cDNA cl... 78 4e-32 emb|AW289859|AW289859 NXNV007A04F Nsf Xylem Normal wood Vertical... 137 4e-32 30 emb|AW234737|AW234737 sf18f10.y1 Gm-c1028 Glycine max cDNA clone... 136 2e-31 emb|AW351059|AW351059 GM210010B10G3R Gm-r1021 Glycine max cDNA 3... 136 2eemb|AW684772|AW684772 NF021H06NR1F1000 Nodulated root Medicago t... 120 5e-31 emb|AW775144|AW775144 EST334295 KV3 Medicago truncatula cDNA clo... 76 9e-31 35 emb|AI495113|AI495113 sb03f02.yl Gm-c1004 Glycine max cDNA clone... 133 1e-30 emb|AW472306|AW472306 si23e07.yl Gm-c1029 Glycine max cDNA clone... 117 2e-30 emb|AW035139|AW035139 EST280401 tomato callus, TAMU Lycopersicon... 131 5e-30 emb|AI496590|AI496590 sb13g02.y1 Gm-c1004 Glycine max cDNA clone... 129 2e-29 emb|AW351151|AW351151 GM210011A10E12R Gm-r1021 Glycine max cDNA ... 81 2e-29 40 emb|AW460064|AW460064 si08g11.yl Gm-c1029 Glycine max cDNA clone... 105 4e-29 emb|AW432674|AW432674 sh83h05.yl Gm-c1016 Glycine max cDNA clone... 126 1e-28 emb|AW507521|AW507521 si53a07.yl Gm-r1030 Glycine max cDNA clone... 106 1e-28 emb|AI461088|AI461088 sa73g10.y1 Gm-c1004 Glycine max cDNA clone... 123 8e-28 emb|AI461045|AI461045 sa73c04.y1 Gm-c1004 Glycine max cDNA clone... 123 8e-28 45 emb|AW010003|AW010003 ST01B12 Pine TriplEx shoot tip library Pin... 123 8e-28 gb|BE021390|BE021390 sm48e04.yl Gm-c1028 Glycine max cDNA clone ... 122 2e-27 gb|BE022539|BE022539 sm86c09.yl Gm-c1015 Glycine max cDNA clone ... 121 4e-27 emb|AW559258|AW559258 EST306094 DSIR Medicago truncatula cDNA cl... 118 3e-26 emb|AW760707|AW760707 sl36a09.yl Gm-c1027 Glycine max cDNA clone... 95 8e-26 50 emb|AI941013|AI941013 sb83c06.yl Gm-c1010 Glycine max cDNA clone... 95 8e-26 emb|AW745074|AW745074 LG1\_386\_D11.b1\_A002 Light Grown 1 (LG1) So... 91 3e-25 emb|AI900284|AI900284 sc03d11.y1 Gm-c1012 Glycine max cDNA clone... 111 4e-25 emb|AW666457|AW666457 sk36g12.y1 Gm-c1028 Glycine max cDNA clone... 114 7e-25 emb|AI522845|AI522845 sa74e03.yl Gm-c1004 Glycine max cDNA clone... 114 7e-25 55 emb[AW350289]AW350289 GM210008A10D4R Gm-r1021 Glycine max cDNA 3... 113 9eemb|AW233771|AW233771 sf26e08.y1 Gm-c1028 Glycine max cDNA clone... 105 1e-24 gb|BE021431|BE021431 sm49a02.y1 Gm-c1028 Glycine max cDNA clone ... 111 4e-24 emb|AW495792|AW495792 NXNV\_065\_D12\_FF Nsf Xylem Normal wood Vert... 110 8e-24 60 emb|AI441701|AI441701 sa60b06.yl Gm-c1004 Glycine max cDNA clone... 110 8e-24 emb[AW686910]AW686910 NF003H08RT1F1000 Developing root Medicago ... 79 9e-24

emb|AW310235|AW310235 sf33b10.x1 Gm-c1028 Glycine max cDNA clone... 110 1e-23 emb|AI495196|AI495196 sa89d04.y1 Gm-c1004 Glycine max cDNA clone... 100 4e-23 emb|AI442650|AI442650 sa43f05.y1 Gm-c1004 Glycine max cDNA clone... 106 1e-22 emb|AW926095|AW926095 HVSMEg0006F02 Hordeum vulgare pre-anthesis... 106 1e-22 emb|AW309915|AW309915 sf26e08.x1 Gm-c1028 Glycine max cDNA clone... 105 2e-22 emb|AI812367|AI812367 1E2 Pine Lambda Zap Xylem library Pinus ta... 104 5e-22 gb|BE022988|BE022988 sm69b05.yl Gm-c1028 Glycine max cDNA clone ... 103 1e-21 emb|AI748713|AI748713 sb61a02.y1 Gm-c1010 Glycine max cDNA clone... 82 1e-21 gb|BE020169|BE020169 sm39e04.y1 Gm-c1028 Glycine max cDNA clone ... 103 1e-21 10 gb|BE021658|BE021658 sm60g09.yl Gm-c1028 Glycine max cDNA clone ... 101 3e-21 emb|AI855670|AI855670 sc32f08.yl Gm-c1014 Glycine max cDNA clone... 100 2e-20 emb|AI920195|AI920195 1725 Pine Lambda Zap Xylem library Pinus t... 73 2e-20 emb|AW686987|AW686987 NF004G02RT1F1018 Developing root Medicago ... 99 2e-20 emb|AW684941|AW684941 NF023D03NR1F1000 Nodulated root Medicago t... 88 1e-19 15 emb|AI494665|AI494665 sb18f10.yl Gm-c1004 Glycine max cDNA clone... 89 1e-19 emb|AW277935|AW277935 sf87g05.y1 Gm-c1019 Glycine max cDNA clone... 97 1e-19 gb|BE023344|BE023344 sm70f11.yl Gm-c1028 Glycine max cDNA clone ... 78 2e-19 emb|AV422621|AV422621 AV422621 Lotus japonicus young plants (two... 60 2e-19 emb|AW127416|AW127416 M110599 DSIL Medicago truncatula cDNA clon... 95 4e-19 20 Ouery= AL030978.64\_at 15431\_at /id\_source genbank /description emb|caa19722.1| (al030978) putative protein [arabidopsis thaliana] /blast score 2.00e-51/ec number /family /chip nova/gb link

http://www3.ncbi.nlm.nih.gov/htbin-

25 post/entrez/query?db=n&form=6&dopt=g&uid=gb|a1030978| /ncgi http://www.ncgr.org/cgi-bin/ff?al030978 (393 letters)

Database: plantfungal

30

40

45

50

55

60

661,018 sequences; 426,114,510 total letters

Searching.....done

E Score

35 Sequences producing significant alignments:

(bits) Value

emb|AW218792|AW218792 EST301272 tomato root during/after fruit s... 104 2e-30 emb|AW218791|AW218791 EST301271 tomato root during/after fruit s... 104 2e-30 emblAI781536|AI781536 EST262415 tomato susceptible, Cornell Lyco... 104 2e-30 emb|AI894486|AI894486 EST263929 tomato callus, TAMU Lycopersicon... 100 3e-29 emb|AW255546|AW255546 ML580 peppermint glandular trichome Mentha... 96 4e-25 emb|AW781055|AW781055 s188d08.yl Gm-c1037 Glycine max cDNA clone... 95 5e-25 emb|AW980335|AW980335 EST391488 GVN Medicago truncatula cDNA clo... 94 1e-24 emb|AW982334|AW982334 HVSMEg0002O20f Hordeum vulgare pre-anthesi... 86 1e-24 gb|H74448|H74448 150 Standard Brassica napus cDNA clone R29F, mR... 92 4e-24 emb|AI960943|AI960943 sc93a04.y1 Gm-c1019 Glycine max cDNA clone... 92 5e-24 emb|AU036790|AU036790 AU036790 Cryptomeria japonica seedling lea... 88 5e-24 emb|AW598247|AW598247 sj42f01.yl Gm-c1008 Glycine max cDNA clone... 80 3e-23 emb|AW100651|AW100651 sd58b12.y1 Gm-c1008 Glycine max cDNA clone... 89 4e-23 emblAW208200|AW208200 M110845e GVSN Medicago truncatula cDNA clo... 88 5e-23 emb|AW559726|AW559726 EST314718 DSIR Medicago truncatula cDNA cl... 83 2e-22 emb|AW101120|AW101120 sd74c08.yl Gm-c1008 Glycine max cDNA clone... 94 2e-22 emb|AW926990|AW926990 HVSMEg0009D05 Hordeum vulgare pre-anthesis... 103 8e-22 gb[BE124173[BE124173 EST394298 DSIL Medicago truncatula cDNA clo... 94 1e-21 emblAT000903|AT000903 AT000903 Brassica rapa guard cell Brassica... 54 2e-20 emb[AW690635]AW690635 NF031H11ST1F1000 Developing stem Medicago ... 75 6e-20 emb|AW126039|AW126039 N100235e rootphos(-) Medicago truncatula c... 75 6e-20 emb|AW126127|AW126127 N100324e rootphos(-) Medicago truncatula c... 75 7e-20 emb|AI812959|AI812959 2A11 Pine Lambda Zap Xylem library Pinus t... 75 2e-19 emb|AI043565|AI043565 L30-578T3 Ice plant Lambda Uni-Zap XR expr... 75 3e-19

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      emb|AW461253|AW461253 NXNV060C05F Nsf Xylem Normal wood Vertical... 70 7e-18
      emb|AW126244|AW126244 N100089e rootphos(-) Medicago truncatula c... 53 2e-17
      emb|AW683890|AW683890 NF003A10NR1F1000 Nodulated root Medicago t... 49 4e-17
      emb|AW922806|AW922806 DG1 46 G06.g1 A002 Dark Grown 1 (DG1) Sorg... 84 4e-17
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      emb|AI779006|AI779006 EST259885 tomato susceptible, Cornell Lyco... 73 1e-12
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25
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      emb|AW784020|AW784020 NXNV_103 A11 F Nsf Xylem Normal wood Verti... 54 8e-08
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      emb|AW309614|AW309614 sf22a01.x1 Gm-c1028 Glycine max cDNA clone...
      gb|L01433|SOYSCAM4X Soybean calmodulin (SCaM-4) mRNA, complete cds. 40 0.008
35
      gb|U20292|STU20292 Solanum tuberosum clone PCM3 calmodulin gene,... 40 0.008
      emb|AW458431|AW458431 sh09b07.yl Gm-c1016 Glycine max cDNA clone...
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      emb|AW706736|AW706736 sk02d02.y1 Gm-c1023 Glycine max cDNA clone... 39 0.022
      gb[M88307]BNACALM Brassica juncea calmodulin mRNA, complete cds.
                                                                         38 0.030
40
      gb|U10150|BNU10150 Brassica napus Naehan calmodulin (bcm1) mRNA,... 38 0.030
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      emb|AW317381|AW317381 sg48f11.yl Gm-c1025 Glycine max cDNA clone... 38 0.041
      emb|AI973796|AI973796 sd10d10.y1 Gm-c1020 Glycine max cDNA clone... 37 0.078
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45
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      emb|AI080805|AI080805 TENU3514 T. cruzi epimastigote normalized ... 35 0.28
      emb|AW696965|AW696965 NF110H11ST1F1095 Developing stem Medicago ... 34 0.52
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50
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      emblAW267797|AW267797 EST305925 DSIR Medicago truncatula cDNA cl... 34 0.52
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55
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	emb AQ367507 AQ367507 toxb0002H01r CUGI Tomato BAC Library Lycop 34 1.9
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60
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5	emb AL034556 PFMAL3P5 Plasmodium falciparum MAL3P5, complete seq 33 1.5
•	
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	emb AC005505 AC005505 Plasmodium falciparum chromosome 12 clone 33 2.0
	emb AC005308 AC005308 Plasmodium falciparum chromosome 12 clone 33 2.0
	emb AL049180 PFMAL13P1 Plasmodium falciparum chromosome 13 strai 33 2.0
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	emb AW458882 AW458882 sh16a07.y1 Gm-c1016 Glycine max cDNA clone 33 2.0
	11/17/00004//17/000004//17/000004//
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,	ombit Constal Constal Plant 4 Linguistic for basic-animo-acid period 22.30
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- •	emb AW035978 AW035978 EST282837 tomato callus, TAMU Lycopersicon 32 3.8
	emb[AF120285]AF120285 Pleurotus ostreatus linear mitochondrial p 32 3.8
	emb AL049181 PFMAL13P4 Plasmodium falciparum chromosome 13 strai 32 3.8
25	emb AW691746 AW691746 NF043G06ST1F1000 Developing stem Medicago 32 3.8
35	emb Z98551 PFMAL3P6 Plasmodium falciparum MAL3P6, complete seque 32 3.8
	emb AW030017 AW030017 EST273272 tomato callus, TAMU Lycopersicon 32 3.8
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	Control of Collock Collock in the control of the collock in the co
45	emb AC011016 AC011016 Leishmania major chromosome 35 clone L218 32 3.8
43	emb AL033503 CAC49C4 C.albicans cosmid Ca49C4. 32 3.8
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	ombi750144SDAC2H1S comba the manual Technology of the State of the Sta
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S	emb AE001422 AE001422 Plasmodium falciparum chromosome 2, sectio 31 5.2
55	emb AE001401 AE001401 Plasmodium falciparum chromosome 2, sectio 31 5.2
	emb AE001380 AE001380 Plasmodium falciparum chromosome 2, sectio 31 5.2
	emb AC004157 AC004157 Plasmodium falciparum chromosome 12 clone 31 5.2
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60	emb AQ949411 AQ949411 Sheared DNA-38N10.TF Sheared DNA Trypanoso 31 5.2

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

Score I

15 Sequences producing significant alignments:

10

(bits) Value

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5	emb AI026521 AI026521 TENU0733 T. cruzi epimastigote normalized 73 1e-16
3	emb X59720 SCCHRIII S.cerevisiae chromosome III complete DNA seq 85 2e-16 gb T36700 T36700 EST101687 S. cerevisiae strain X2180-1A Sacchar 79 1e-14
	emb AW650769 AW650769 EST329223 tomato germinating seedlings, TA 79 1e-14
•	emb AW739119 AW739119 gb26a12.y1 Moss EST library PPN Physcomitr 79 1e-14
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	emb AA948748 AA948748 L0-267M13R Ice plant Lambda Uni-Zap XR exp 66 1e-10
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	emb AW686356 AW686356 NF040G07NR1F1000 Nodulated root Medicago t 35 0.36
25	emb AV389829 AV389829 AV389829 Chlamydomonas reinhardtii C9 Chla 34 0.49
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	emb AW329632 AW329632 N200892e rootphos(-) Medicago truncatula c 30 1.2
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	emb AW278010 AW278010 sf89d09.y1 Gm-c1019 Glycine max cDNA clone 30 3.6
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60	http://www3.ncbi.nlm.nih.gov/htbin-
	post/entrez/query?db=n&form=6&dopt=g&uid=gb a1024486  /ncgi

http://www.ncgr.org/cgi-bin/ff?al024486 (844 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching.....

Score E

10

5

Sequences producing significant alignments: (bits) Value emb|AI489421|AI489421 EST247760 tomato ovary, TAMU Lycopersicon ... 336 1e-91 emb|AW329288|AW329288 N200509e rootphos(-) Medicago truncatula c... 307 9e-83 emb|AW733754|AW733754 sk77g11.yl Gm-c1016 Glycine max cDNA clone... 290 9e-78 15 emb|AW561007|AW561007 EST316055 DSIR Medicago truncatula cDNA cl... 287 8e-77 emb|AI489189|AI489189 EST247528 tomato ovary, TAMU Lycopersicon ... 273 1e-72 emb|AI488043|AI488043 EST246365 tomato ovary, TAMU Lycopersicon ... 269 2e-71 emb|AW625207|AW625207 EST319114 tomato radicle, 5 d post-imbibit... 258 5e-68 emb|AW559910|AW559910 EST314958 DSIR Medicago truncatula cDNA cl... 258 5e-68 20 emblAW559909|AW559909 EST314957 DSIR Medicago truncatula cDNA cl... 245 4e-64 emb|AI487722|AI487722 EST246044 tomato ovary, TAMU Lycopersicon ... 241 5e-63 emb|AW218466|AW218466 EST303649 tomato radicle, 5 d post-imbibit... 238 3e-62 emb|AW624963|AW624963 EST313792 tomato radicle, 5 d post-imbibit... 234 8e-61 emb[AW774997]AW774997 EST334148 KV3 Medicago truncatula cDNA clo... 217 1e-55 25 emb|AA495616|AA495616 c425 Zhou and Ragan 1993 Gracilaria gracil... 118 3e-52 emb|AW678376|AW678376 WS1\_15\_G07.g1\_A002 Water-stressed 1 (WS1) ... 175\_5e-50 emb|AW760767|AW760767 sl36g03.yl Gm-c1027 Glycine max cDNA clone... 194 8e-49 emb|AI416755|AI416755 sa18h08.x1 Gm-c1005 Glycine max cDNA clone... 175 5e-44 gb|BE020624|BE020624 sm51d11.yl Gm-c1028 Glycine max cDNA clone ... 175 4e-43 30 emb|AW218418|AW218418 EST303601 tomato radicle, 5 d post-imbibit... 174 9e-43 emb|AW684921|AW684921 NF023B02NR1F1000 Nodulated root Medicago t... 138 8e-40 emb|AL035218|SPCC1281 S.pombe chromosome III cosmid c1281. 81 3e-37 emb|AW222782|AW222782 EST299593 tomato fruit red ripe, TAMU Lyco... 149 4e-35 emb|AW098211|AW098211 ga07f10.y1 Moss EST library CPU Ceratodon ... 147 9e-35 35 emb|Z48639|SC9920 S.cerevisiae chromosome XIII cosmid 9920. emb|AA660525|AA660525 00411 MtRHE Medicago truncatula cDNA 5', m... 104 1e-33 emb|Z28301|SCYKR076W S.cerevisiae chromosome XI reading frame OR... 77 5e-32 emb|AW931289|AW931289 EST357132 tomato fruit mature green, TAMU ... 123 1e-27 emb|X85807|SCCVIIRA S.cerevisiae chromosome VII 27kbp right arm ... 51 2e-23 40 emb|Z72939|SCYGR154C S.cerevisiae chromosome VII reading frame O... 51 2e-23 emb|AA788015|AA788015 r4b01a1.r1 Aspergillus nidulans 24hr asexu... 90 1e-22 emb|AW931288|AW931288 EST357131 tomato fruit mature green, TAMU ... 106 2e-22 emb|AW686971|AW686971 NF004E10RT1F1082 Developing root Medicago ... 104 1e-21 emb|AI416795|AI416795 sa18h08.y1 Gm-c1005 Glycine max cDNA clone... 98 7e-20 45 emb|AI327824|AI327824 j0c06a1\_rl Aspergillus nidulans 24hr asexu... 77 le-17 emb|AI212279|AI212279 x1e09a1.rl Aspergillus nidulans 24hr asexu... 90 2e-17 emb|AI165879|AI165879 B002P45U Hybrid aspen plasmid library Popu... 84 2e-15 emb|AA966531|AA966531 w5h12a1.rl Aspergillus nidulans 24hr asexu... 82 5e-15 emb|AW931345|AW931345 EST357188 tomato fruit mature green, TAMU ... 82 7e-15 50 emblAW442112|AW442112 EST311508 tomato fruit red ripe, TAMU Lyco... 80 2e-14 emb|AW704348|AW704348 sk18b11.y1 Gm-c1028 Glycine max cDNA clone... 77 2e-13 emb|AW221318|AW221318 EST297787 tomato fruit mature green, TAMU ... 68 9e-11 emb|AQ875035|AQ875035 V120D9 mTn-3xHA/lacZ Insertion Library, st... 67 2e-10 emb|AW934230|AW934230 EST360073 tomato fruit mature green, TAMU ... 66 3e-10 55 emb|AA787150|AA787150 m8a03a1.rl Aspergillus nidulans 24hr asexu... 58 1e-07 emb|AQ492097|AQ492097 V111F10 mTn-3xHA/lacZ Insertion Library Sa... 55 9e-07 emb|AA788014|AA788014 r4b01a1.fl Aspergillus nidulans 24hr asexu... 35 4e-04 emb|AA787149|AA787149 m8a03a1.fl Aspergillus nidulans 24hr asexu... 46 5e-04 emb[AC007865]AC007865 Trypanosoma brucei chromosome II clone RPC... 38 0.12 60 emb|AA788490|AA788490 13a06a1.fl Aspergillus nidulans 24hr asexu... 35 0.58 emb|AV420850|AV420850 AV420850 Lotus japonicus young plants (two... 29 1.0

	emb AF041468 AF041468 Guillardia theta complete plastid genome. 35 1.1
	emb AW037453 AW037453 EST279002 tomato mixed elicitor, BTI Lycop 35 1.1
	emb Z34288 SCXCDNA S.cerevisiae (S288C) X chromosome DNA (17137bp). 34 1.5
	emb Z49346 SCYJL071W S.cerevisiae chromosome X reading frame ORF 34 1.5
5	2. LV 712 COLUMN 18 11 11 11 11 11 11 11 11 11 11 11 11
,	emb X71362 HVDHN7 H.vulgare gene for dehydrin 7. 34 1.5
	emb[X88851]SCESTGENE S.cerevisiae DNA for hypotetical proteins a 34 1.5
	emb AF043087 AF043087 Hordeum vulgare dehydrin 1 (dhn1) gene, co 34 1.5
	emb AI988341 AI988341 sc99e11.yl Gm-c1020 Glycine max cDNA clone 34 2.1
	emb Y10224 CMAO1 C.melo ao1 gene, partial. 34 2.1
10	emb AI900471 AI900471 sc11b06.y1 Gm-c1012 Glycine max cDNA clone 34 2.1
10	and A Orgaso Ala Orgaso A Million and College and Coll
	emb AQ874584 AQ874584 V111B3 mTn-3xHA/lacZ Insertion Library, st 34 2.1
	emb AW279161 AW279161 sf67b09.yl Gm-c1013 Glycine max cDNA clone 34 2.1
	emb Z99531 SPAC19D5 S.pombe chromosome I cosmid c19D5. 33 2.8
	emb AJ273736 AJ273736 AJ273736 Metarhizium anisopliae ARSEF 2575 33 2.8
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	emb AW982536 AW982536 HVSMEg0003I12f Hordeum vulgare pre-anthesi 33 3.9
•	amble 12001/4/WI A 1001/4 When arranged to the DNA Course A Course A 22 20
	emb AJ229614 KLAJ9614 Kluyveromyces lactis DNA fragment for sequ 33 3.9
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43.	661,018 sequences; 426,114,510 total letters
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_	emb AF053127 AF053127 Malus domestica leucine-rich receptor-like 169 8e-99
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      Database: plantfungal
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    Score

      Sequences producing significant alignments:
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      emb|AF166121|AF166121 Hordeum vulgare Cf2/Cf5 disease resistance... 94 3e-22
      emblAI779817|AI779817 EST260696 tomato susceptible, Cornell Lyco... 100 1e-21
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      emb|AI727073|AI727073 BNLGHi7256 Six-day Cotton fiber Gossypium ... 95 2e-18
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      emb[AW735867]AW735867 EST336635 tomato flower buds 0-3 mm, Corne... 89 2e-16
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      emb[AI488443]AI488443 EST246782 tomato ovary, TAMU Lycopersicon ... 88 3e-16
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      emb|AW127035|AW127035 ga20c02.yl Moss EST library PPU Physcomitr... 88 4e-16
      emb|AI895669|AI895669 EST265112 tomato callus, TAMU Lycopersicon... 88 4e-16
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emb|AW618685|AW618685 EST320671 L. pennellii trichome, Cornell U... 88 4e-16 emb|AI894989|AI894989 EST264432 tomato callus, TAMU Lycopersicon... 88 4e-16 emb|AI782076|AI782076 EST262955 tomato susceptible, Cornell Lyco... 87 5e-16 emb|AW648795|AW648795 EST327249 tomato germinating seedlings, TA... 87 5e-16 emb|AW694942|AW694942 NF081G02ST1F1018 Developing stem Medicago ... 78 6e-16 emb|AV415057|AV415057 AV415057 Lotus japonicus young plants (two... 87 7e-16 emb|AW399097|AW399097 EST309597 L. pennellii trichome, Cornell U... 63 8e-16 emb|AW761367|AW761367 sl66e07.yl Gm-c1027 Glycine max cDNA clone... 86 1e-15 emb|AW649208|AW649208 EST327662 tomato germinating seedlings, TA... 86 1e-15 10 emb|AV428126|AV428126 AV428126 Lotus japonicus young plants (two... 85 2e-15 emb|AW290705|AW290705 NXNV045C11F Nsf Xylem Normal wood Vertical... 85 2e-15 emb|AI352869|AI352869 MB70-4A PZ204.BNlib Brassica napus cDNA cl... 85 2e-15 emblAA661049|AA661049 00946 MtRHE Medicago truncatula cDNA 5' si... 85 2e-15 emb|AW455303|AW455303 EST311841 tomato root during/after fruit s... 85 3e-15 15 emb|AW869870|AW869870 NXNV\_122\_E02\_F Nsf Xylem Normal wood Verti... 85\_3e-15 emb|AW398651|AW398651 EST309151 L. pennellii trichome, Cornell U... 84 4e-15 emb|AW398661|AW398661 EST309161 L. pennellii trichome, Cornell U... 84 4e-15 emb|AF049920|AF049920 Petunia x hybrida PGPS/D4 (PGPS/D4) mRNA, ... 84 4e-15 emb|AW624634|AW624634 EST322579 tomato flower buds 3-8 mm, Corne... 59 4e-15 20 emb|AW618879|AW618879 EST320865 L. pennellii trichome, Cornell U... 62 4e-15 gb|BE035885|BE035885 MO11F02 MO Mesembryanthemum crystallinum cD....\_84 5e-15 Query= AL096882.91\_s\_at 16817\_s\_at /id\_source genbank /description emb|cab51412.1| (al096882) acc synthase (atacs-6) [arabidopsis 25 thaliana] /blast score 0 /ec number /family /chip nova /gb link /ncgi (1488 letters) Database: plantfungal 30 661,018 sequences; 426,114,510 total letters Searching..... Score E 35 Sequences producing significant alignments: (bits) Value emb[X82273]BOACCS B.oleracea mRNA for ACC synthase. 986 0.0 emb[X72676]BJMACC B. juncea mRNA for 1-Aminocyclopropane-1-carbo... 875 0.0 emb|AF057563|AF057563 Nicotiana glutinosa 1-aminocyclopropane-1-... 765 0.0 40 emb|AJ005002|NTAJ5002 Nicotiana tabacum mRNA for 1-aminocyclopro... 761 0.0 emb|AB034992|AB034992 Malus domestica MdACS-5A mRNA for 1-aminoc... 754 0.0 emb|AB034993|AB034993 Malus domestica MdACS-5B mRNA for 1-aminoc... 755 0.0 gb|U72389|LEU72389 Lycopersicon esculentum 1-aminocyclopropane-1... 756 0.0 gb[U72390]LEU72390 Lycopersicon esculentum 1-aminocyclopropane-1... 753 0.0 45 emb|AB033503|AB033503 Populus euramericana peacs-2 mRNA for 1-am... 741 0.0 gb|U68216|CPU68216 Carica papaya ACC synthase mRNA, complete cds. 461 0.0 emb|AF061605|AF061605 Nicotiana glutinosa ACC synthase mRNA, com... 742 0.0 emb|AB013100|AB013100 Lycopersicon esculentum LE-ACS6 mRNA for 1... 741 0.0 emb|AJ012551|CSI012551 Citrus sinensis mRNA for ACC synthase. 50 emb|AB013346|AB013346 Lycopersicon esculentum mRNA for 1-aminocy... 739 0.0 emb|AB033502|AB033502 Populus euphratica peacs-1 mRNA for 1-amin... 620 0.0 gb|U88971|PHU88971 Pelargonium hortorum 1-aminocyclopropane-1-ca... 455 0.0 emb|Z11613|VRACCSYNM V.radiata mRNA for ACC synthase. 456 0.0 emb[X98492]NTACCS Nicotiana tabacum mRNA for ACC-synthase (clone... 455 0.0 55 emb[X67100]GMCACCS1 G.max mRNA for ACC synthase. 454 0.0

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60

emb|AF016459|AF016459 Pisum sativum 1-aminocyclopropane-1-carbox... 445 0.0 emb|AF109927|AF109927 Musa acuminata 1-aminocyclopropane-1-carbo... 707 0.0 dbi|D30805|CUS1A1CS Melon mRNA for 1-aminocyclopropane-1-carboxy... 705 0.0 emb|AB021906|AB021906 Musa acuminata MA-ACS1 mRNA for ACC syntha... 703 0.0 5 emb|AB006803|AB006803 Cucumis sativus CS-ACS1 mRNA for ACC synth... 703 0.0 emb|AF129508|AF129508 Musa acuminata 1-aminocyclopropane-1-carbo... 702 0.0 emb[X65982[NTXACCSYN N.tabacum mRNA for 1-aminocyclopropane-1-ca... 438 0.0 emb|AB031026|AB031026 Prunus mume PM-ACS1 mRNA for ACC synthase,... 434 0.0 emb|AB015625|AB015625 Pyrus pyrifolia pPPACS3 mRNA for 1-aminocy... 418 0.0 10 dbj[D01032|CUCACCW Cucurbita maxima mRNA for 1-aminocyclopropane... 695 0.0 dbj|E03724|E03724 cDNA encoding detriment induced type 1-aminocy... 695 0.0 gb]M58323|CUCACCSYN Cucurbita pepo 1-aminocyclopropane-1-carboxy... 692 0.0 emb[Y11357]CP1A1CS C.papaya mRNA for 1-aminocyclopropane-1-carbo... 430 0.0 emb|X62536|LEACC L.esculentum mRNA for ACC synthase. 15 emb|AF057562|AF057562 Nicotiana glutinosa 1-aminocyclopropane-1-... 437 0.0 emb[X59145]LEACC2MR Lycopersicon esculentum LE-ASCC2 mRNA (ptACC... 326 0.0 ) gb[U79999]MAU79999 Musa acuminata ACC synthase (acs3) mRNA, comp... 687 0.0 emb[Y15739]MAACSYNTH Musa acuminata mRNA for 1-aminocyclopropane... 687 0.0 gb]M34289|TOMACS Tomato 1-aminocyclopropane-1-carboxylate syntha... 327 0.0 20 gb|M63490|TOMACCS Tomato 1-aminocyclopropane-1-carboxylate synth... 682 0.0 emb|X59146|LEACC4MR Lycopersicon esculentum LE-ACC4 mRNA (ptACC4... 681 0.0 gb[U17229]PHU17229 Pelargonium hortorum clone pGAC-1 1-aminocycl... 438 0.0 gb|U17231|PHU17231 Pelargonium hortorum clone pGAC-2 1-aminocycl... 423 0.0 emb|AF038945|AF038945 Rumex palustris 1-aminocyclopropane-1-carb... 407 0.0 25 gb|L20634|POTACCSYN Solanum tuberosum 1-aminocyclopropane-1-carb... 403 0.0 emb|AJ276295|CSI276295 Citrus sinensis partial mRNA for ACC synt... 446 0.0 emb|AF239987|AF239987 Prunus persica ACC synthase ACS1 mRNA, par... 662 0.0 emb|AF178076|AF178076 Carica papaya 1-aminocyclopropane-1-carbox... 395 0.0 emb[X82265]CAACC1 C.anuum mRNA for 1-aminocylopropane-1-carboxyl... 639 0.0 30 gb[U70842|STU70842 Solanum tuberosum 1-aminocyclopropane-1-carbo... 637 0.0 emb|AF144746|AF144746 Solanum melongena 1-aminocyclopropane-1-ca... 636 0.0 emblAJ012577|CPA012577 Carica papaya mRNA for 1-aminocyclopropan... 387 e-180 gb[U73815]MDU73815 Malus domestica ACC synthase (MdACS-2) mRNA, ... 623 e-178 dbj|D37937|D37937 Cucumis melo mRNA for 1-aminocyclopropane-1-ca... 380 e-177 35 emb|Z11562|VRACCSYN V.radiata mRNA for 1-aminocyclopropane-1-car... 375 e-176 emb|AF177769|AF177769 Carica papaya 1-aminocyclopropane-1-carbox... 369 e-175 emb|AF083814|AF083814 Antirrhinum majus ACC synthase 1 (ACS1) mR... 605 e-172 emb|AF178077|AF178077 Carica papaya 1-aminocyclopropane-1-carbox... 316 e-172 emb|AF239989|AF239989 Prunus persica ACC synthase ACS25 mRNA, pa... 597 e-170 40 gb[U22523]MIU22523 Mangifera indica 1-aminocyclopropane-carboxyl... 359 e-169 emb[X66605]DCACCS D.caryophyllus mRNA for 1-aminocyclopropane-1-... 396 e-168 gb|L31347|MAUACCSYN Malus domestica 1-aminocyclopropane-1-carbox... 378 e-168 emb|AB015624|AB015624 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 377 e-168 emb|AF049137|AF049137 Dianthus caryophyllus 1-aminocyclopropane-... 338 e-167 45 emb|AF170705|AF170705 Mangifera indica 1-aminocyclopropane-1-car... 290 e-167 gb[U03294]MSU03294 Malus sylvestris 1-aminocyclopropane-1-carbox... 372 e-166 emb|Z77854|PSPACS1 Phalaenopsis species mRNA for 1-aminocyclopro... 513 e-166 emb|AJ277160|CPA277160 Carica papaya partial paccs1A gene for 1-... 266 e-165 gb[U73816]MDU73816 Malus domestica ACC synthase (MdACS-3) mRNA, ... 505 e-164 50 emb|AB007449|AB007449 Actinidia deliciosa mRNA for 1-aminocyclop... 355 e-164 emblAB007639|AB007639 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 502 e-164 dbj|D01033|CUCACCA Cucurbita maxima mRNA for 1-aminocyclopropane... 366 e-164 emb[X87112]PCPCACS1G P.communis mRNA for 1-aminocyclopropane-1-c... 363 e-163 gb[U17972|LEU17972 Lycopersicon esculentum 1-aminocyclopropane-1... 362 e-163 55 emb|AB015495|AB015495 Passiflora edulis PE-ACS2 mRNA for ACC syn... 368 e-162 emblAB006805|AB006805 Cucumis sativus CS-ACS3 mRNA for ACC synth... 360 e-160 emb|AB000679|AB000679 Vigna radiata mRNA for 1-aminocyclopropane... 365 e-158 gb|U34987|VRU34987 Vigna radiata 1-aminocyclopropane-1-carboxyli... 325 e-157 gb[M66619]DINCARACC D.caryophyllus 1-aminocyclopropane-1-carboxy... 496 e-156 60 gb[U34986[VRU34986 Vigna radiata 1-aminocyclopropane-1-carboxyli... 362 e-156 gb[U64031]DCU64031 Dendrobium crumenatum ACC synthase gene, comp... 259 e-154

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emb|AF016458|AF016458 Pisum sativum 1-aminocyclopropane-1-carbox... 355 e-154 emb|Z27233|STACCAS1 S.tuberosum (STAC1) gene for amino cycloprop... 237 e-152 emb|Z27234|STACCAS2 S.tuberosum STACS2 gene for 1-Aminocycloprop... 232 e-152 emb|AF043122|AF043122 Lycopersicon esculentum ACC synthase (LE-A... 236 e-150 gb|U37774|CMU37774 Cucurbita maxima 1-aminocyclopropane-1-carbox... 234 e-150 gb|L34171|TOMACS3A Lycopersicon esculentum 1-aminocyclopropane-1... 235 e-149 gb|U18055|LEU18055 Lycopersicon esculentum 1-aminocyclopropane-1... 235 e-149 emb|AF151961|AF151961 Vigna radiata 1-aminocyclopropane-1-carbox... 232 e-146 emb|Z12135|VRACCSYN4 V.radiata gene for ACC synthase (pMAC-4). 10 gb|L07883|DORAMICARB Moth orchid 1-aminocyclopropane-1-carboxyla... 243 e-144 emb|AF074927|AF074927 Sinapis arvensis 1-aminocyclopropane-1-car... 330 e-142 emb|AF083815|AF083815 Antirrhinum majus ACC synthase 2 (ACS2) mR... 482 e-141 emb|AB021907|AB021907 Musa acuminata MA-ACS2 mRNA for ACC syntha... 310 e-140 emb|AB021908|AB021908 Musa acuminata MA-ACS3 mRNA for ACC syntha... 298 e-138 15 Query= AC000107.5\_at 17278\_at /id\_source genbank /description gb|aad36959.1|ac000107\_5 (ac000107) f17f8.5 [arabidopsis thaliana] /blast\_score 0 /ec\_number /family /chip nova /gb\_link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|ac000107| /ncgi 20 http://www.ncgr.org/cgi-bin/ff?ac000107 (1950 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

25

30

Searching.....done

Score E

Sequences producing significant alignments:

(bits) Value

gb|U79958|PSU79958 Pisum sativum BP-80 vacuolar sorting receptor... 630 0.0

emb|AB006809|AB006809 Cucurbita sp. mRNA for PV72, complete cds. 606 0.0 emb|AW267745|AW267745 EST305873 DSIR Medicago truncatula cDNA cl... 350 e-124 emb|AW931583|AW931583 EST357426 tomato fruit mature green, TAMU ... 165 e-112 emb|AW309187|AW309187 sg05d06.y1 Gm-c1019 Glycine max cDNA clone... 326 e-108 emb|AW622833|AW622833 EST306903 tomato flower buds 3-8 mm, Corne... 387 e-106 emb|AW774434|AW774434 EST333585 KV3 Medicago truncatula cDNA clon... 294 e-100 emb|AW689392|AW689392 NF018F12ST1F1000 Developing stem Medicago ... 338 e-100 gb|BE054150|BE054150 GA\_Ea0034H17f Gossypium arboreum 7-10 dpa ... 195 2e-99 emb|AW737948|AW737948 EST36666 tomato susceptible, Cornell Lyco... 332 6e-90 emb|AW737948|AW737948 EST339375 tomato flower buds, anthesis, Co... 225 1e-89 emb|AW615949|AW615949 EST325315 tomato flower buds 0-3 mm, Corne... 217 3e-86 emb|AW932529|AW932529 EST358372 tomato fruit mature green, TAMU ... 303 1e-83 emb|AW685785|AW685785 NF030C07NR1F1000 Nodulated root Medicago t... 157 2e-81 emb|AW747297|AW747297 WS1 67 G06 b1 A002 Water-stressed 1 (WS1) 286 4e-76

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WO 02/22675 gb|U18916|SCE9781 Saccharomyces cerevisiae chromosome V cosmids ... 35 2.8 dbj|D13340|CHL58S25S Chlorella ellipsoidea genes for 5.8S rRNA a... 35 2.8 emb|AW648383|AW648383 EST326837 tomato germinating seedlings, TA... 33 3.3 emb|AI778616|AI778616 EST259495 tomato susceptible, Cornell Lyco... 33 3.3 emb|AA749476|AA749476 L30-23M13T3 Ice plant Lambda Uni-Zap XR ex... emb|AA938798|AA938798 L0-249M13R Ice plant Lambda Uni-Zap XR exp... emb|AI822187|AI822187 L0-664T3 Ice plant Lambda Uni-Zap XR expre... 31 3.3 emb|AI822671|AI822671 L0-1192T3 Ice plant Lambda Uni-Zap XR expr... 31 3.3 10 Query= AC004684.168 s at 17744 s at /id\_source genbank/description gb|aac23646.1| (ac004684) putative alcohol dehydrogenase [arabidopsis thaliana] /blast score 1.00e-170 /ec number /family /chip nova /gb link /ncgi (885 letters) 15 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching.....done 20 Score E Sequences producing significant alignments: (bits) Value emb[X97606]MSRNAAAA M.sativa mRNA translated from abscisic activ... 306 e-115 25 emb[AW666569]AW666569 GA\_Ea0005J12 Gossypium arboreum 7-10 dpa ... 310 6e-91 gb|BE123903|BE123903 EST394028 DSIL Medicago truncatula cDNA clo... 304 7e-88 emb|AW329160|AW329160 N200369e rootphos(-) Medicago truncatula c... 224 2e-80 emb|AI794650|AI794650 sb67a05.y1 Gm-c1019 Glycine max cDNA clone... 286 2e-76

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(1233 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

15

Score

Sequences producing significant alignments:

(bits) Value

20

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- 25 emb|AW832261|AW832261 sm21g08.yl Gm-c1027 Glycine max cDNA clone... 372 e-102 emb|AW507609|AW507609 si42a06.y1 Gm-r1030 Glycine max cDNA clone... 365 e-100 emb|AI974103|AI974103 sd16e09.yl Gm-c1020 Glycine max cDNA clone... 202 9e-91 emb|AV421952|AV421952 AV421952 Lotus japonicus young plants (two... 304 1e-81 emb|AW509448|AW509448 si37c07.yl Gm-r1030 Glycine max cDNA clone... 297 4e-81
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gblaab64049.1 (ac002333) putative endochitinase [arabidopsis thaliana] /blast\_score 1.00e-161 /ec\_number /family /chip nova /gb\_link /ncgi (834 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 10 E Score Sequences producing significant alignments: (bits) Value gb[U21848]BNU21848 Brassica napus chitinase class IV (LSC222) mR... 225 e-134 15 emb|X61488|BNCHITIN B.napus mRNA for chitinase. gblU97521|VVU97521 Vitis vinifera class IV endochitinase (VvChi4... 173 3e-77 gb|U97522|VVU97522 Vitis vinifera class IV endochitinase (VvChi4... 174 2e-76 emb[X57187]PVCHITIN P.vulgaris mRNA for chitinase. emb|AF112966|AF112966 Triticum aestivum chitinase IV precursor (... 157 5e-70 20 dbj|D45181|D45181 Chenopodium amaranticolor mRNA for chitinase, ... 100 5e-67 gb[U52845]DCU52845 Daucus carota class IV chitinase EP3-1/H5 (EP... 114 7e-67 gb[U52846]DCU52846 Daucus carota class IV chitinase EP3-2/H1 (EP... 111 9e-66 gb|BE034166|BE034166 MG05H02 MG Mesembryanthemum crystallinum cD... 129 1e-65 gb[U52847]DCU52847 Daucus carota class IV chitinase EP3-3/E7 (EP... 110 8e-65 25 gb|U52848|DCU52848 Daucus carota class IV chitinase EP3B/E6 (EP3... 111 2e-64 gb|BE034975|BE034975 ML07H10 ML Mesembryanthemum crystallinum cD... 125 2e-64 gb|BE034428|BE034428 MH04G02 MH Mesembryanthemum crystallinum cD... 123 2e-63 gb|BE034976|BE034976 ML07H11 ML Mesembryanthemum crystallinum cD... 120 6e-63 dbj|D45183|D45183 Chenopodium amaranticolor mRNA for chitinase, ... 96 1e-62 30 dbj|D45184|D45184 Chenopodium amaranticolor mRNA for chitinase, ... 100 3e-62 dbi|D45182|D45182 Chenopodium amaranticolor mRNA for chitinase, ... 100 4e-60 emb[X75945]BVCH4RNA B.vulgaris Ch4 mRNA for chitinase. 92 6e-60 emb|A23392|A23392 B.vulgaris mRNA for chitinase 4 (B15). 92 6e-60 emb[X88803]VURNACHI4 V.unguiculata mRNA for chitinase clase 4 (p... 118 3e-59 emblAI897733|AI897733 EST267176 tomato ovary, TAMU Lycopersicon ... 110 1e-58 35 gb|BE034406|BE034406 MH04D10 MH Mesembryanthemum crystallinum cD... 105 2e-58 gb|L25826|BEUSP2X Sugar beet chitinase (SP2) mRNA, complete cds. 158 3e-58 gb]BE034497|BE034497 MH05H03 MH Mesembryanthemum crystallinum cD... 89 8e-58 gb|L42467|PIACHI Picea glauca chitinase (chi) mRNA, complete cds. 93 2e-57 40 emb|AF090336|AF090336 Citrus sinensis chitinase CHI1 (chi1) mRNA... 166 2e-57 emb|A1897843|A1897843 EST267286 tomato ovary, TAMU Lycopersicon ... 103 1e-56 emb|AF112963|AF112963 Triticum aestivum chitinase II precursor (... 149 9e-56 gb|BE035287|BE035287 MM04H04 MM Mesembryanthemum crystallinum cD... 162 4e-54 emb[AW680953]AW680953 WS1\_9\_A06.b1\_A002 Water-stressed 1 (WS1) S... 178 3e-52 45 gb|BE033502|BE033502 ME03F10 ME Mesembryanthemum crystallinum cD... 87 4e-50 gb[M94105]ALCCHITIN Allium sativum chitinase mRNA, 3' end. 100 6e-49 emb|AI898279|AI898279 EST267722 tomato ovary, TAMU Lycopersicon ... 83 1e-48 gb[U83592]MSU83592 Medicago sativa class I chitinase mRNA, compl... 109 1e-48 gb[U83591[MSU83591 Medicago sativa class I chitinase mRNA, compl... 109 1e-48 50 emb|AW030814|AW030814 EST274069 tomato callus, TAMU Lycopersicon... 83 3e-48 emb|AW035013|AW035013 EST279284 tomato callus, TAMU Lycopersicon... 83 3e-48 emblAW029733|AW029733 EST272988 tomato callus, TAMU Lycopersicon... 83 3e-48 gb|BE034481 |BE034481 MH05F02 MH Mesembryanthemum crystallinum cD... 162 5e-48 emb|AW924422|AW924422 WS1 69 C06.b1 A002 Water-stressed 1 (WS1) ... 140 7e-48 55 emb|AI485982|AI485982 EST244303 tomato ovary, TAMU Lycopersicon ... 83 1e-47 emb|AF061805|AF061805 Elaeagnus umbellata acidic chitinase mRNA,... 99 1e-47 emb|AW648023|AW648023 EST326477 tomato germinating seedlings, TA... 83 3e-47 gb|L37876|PEACHI2I Pisum sativum chitinase class I (chi2) gene, ... 109 3e-47 gb|BE034447|BE034447 MH05A08 MH Mesembryanthemum crystallinum cD... 87 5e-47 60 emb[X15494|STCHITIN Potato endochitinase gene (EC 3.2.1.14). emb[X07130|STCHIT Solanum tuberosum mRNA for endochitinase (EC 3... 95 6e-47

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      emb|AW031815|AW031815 EST275269 tomato callus, TAMU Lycopersicon... 31 0.42
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      emb|AW218030|AW218030 EST296745 tomato flower buds, anthesis, Co... 31 0.42
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      emb|Z50171|TBBSLARNA T.brucei brucei SLA RNA gene.
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      emb[V00694]MISC13 Saccharomyces cerevisiae mitochondrion COX/OXI... 35 3.0
      gb|L36897|YSCMTCG13 Saccharomyces cerevisiae mitochondrion oxi3 ... 35 3.0
      emb|AW933203|AW933203 EST359046 tomato fruit mature green, TAMU ... 35 3.0
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	emb AW257203 AW257203 EST305340 KV2 Medicago truncatula cDNA clo 35 4.2
5	emb AC005504 AC005504 Plasmodium falciparum chromosome 12, *** S 35 4.2
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	emb AA739776 AA739776 541 PtIFG2 Pinus taeda cDNA clone 8876M 3' 35 4.2
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	emb AA739843 AA739843 608 PtIFG2 Pinus taeda cDNA clone 8988M 3' 35 4.2
	emb AQ849884 AQ849884 LMAJFV1_lm51b04.x1 Leishmania major FV1 ra 34 5.7
15	a langua amania ni sa amana a sa sa sa sa sa sa sa sa sa sa sa sa
13	
	emb AW735973 AW735973 EST336741 tomato flower buds 0-3 mm, Corne 34 5.7
	emb AW668487 AW668487 GAEa0014C10 Gossypium arboreum 7-10 dpa 34 5.7
20	emb AL110295 SPBC106 S.pombe chromosome II cosmid c106. 34 5.7
20	gb U08622 SPU08622 Schizosaccharomyces pombe cAMP-dependent prot 34 5.7
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	emb AV407948 AV407948 AV407948 Lotus japonicus young plants (two 34 5.7
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25	emb AW719775 AW719775 LjNEST9G5r Lotus japonicus nodule library 34 5.7
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	emb AL132675 SPAC144 S.pombe chromosome I cosmid c144. 34 7.8
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30	gb B07277 B07277 G267T3 MVAT4 sheared genomic library Trypanosom 34 7.8
	emb AW982385 AW982385 HVSMEg0003C03f Hordeum vulgare pre-anthesi 34 7.8
	emb AW442327 AW442327 EST311723 tomato fruit red ripe, TAMU Lyco 34 7.8
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~ ~	emb[X15601]CMCHPSBA C.moewusii chloroplast psbA gene. 34 7.8
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	thaliana] /blast_score 0 /ec_number /family /chip nova /gb_link
40	http://www3.ncbi.nlm.nih.gov/htbin-
40	post/entrez/query?db=n&form=6&dopt=g&uid=gb af071527 /ncgi
	http://www.ncgr.org/cgi-bin/ff?af071527
	(1926 letters)
	Database: plantfungal
45	661,018 sequences; 426,114,510 total letters
	Searchingdone
	Score E
50	Score E Sequences producing significant alignments: (bits) Value
50	ocquences producing significant angiments. (ons) value
	emb AI894928 AI894928 EST264371 tomato callus, TAMU Lycopersicon 58 2e-07
	gb BE033853 BE033853 MG01B02 MG Mesembryanthemum crystallinum cD 55 2e-06
	emb AW621748 AW621748 EST312546 tomato root during/after fruit s 53 8e-06
55	emb AI729861 AI729861 BNLGHi5428 Six-day Cotton fiber Gossypium 52 2e-05
	* 1 · * * * * * * * * * * * * * * * * *
	emb AW035961 AW035961 EST282820 tomato callus, TAMU Lycopersicon 51 3e-05 emb AV420300 AV420300 AV420300 Lotus japonicus young plants (two 49 1e-04
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	emb AW694103 AW694103 NF072D04ST1F1041 Developing stem Medicago 48 3e-04
60	******
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emb|AZ124239|AZ124239 T223103b Medicago truncatula BAC library M... 43 0.003 emb|AV414054|AV414054 AV414054 Lotus japonicus young plants (two... 44 0.004 emb|AI487562|AI487562 EST245884 tomato ovary, TAMU Lycopersicon ... 44 0.005 emb|Z69731|SPAC6C3 S.pombe chromosome I cosmid c6C3. emb|AW164678|AW164678 se76b06.y1 Gm-c1023 Glycine max cDNA clone... 43 0.007 emb|AI489135|AI489135 EST247474 tomato ovary, TAMU Lycopersicon ... 43 0.007 emb|AI736949|AI736949 sb35a01.yl Gm-c1013 Glycine max cDNA clone... 42 0.017 emb|AW736367|AW736367 EST332286 KV3 Medicago truncatula cDNA clo... 42 0.017 emb|AI164848|AI164848 A069p65u Hybrid aspen plasmid library Popu... 41 0.033 10 emb|AQ638202|AQ638202 927P1-9H10.TP 927P1 Trypanosoma brucei gen... 41 0.033 emb|AW040703|AW040703 EST283567 tomato mixed elicitor, BTI Lycop... 41 0.033 emb]AI054990|AI054990 coau0002L09 Cotton Boll Abscission Zone cD... 41 0.033 emb|AI166315|AI166315 xylem.est.16 Poplar xylem Lambda ZAPII lib... 37 0.035 emb|AI055156|AI055156 coau0003D05 Cotton Boll Abscission Zone cD... 40 0.045 15 emb|AW256577|AW256577 EST304714 KV2 Medicago truncatula cDNA clo... 30 0.064 emb|AA783893|AA783893 c8g07a1.rl Aspergillus nidulans 24hr asexu... 30 0.16 emb|AW053754|AW053754 L30-1647T3 Ice plant Lambda Uni-Zap XR exp... 38 0.30 emb|AI054715|AI054715 coau0001M19 Cotton Boll Abscission Zone cD... 37 0.41 gb|BE035556|BE035556 MO09B06 MO Mesembryanthemum crystallinum cD... 29 0.69 20 gb|BE036920|BE036920 MP09B06 MP Mesembryanthemum crystallinum cD... 29 0.71 emb|AL033534|SPBC215 S.pombe chromosome II cosmid c215. emb|AW932897|AW932897 EST358740 tomato fruit mature green, TAMU ... 36 1.1 emb|AW596694|AW596694 sj15e12.y1 Gm-c1032 Glycine max cDNA clone... 35 1.5 emb[X02390]ANTRPC1 Aspergillus nidulans trpC gene. 35 2.0 emb|X52152|SCIME S. cerevisiae IME1 gene for Ime1, a meiosis ind... 35 2.0 25 emb|Z49595|SCYJR095W S.cerevisiae chromosome X reading frame ORF... gb|L47610|PIAEMB4R Picea glauca EMB4 mRNA. emb|Z49594|SCYJR094C S.cerevisiae chromosome X reading frame ORF... 35 2.0 emb|AI622956|AI622956 TENG0590 T. Cruzi epimastigote normalised ... 35 2.0 30 gb|M37188|YSCIME1 S.cerevisiae meiosis-inducing protein (IME1) g... 35 2.0 gb|BE034285|BE034285 MH02G01 MH Mesembryanthemum crystallinum cD... 35 2.8 emb|AW256576|AW256576 EST304713 KV2 Medicago truncatula cDNA clo... 35 2.8 emb|AI167045|AI167045 xylem.est.817 Poplar xylem Lambda ZAPII li... 35 2.8 emb|AW696259|AW696259 NF104F08ST1F1074 Developing stem Medicago ... 29 3.2 35 emb|AI974296|AI974296 T110298e KV0 Medicago truncatula cDNA clon... 34 3.8 emb|AI756377|AI756377 EtESTea12b12.yl Eimeria M5-6 Merozoite sta... 34 3.8 emb|AZ217340|AZ217340 Sheared DNA-117G6.TF Sheared DNA Trypanoso... 34 3.8 emb|AQ988815|AQ988815 11E1D03NE.R1 C. parvum Lambda Zap Express ... 34 3.8 emb|AW565712|AW565712 LG1 348 C03.g1 A002 Light Grown 1 (LG1) So... 34 3.8 40 gb|L20566|ASNACDP Aspergillus niger acid phospatase complete cds. emb|AW869952|AW869952 NXNV\_120\_C10\_F Nsf Xylem Normal wood Verti... 34 5.3 emb|AW691961|AW691961 NF046B02ST1F1000 Developing stem Medicago ... 34 5.3 emb|AF007946|AF007946 Trautvetteria carolinensis maturase (matK)... 34 5.3 emb|AW185605|AW185605 se79h09.yl Gm-c1023 Glycine max cDNA clone... 33 7.2 45. emb|AQ639089|AQ639089 927P1-1D11.TP 927P1 Trypanosoma brucei gen... 33 7.2 emb|AW731466|AW731466 GA\_Ea0030J17 Gossypium arboreum 7-10 dpa ... 33 7.2 emb|AW623353|AW623353 EST321298 tomato flower buds 3-8 mm, Corne... 33 7.2 emb|AI781323|AI781323 EST262202 tomato susceptible, Cornell Lyco... 33 7.2 emb|AZ214634|AZ214634 Sheared DNA-68B12.TR Sheared DNA Trypanoso... 33 7.2 50 emb|AQ656501|AQ656501 Sheared DNA-12B14.TF Sheared DNA Trypanoso... 33 7.2 emb|AW932874|AW932874 EST358717 tomato fruit mature green, TAMU ... 33 7.2 emb|AW926434|AW926434 HVSMEg0007D10 Hordeum vulgare pre-anthesis... 33 7.2 emblAQ945431|AQ945431 Sheared DNA-44E8.TR Sheared DNA Trypanosom... 33 7.2 emb|AQ945478|AQ945478 Sheared DNA-45J6.TF Sheared DNA Trypanosom... 33 7.2 55 emb|AL116771|CNS01DFV Botrytis cinerea strain T4 cDNA library un... 33 7.2 emb|AL035476|PFMAL4P3 Plasmodium falciparum chromosome 4 strain ... 33 7.2 emb|AW776850|AW776850 EST335915 DSIL Medicago truncatula cDNA cl... 33 7.2 emb|AZ219905|AZ219905 Sheared DNA-61D10.TF Sheared DNA Trypanoso... 33 7.2 emb|AZ216687|AZ216687 Sheared DNA-84G10.TR Sheared DNA Trypanoso... 33 7.2 60 emb[X87941|SCDNAGENS S.cerevisiae CRM1, YML9, PET54, SM11, PHO81... 28 7.5 emb|AZ124240|AZ124240 T223104b Medicago truncatula BAC library M... 31 7.9

emb|AI213023|AI213023 y6e12a1.fl Aspergillus nidulans 24hr asexu... 33 9.9 emb|AA556848|AA556848 690 Loblolly pine C Pinus taeda cDNA clone... 33 9.9 emb|AA965673|AA965673 m2g10a1.fl Aspergillus nidulans 24hr asexu... 33 9.9 emb|AI212224|AI212224 w9h05a1.fl Aspergillus nidulans 24hr asexu... 33 9.9 emb|AW924917|AW924917 WS1 73 B03.b1 A002 Water-stressed 1 (WS1) ... 33 9.9 emb|AW127359|AW127359 M110532 DSIL Medicago truncatula cDNA clon... 33 9.9 emb[Y15418]CCY15418 Coprinus cinereus acs-1 gene. emb|AW694961|AW694961 NF081H09ST1F1079 Developing stem Medicago ... 33 9.9 emb|AW991081|AW991081 SsS0293 Suaeda salsa ZAP cDNA library Suae... 33 9.9 10 emb|AW399343|AW399343 EST309843 L. pennellii trichome, Cornell U... 33 9.9 emb|AA784746|AA784746 g2a12a1.fl Aspergillus nidulans 24hr asexu... 33 9.9 emb|Z73017|SCYGR232W S.cerevisiae chromosome VII reading frame O... 28 10.0 Query= AC003028.196\_at 19284\_at /id\_source genbank /description 15 gb|aac27173.1| (ac003028) putative anthocyanidin synthase [arabidopsis thaliana] /blast score 0 /ec number /family /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|ac003028|/ncgi http://www.ncgr.org/cgi-bin/ff?ac003028 20 (1062 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 25 Searching......done E Score Sequences producing significant alignments: (bits) Value 30 emb|AI486803|AI486803 EST245125 tomato ovary, TAMU Lycopersicon ... 358 6e-98 emb|AI486879|AI486879 EST245201 tomato ovary, TAMU Lycopersicon ... 314 8e-85 emb|AW574302|AW574302 EST316893 GVN Medicago truncatula cDNA clo... 255 1e-76 emb|AW459665|AW459665 sh90c10.y1 Gm-c1016 Glycine max cDNA clone... 285 5e-76 emb|AI898341|AI898341 EST267784 tomato ovary, TAMU Lycopersicon ... 282 3e-75 35 gb|LA2466|PIAEFE Picea glauca ethylene-forming enzyme (EFE) mRNA... 156 4e-75 emb|AW981190|AW981190 EST392384 DSIL Medicago truncatula cDNA cl... 262 2e-73 emb|AI777799|AI777799 EST258678 tomato susceptible, Cornell Lyco... 267 1e-70 emb|AW574101|AW574101 EST316692 GVN Medicago truncatula cDNA clo... 198 2e-67 emb|AW268031|AW268031 EST306309 DSIR Medicago truncatula cDNA cl... 126 6e-65 40 emb|AW775553|AW775553 EST334618 DSIL Medicago truncatula cDNA cl... 210 3e-63 emb|AW509400|AW509400 si22f08.yl Gm-c1029 Glycine max cDNA clone... 237 8e-62 emb|AB003779|AB003779 Perilla frutescens mRNA for leucoanthocyan... 115 5e-59 emb|AW349732|AW349732 GM210006A11F12R Gm-r1021 Glycine max cDNA ... 228 6e-59 gb|U97530|PAU97530 Prunus armeniaca ethylene-forming-enzyme-like... 143 1e-58 45 emb|AF082862|AF082862 Pisum sativum unknown mRNA, partial cds. 220 2e-56 emb|Z22543|PHFLASYNA P.hybrida flavonol synthase mRNA. 99 1e-55 emb|X83229|NTRNA1A1C N.tabacum mRNA for 1-aminocyclopropane-1-ca... 176 1e-54 emblAF184273|AF184273 Daucus carota leucoanthocyanidin dioxygena... 110 9e-54 emb|AW030000|AW030000 EST273255 tomato callus, TAMU Lycopersicon... 186 1e-53 50 emb|AF026058|AF026058 Matthiola incana anthocyanidin synthase mR... 116 2e-53 emb|AF184274|AF184274 Daucus carota leucoanthocyanidin dioxygena... 110 2e-53 emb|AF117269|AF117269 Malus domestica anthocyanidin synthase (AN... 114 3e-53 emb[X71360]MSPANTHHY Malus sp. mRNA for anthocyanidin hydroxylase. 114 3e-53 emb|AW686531|AW686531 NF042D05NR1F1000 Nodulated root Medicago t... 166 2e-52 55 gb|U54566|NGU54566 Nicotiana glutinosa 1-aminocyclopropane-1-car... 174 2e-52 emb|AF001391|AF001391 Matthiola incana putative flavonol synthas... 118 3e-52 emb|AF119095|AF119095 Malus domestica flavonol synthase (FLS) mR... 91 2e-51 emb|X70786|PHFLV3HDX P.hybrida mRNA for putative flavanone 3-hyd... 112 5e-51 emb|AB012205|AB012205 Lactuca sativa Ls3h1 mRNA for gibberelin 3... 137 9e-51 60 emb|AF028602|AF028602 Ipomoea purpurea anthocyanidin synthase (A... 108 1e-50 emb|AB013101|AB013101 Lycopersicon esculentum LE-ACO4 mRNA for 1... 157 2e-50

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emb|AB023786|AB023786 Ipomoea batatas ans I mRNA for anthocyanid... 111 4e-50 emb[X75966]VVLDOX V.vinifera LDOX mRNA for leucoanthocyanidin di... 105 6e-50 emb[X04792]LEETHYBR Tomato mRNA expressed during fruit ripening ... 159 1e-49 emb|AB023787|AB023787 Ipomoea batatas ans II mRNA for anthocyani... 110 1e-49 emb[A35021]A35021 L.esculentum pTOM13. 159 le-49 emb|AF053354|AF053354 Phaseolus vulgaris 1-aminocyclopropane-1-c... 159 2e-49 gb[U06046[VRU06046 Vigna radiata clone pVR-ACO1 1-aminocycloprop... 157 3e-49 emb[X58885]LEEFEMR L.esculentum mRNA for ethylene-forming enzyme... 157 3e-49 emb|AB003514|AB003514 Actinidia deliciosa mRNA for 1-aminocyclop... 157 4e-49 10 gb[U68215]CPU68215 Carica papaya ACC oxidase mRNA, complete cds. emb|AF117270|AF117270 Malus domestica flavanone 3-hydroxylase (F... 100 5e-49 emb[X69664]MSF3HA Malus sp. mRNA for naringenin,2-oxoglutarate,3... 100 5e-49 gb[M97961]ACTACCOXI Actinidia deliciosa ACC oxidase homologue pr... 154 7e-49 emb|AB010991|AB010991 Lycopersicon esculentum Le3OH-1 mRNA for 3... 133 7e-49 15 emb|AF115262|AF115262 Trifolium repens mature green leaf 1-amino... 158 7e-49 gb[U19856]PHU19856 Pelargonium hortorum 1-aminocyclopropane-1-ca... 153 1e-48 gb|U07953|PHU07953 Pelargonium hortorum 1-aminocyclopropane-1-ca... 159 1e-48 gb|U54565|NGU54565 Nicotiana glutinosa 1-aminocyclopropane-1-car... 156 1e-48 emb|AW459318|AW459318 sh23e02.y1 Gm-c1016 Glycine max cDNA clone... 165 1e-48 20 gb|U74081|IPU74081 Ipomoea purpurea flavanone 3-hydroxylase (F3H... 99 2e-48 emb|AF254125|AF254125 Carica papaya ACC oxidase mRNA, complete cds. 150 2e-48 emb|AB012857|AB012857 Nicotiana tabacum mRNA for ACC oxidase, co... 155 2e-48 emb|AB002667|AB002667 Vigna angularis mRNA for ACC-oxidase, comp... 158 2e-48 emb|AB023789|AB023789 Ipomoea batatas f3h II mRNA for flavanone ... 96 2e-48 25 emb|AB023790|AB023790 Ipomoea batatas f3h III mRNA for flavanone... 96 2e-48 gb[U06047|VRU06047 Vigna radiata clone pVR-ACO2 1-aminocycloprop... 157 3e-48 gb|BE058138|BE058138 snl1h05.yl Gm-c1016 Glycine max cDNA clone ... 193 3e-48 emb[Y10749]BPACORNA B.pendula ACO mRNA. 150 3e-48 gb|U67861|PHU67861 Pelargonium hortorum 1-aminocyclopropane-1-ca..., 154 5e-48 30 emb[X87097]PCPCACO1G P.communis mRNA for 1-aminocyclopropane-1-c... 155 5e-48 emb|AF115263|AF115263 Trifolium repens senescent leaf 1-aminocyc... 162 5e-48 emb|AW685998|AW685998 NF032F07NR1F1000 Nodulated root Medicago t... 159 5e-48 emb|AI440611|AI440611 sa68d07.yl Gm-c1004 Glycine max cDNA clone... 192 5e-48 emb|AB023788|AB023788 Ipomoea batatas f3h I mRNA for fravanone 3... 95 6e-48 35 gb[U23066]PAU23066 Persea americana flavanone 3-hydroxylase mRNA... 100 6e-48 gblM98357|PEAAMINCAR Pea 1-aminocyclopropane-1-carboxylate oxida... 155 6e-48 emb[X98493]NTACCO Nicotiana tabacum mRNA for ACC-oxidase (clone ... 154 9e-48 emb[X81628]BOACCOX1 B.oleracea mRNA for ACC oxidase (ACC0x1). 148 le-47 gb[L35152]DINACCA Dianthus caryophyllus amino-cyclopropane carbo... 158 le-47 40 emb|AW685110|AW685110 NF025C04NR1F1000 Nodulated root Medicago t... 158 1e-47 emb|AB032198|AB032198 Nicotiana tabacum mRNA for gibberellin 3be... 135 2e-47 gb|L37103|DORCAROXI Doritaenopsis sp. 1-aminocyclopropane-1-carb... 125 2e-47 emb|AB012206|AB012206 Lactuca sativa Ls3h2 mRNA for gibberellin ... 118 2e-47 dbi|D67038|D67038 Pyrus pyrifolia mRNA for ACC oxidase, complete... 152 2e-47 45 emb[Y10034]PP1AMCSYN R.palustris mRNA for 1-aminocyclopropane-1-... 159 2e-47 gb[M62380]DINCARSR Carnation senescence related protein RNA, com... 157 2e-47 gb[M81794]MAURRP Malus sylvestris ripening-related protein mRNA,. 153 2e-47 emb[X61390]MDETR M.domestica ethylene-related mRNA. 153 2e-47 emb|AB011796|AB011796 Citrus unshiu CitFLS mRNA for flavonol syn... 104 3e-47 50 emb|AJ001646|MDACCOXI2 Malus domestica mRNA for ACC oxidase. dbj[D83041]D83041 Ipomoea nil mRNA for 2-oxogulutarate 3-dioxyge... 95 4e-47 gb|L29405|HNNACC Helianthus annuus 1-1-aminocyclopropane-1-carbo... 152 4e-47 emb|AF115261|AF115261 Trifolium repens stolon apex 1-aminocyclop... 156 4e-47 emb|AF240764|AF240764 Eustoma russellianum flavonol synthase (fl... 100 6e-47 55 emb|Z46349|NTACCDEAM N.tabacum mRNA for 1-aminocyclopropane-1-ca... 150 8e-47 gb|L07912|DORACCOXID Moth orchid ACC oxidase mRNA, complete cds. emb[X77232]PPPAO1 P.persica PAO1 mRNA for 1-aminocyclopropane-1-... 156 1e-46 emb|Z11750|BJEFEMR B.juncea mRNA for ethylene-forming enzyme. 146 1e-46 emb|AF026793|AF026793 Prunus armeniaca 1-aminocyclopropane-1-car... 156 1e-46 60 emb[X72594]MIFHT M.incana mRNA for flavanone 3-beta-hydroxylase. 94 2e-46 gb[M90294]PETEFE Petunia hybrida L. ethylene forming enzyme mRNA... 152 2e-46

emb|AB010992|AB010992 Lycopersicon esculentum Le3OH-2 mRNA for 3... 133 2e-46 emb|X75965|VVF3H V.vinifera F3H mRNA for flavanone 3-hydroxylase. 95 3e-46 emb|AB031027|AB031027 Primus mume PM-ACO1 mRNA for ACC oxidase, ... 155 4e-46 emb|AW775815|AW775815 EST334880 DSIL Medicago truncatula cDNA cl... 154 4e-46 emb|AF041479|AF041479 Rumex palustris 1-aminocyclopropane-1-carb... 154 7e-46 emb|X81629|BOACCOX2 B.oleracea mRNA for ACC oxidase (ACC0x2). 149 7e-46 emb|AW775355|AW775355 EST334420 DSIL Medicago truncatula cDNA cl... 162 8e-46

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15 (1731 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

20 Searching.....done

Score E Sequences producing significant alignments: (bits) Value 25 emb|AF023472|AF023472 Hordeum vulgare peptide transporter (ptr1)... 162 e-119 emb|AW929517|AW929517 EST338305 tomato flower buds 8 mm to pre-a... 162 3e-90 emb|AF016713|AF016713 Lycopersicon esculentum oligopeptide trans... 138 3e-89 emb|AF213936|AF213936 Prunus dulcis amino acid/peptide transport... 132 2e-87 emb[Y13862]NTENRT1 Nicotiana tabacum enr-T1 gene. 173 9e-81 30 emb|X92852|LENIT2 L.esculentum exon 1 of NIT2 gene. 140 le-80 emb|AW773639|AW773639 EST332625 KV3 Medicago truncatula cDNA clo... 298 9e-80 emb[X92853]LEMIT1 L.esculentum exon 1 of NIT1 gene. 150 9e-74 gb|U17987|BNU17987 Brassica napus RCH2 protein mRNA, complete cds. 80 5e-73 emb|AW929740|AW929740 EST354010 tomato flower buds 8 mm to pre-a... 163 3e-58 35 emb|AW278758|AW278758 sf97b05.yl Gm-c1019 Glycine max cDNA clone... 127 2e-47 emb|AW186088|AW186088 se63e01.y1 Gm-c1019 Glycine max cDNA clone... 176 4e-43 emb|AQ917114|AQ917114 T233143b Medicago truncatula BAC library M... 112 6e-42 emb|AF080545|AF080545 Nepenthes alata peptide transporter (PTR1)... 111 1e-41 emblAW677489|AW677489 DG1 8 A08.b1 A002 Dark Grown 1 (DG1) Sorgh... 126 2e-35 40 emb|AW219289|AW219289 EST301771 tomato root during/after fruit s... 144 3e-34 emb|Z69370|CSNITR1 C.sativus mRNA for nitrite transporter. 76 3e-34 emb|AW310175|AW310175 sf32c04.x1 Gm-c1028 Glycine max cDNA clone... 146 5e-34 emb|AW931623|AW931623 EST357466 tomato fruit mature green, TAMU ... 76 8e-34 emb|AW455271|AW455271 EST311931 tomato root during/after fruit s... 144 3e-33 45. emb|AW202366|AW202366 sf14b11.yl Gm-c1027 Glycine max cDNA clone... 144 3e-33 emb|AW694006|AW694006 NF071D12ST1F1101 Developing stem Medicago ... 79 7e-33 emb|AW691380|AW691380 NF040G02ST1F1000 Developing stem Medicago ... 142 1e-32 emb|AW233936|AW233936 sf32c04.y1 Gm-c1028 Glycine max cDNA clone... 82 4e-32 emb|AW774447|AW774447 EST333598 KV3 Medicago truncatula cDNA clo... 85 8e-32 50

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(1290 letters)

5

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching.....done

10

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Sequences producing significant alignments:

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Database: plantfungal 661,018 sequences; 426,114,510 total letters 60

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Searching done

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20	emb AW738043 AW738043 EST339470 tomato flower buds, anthesis, Co 43 0.005 emb AW774865 AW774865 EST334016 KV3 Medicago truncatula cDNA clo 43 0.007 emb AW832220 AW832220 sm21a01.yl Gm-c1027 Glycine max cDNA clone 38 0.011 emb AW931008 AW931008 EST356851 tomato fruit mature green, TAMU 39 0.089 emb AW396940 AW396940 sg65b05.yl Gm-c1007 Glycine max cDNA clone 39 0.089
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30	emb AW720369 AW720369 LjNEST21g3r Lotus japonicus nodule library 36 0.83 emb Z75009 SCYOR101W S.cerevisiae chromosome XV reading frame OR 35 1.6 gb U32307 SCU32307 Saccharomyces cerevisiae oligosaccharyltransf 35 1.6 emb X94335 SC130KBXV S.cerevisiae 130kb DNA fragment from chromo 35 1.6 emb Z75010 SCYOR102W S.cerevisiae chromosome XV reading frame OR 35 1.6
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55	emb AW696984 AW696984 NF111A10ST1F1072 Developing stem Medicago       33 7.6         gb L07391 NEUNIT6X Neurospora crassa nitrite reductase (nit-6) g       33 7.6         emb AQ645907 AQ645907 RPCI93-DpnII-28N21.TV RPCI93-DpnII Trypano       33 7.6
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post/entrez/query?db=n&form=6&dopt=g&uid=gb|ac005956| /ncgi http://www.ncgr.org/cgi-bin/ff?ac005956 (711 letters)

5 Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

10 Score

Sequences producing significant alignments:

(bits) Value

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      emb|AI727213|AI727213 BNLGHi7517 Six-day Cotton fiber Gossypium ... 70 2e-11
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60
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Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching.....done

5

#### Score E

Sequences producing significant alignments:

(bits) Value

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	emb AF114171 AF114171 Sorghum bicolor BAC clone 25.M18, complete 33 7.8
	emb AI900865 AI900865 sb95d08.yl Gm-c1012 Glycine max cDNA clone 33 7.8
50	emb AZ124341 AZ124341 T223084b Medicago truncatula BAC library M 33 7.8
-	omop 1224341 p 22124341 12230040 Medicago d unicaldia DAC notaty Wi 33 7.0
	Overer AI 025670 144 at 17652 at lid assume combant /description
	Query= AL035679.144 at 17653 at /id_source genbank /description
	emb cab38823.1  (al035679) putative protein [arabidopsis thaliana]
5 <i>5</i>	/blast_score 0 /ec_number /family /chip nova /gb_link
55	http://www3.ncbi.nlm.nih.gov/htbin-
	post/entrez/query?db=n&form=6&dopt=g&uid=gb al035679  /ncgi
	http://www.ncgr.org/cgi-bin/ff?al035679
	(1455 letters)

Database: plantfungal
661,018 sequences; 426,114,510 total letters

5 (bits) Value Sequences producing significant alignments: emb|AW091895|AW091895 EST285075 tomato mixed elicitor, BTI Lycop... 306 3e-82 emb|AW040183|AW040183 EST282682 tomato mixed elicitor, BTI Lycop... 306 3e-82 emb[AW776130]AW776130 EST335195 DSIL Medicago truncatula cDNA cl... 204 2e-51 10 emb|AI897609|AI897609 EST267052 tomato ovary, TAMU Lycopersicon ... 182 4e-45 emb|AI055000|AI055000 coau0002L23 Cotton Boll Abscission Zone cD... 90 1e-42 emb|AI055639|AI055639 coau0004K19 Cotton Boll Abscission Zone cD... 107 2e-22 emb|AW693198|AW693198 NF061D12ST1F1000 Developing stem Medicago ... 66 6e-10 emb|AW684256|AW684256 NF014F09NR1F1000 Nodulated root Medicago t... 43 0.007 15 emb|AW667681|AW667681 GA\_Ea0010D15 Gossypium arboreum 7-10 dpa ... 43 0.007 emb|AW736130|AW736130 EST332126 KV3 Medicago truncatula cDNA clo... 39 0.12 emb|AW683515|AW683515 NF015C12LF1F1097 Developing leaf Medicago ... 36 0.21 emb|AI781902|AI781902 EST262781 tomato susceptible, Cornell Lyco... 36 0.57 gb[U12141|SCU12141 Saccharomyces cerevisiae chromosome XIV left ... 35 0.73 20 emb|AW277626|AW277626 sf84e07.yl Gm-c1019 Glycine max cDNA clone... 36 0.79 emb|Z71330|SCYNL054W S.cerevisiae chromosome XIV reading frame O... 35 0.80 emb|AW692595|AW692595 NF057C11ST1F1000 Developing stem Medicago ... 35 1.1 emb[X54145]ANPECT A.niger gene for pectinesterase. 35 1.5 35 1.5 emb|A35008|A35008 A.niger PE gene. 25 emb|A34997|A34997 A.niger pectinesterase coding sequence. 35 1.5 emb|AQ162316|AQ162316 mgxb0012I08r CUGI Rice Blast BAC Library P... 34 2.8 emb|AF132029|AF132029 Hortonia floribunda chloroplast atpB-rbcL ... 34 2.8 emb|AW567917|AW567917 si67c02.yl Gm-r1030 Glycine max cDNA clone... 34 2.8 emb|AQ951657|AQ951657 Sheared DNA-48F5.TR Sheared DNA Trypanosom... 34 2.8 30 gb[U81520]CIU81520 Cichorium intybus sucrose:sucrose 1-fructosyl... 34 3.9 emb]AZ047925]AZ047925 LMAJFV1 lm68e11.x1 Leishmania major FV1 ra... 34 3.9 emb|AI730144|AI730144 BNLGHi6313 Six-day Cotton fiber Gossypium ... 34 3.9 emb|AQ942723|AQ942723 Sheared DNA-42J12.TR Sheared DNA Trypanoso... 34 3.9 gb[U05812|HMU05812 Herpetomonas muscarum ATCC 30261 kinetoplast ... 34 3.9 emb|AW681095|AW681095 WS1 8 B09.g1 A002 Water-stressed 1 (WS1) S... 34 3.9 emb|AL049180|PFMAL13P1 Plasmodium falciparum chromosome 13 strai... 27 4.9 emb|AW832107|AW832107 sm30e05.y1 Gm-c1028 Glycine max cDNA clone... 33 5.3 gb[N82224]N82224 TgESTzy37d01.rl TgRH Tachyzoite cDNA Toxoplasma... 33 5.3 emb|A94222|A94222 Sequence 5 from Patent EP0952222. 40 emblAA741851\AA741851 LmLv39p3/584B Leishmania major promastigot... 33 5.3 emb[AW728096]AW728096 GA Ea0029O02 Gossypium arboreum 7-10 dpa ... 33 7.3 emb|AW278032|AW278032 sf89g07.yl Gm-c1019 Glycine max cDNA clone... 33 7.3 gb[N82117]N82117 TgESTzy36b06.r1 TgRH Tachyzoite cDNA Toxoplasma... 33 7.3 emb|AL355930|NCB2O8 Neurospora crassa DNA linkage group II BAC c... 32 10.0 45. emb|AW688738|AW688738 NF011A11ST1F1000 Developing stem Medicago ... 32 10.0 emb|AI781529|AI781529 EST262408 tomato susceptible, Cornell Lyco... 32 10.0 emb|AI397765|AI397765 NCC5A11T7 Conidial Neurospora crassa cDNA ... 32 10.0 emb|AF127239|AF127239 Nicotiana tabacum cultivar Burley 21 argin... 32 10.0 emb[AW180260]AW180260 MgA0351f MgA Library Mycosphaerella gramin... 32 10.0 50 emb|AF127240|AF127240 Nicotiana tabacum cultivar Xanthi arginine... 32 10.0 gb|BE055666|BE055666 GA Ea0009H20f Gossypium arboreum 7-10 dpa ... 32 10.0 gb|BE036548|BE036548 MP01C02 MP Mesembryanthemum crystallinum cD... 32 10.0 emb|AW038750|AW038750 EST280611 tomato mixed elicitor, BTI Lycop... 32 10.0 emb|AL133484|LMFL236 Leishmania major Friedlin chromosome 19 cos... 32 10.0 gb|BE123936|BE123936 EST394061 DSIL Medicago truncatula cDNA clo... 32 10.0

# Example 4

# Immediate Early Transcriptional Responses Controlled by a Salicylic Acid Dependent Local Resistance Pathway

Salicylic Acid (SA) is an important mediator of local and systemic

defense responses. In Arabidopsis, accumulation of SA is essential for local resistance against many pathogens including Peronospora parasitica (Peronospora) isolates. Furthermore, numerous defense-related genes can be activated by SA treatment. In addition to local resistance, SA-accumulation is required for systemic acquired resistance (SAR). In the SAR pathway, SA has been shown to act upstream of NPRI, which is also essential for SAR. Short application of the SA analog BTH (e.g., 4 hours) to plant tissue appears to simulate local defense responses to Peronospora parasitica, whereas sustained SA treatment (e.g., 48 hours) elicits SAR-related plant responses (Maleck et al., 2000).

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A previous cDNA microarray study identified a cluster of roughly 30 coregulated genes that appear to be specifically involved in resistance of the *Arabidopsis* ecotype WS against the *Peronospora* isolate Noco2 (Maleck et al., 2000). These genes are strongly activated in response to infection with Noco2 (incompatible interaction) and 4 hours after BTH treatment. However, these genes were not markedly activated during a compatible interaction with the *Peronospora* isolate Emwal or an incompatible interaction with *Pseudomonas syringaea* (DC3000 avrRpt2) bacteria. BTH treatment for 48 hours (that simulates SAR) even strongly repressed these genes. Thus, a rapid and transient SA peak may control Noco2 triggered defense genes.

Resistance of the Arabidopsis ecotype Col-0 against the *Peronospora* isolate Emoy2 appears to be regulated by a similar pathway that also is dependent on accumulation of SA. Emoy2 resistance in Col-0 is controlled by the resistance gene *RPP4*. To further explore early SA-dependent gene regulatory events and to relate these to *Peronospora*-induced resistance responses, immediate-early target genes of SA were identified using Affymetrix chip experiments with the protein biosynthesis inhibitor cycloheximide (CHX). Genes of this category likely are linked to transcriptional regulators acting closely downstream of SA. Results from these "SA-chip" experiments were combined with those from a set of chip experiments examining expression

profiles triggered in the *Arabidopsis* ecotype Col-0 by the *Peronospora* isolate Emoy2 to identify clusters of co-regulated *RPP4* controlled SA-dependent immediate early genes.

# Results

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To identify genes controlled by the RPP4 pathway, interactions between the Peronospora isolate Emoy2 and Col-0 wild type plants, the defense mutants ndr1, npr1 and pad4 as well as transgenic NahG plants were examined by chip experiments. The interactions between Emoy2 and Col-0, ndr1 or npr1 are incompatible (plant is disease resistant), whereas the interactions involving pad4 and NahG are compatible (plant is susceptible, i.e., disease develops). For each interaction three different time points were analyzed: 0, 12 and 48 hours post infection with 50,000 Emoy2 spores/ml. Genes that were at one or more time points more strongly expressed in Col-0, ndr1 and npr1 as compared to pad4 and NahG were considered as controlled by the RPP4 pathway. Genes represented by 271 probe sets showed at least one 2-fold expression difference in comparison between Col-0 and pad4 or NahG and were selected for further analysis.

To identify immediate early target genes of SA, Col-0 wt seedlings were pretreated with CHX or mock treated 15 minutes before spraying with 2 mM SA or water and harvested 2 hours later (Table 28).

	Table 28	
	<u>Plants</u>	<b>Treatment</b>
	Col-0	untreated (mock)
25	Col-0	+SA
	Col-0	+SA+CHX
	Col-0	+ CHX

271 probe sets were found to be upregulated in a *RPP4* pathway-specific manner (SEQ ID NOs: 400-684). Emoy2-induced upregulation of these genes is compromised in pad4 and *NahG* plants. These RPP4 controlled genes were further analyzed concerning their responses to SA, CHX and SA + CHX. The 271 probe sets were subcategorized by K-means clustering over the SA/CHX data set. Nine K-means clusters comprised three subgraphs. For each of these subgraphs, the first data point represented the untreated control value, the second data point represented the response to combined SA/CHX treatment, the third

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data point represented CHX treatment alone and the fourth data point represented SA treatment alone. K-means clustering defined two categories of RPP4 controlled CHX/SA-responsive genes: immediate early genes (SA-responsive, not CHX affected) and super-induced genes (additive effects of SA and CHX).

Two K-means clusters represent RPP4-controlled SA-induced immediate early genes (Table 29; SEQ ID NOs: 150, 159, 117, 126, 208, 428, 426, 436, 430, 434, 478, 641, 609, 615 and 526). RPP4-controlled upregulation of these genes is SA dependent and SA induction is not compromised by CHX, which blocks or reduces protein biosynthesis. Thus, all essential regulators that mediate SA-responsiveness of these genes must be already preformed and, hence, are likely to act closely downstream of SA. This set of 15 genes includes seven potential regulator genes that may be involved in the control of secondary response genes (which require protein biosynthesis) or other late responses.

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Table 29

gb|AAC23641.1| (AC004684) putative receptor-like protein 12354 g at kinase . 14978 at gb|AAB64024.1| (AC002333) putative glucosyltransferase 20 15479 at emb|CAB39671.1| (AL049483) putative protein 15616 s at emb|CAA08794.1| (AJ009696) wall-associated kinase 1 gb|AAB97145.1| (AF000977) MEK1 16061 s at 16109 s at gb|AAC05342.1| (AC002521) putative protein kinase 25 16140 s\_at emb|CAB42872.1| (AJ012423) wall-associated kinase 2 16603 s at gb|AAB58497.1| (U81293) UDP-glucose:indole-3-acetate beta-D-glucosyltransferase 17499 s at gb|AAD19610.1| (AF107726) cyclic nucleotide gated channel 17930 s at emb|CAA07352.1| (AJ006960) peroxidase 30 18966 at gb|AAC95196.1| (AC004561) putative glutathione S-transferase emb|CAB10219.1| (Z97336) hypothetical protein - weak 20429 s at similarity to NF-kappa-B emb|CAB41928.1| (AL049751) short-chain alcohol 20685 at dehydrogenase like protein emb|CAA19683.1| (AL024486) putative protein 35 13702 s at gb|AAD15461.1| (AC006067) unknown protein 14704 s at To further subcategorize these genes and to more precisely define a cluster of strictly co-regulated genes, the set of 15 genes was analyzed by Kmeans clustering over the RPP4 data set (Emoy2 infections). This allows to use 40 RPP4 triggered expression timing as an additional criterion for subclustering.

Normalized absolute expression levels at 0, 12 and 48 hpi in Emoy2 infected

Col-0 (w.t.), ndr1, npr1, pad4 and NahG plants are displayed. Genes that share a pronounced almost linear and sustained expression increase were chosen for further analysis. These 5 genes (Table 30, which show a pronounced Emoy2-induced SA-dependent but npr1-independent linear expression increase and which encode potential regulators) are upregulated by the RPP4 pathway, respond to Emoy2 recognition with a steep, sustained, almost linear expression increase. This pronounced "upregulation" is SA dependent and independent of de novo protein biosynthesis. Transcription factors that mediate this response must be present when SA is perceived and may act closely downstream of SA.

The mechanism that switches these genes on may involve posttranslational modifications of such preformed transcription factors.

T	a	bl	e	·3	0

12354\_g\_at gb|AAC23641.1| (AC004684) putative receptor-like protein kinase

15616\_s\_at emb|CAA08794.1| (AJ009696) wall-associated kinase 1

16140\_s\_at emb|CAB42872.1| (AJ012423) wall-associated kinase 2

17499\_s\_at gb|AAD19610.1| (AF107726) cyclic nucleotide gated channel 20429\_s\_at emb|CAB10219.1| (Z97336) hypothetical protein - weak similarity to NF-kappa-B

To identify the potential binding sites of such key transcription factors, the promoters of these five genes were searched for conserved compact sequence motifs (Table 31; SEQ ID NOs: 757-764; SEQ ID NO:765 is a consensus sequence). Using the program "AlignACE", a strikingly conserved motif was found that is present in all five promoters. This motif resembles the described consensus binding site of WRKY transcription factors, W box. However, the W box core motif, TGAC, is followed by an "A". The permutation TGACA has never been associated with WRKY binding. In addition to this, four more positions are highly conserved. This motif may be the specific binding site of an unconventional WRKY transcription factor or a factor of a novel so far unknown type. Yeast one hybrid screens can reveal the identity of this factor.

### Table 31

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W box-like Motif

35 MAP Score: 4.4

ACAGTGACA 0 391 1 (12345) SEQ ID NO:757

ACAGTGACA 1 236 1 (16140) SEQ ID NO:758

	ACAGTGACA 1 317 1	(16140)	<b>SEQ ID NO:759</b>
	ACAGTGACA 2 281 1	(15616)	<b>SEQ ID NO:760</b>
	ACAGTAACA 3 84 1	(17498)	<b>SEQ ID NO:761</b>
	AAAGTAACA 3 1557 0	(17498)	<b>SEQ ID NO:762</b>
5	AAAGTGACA 4 1840 0	(20429)	<b>SEQ ID NO:763</b>
•	AAAGTGACA 4 2131 0 *********	(20429)	SEQ ID NO:764
	ACAGTGACA A		SEQ ID NO:765

10 Expt. Frequency: 0.02/1kb Obs. in SOMc7: 0.06/lkb Obs. in this cluster: 0.50/1kb

15 One K-means cluster of the 271 RPP4 controlled probe sets represents RPP4-controlled SA/CHX super-induced genes (Table 32; SEQ ID NOs: 214, 621, 71, 439, 78, 80, 264, 265, 613, 583, 594, 616, 355, 503, 168, 447, and 551). RPP4-controlled upregulation of these genes is SA-dependent; these genes are weakly inducible upon both SA and CHX treatment alone. The response to 20 combined treatment with SA and CHX is stronger than the response to either single treatment. Induction of genes in response to CHX alone has been described before and has been attributed to hypothetical proteins that are rapidly turned over and either repress transcription of the respective gene or control degradation of the respective mRNA. Block of synthesis of such hypothetical 25 proteins may lead to increased mRNA levels by either increased transcription or reduced mRNA degradation. Like the set of immediate early SA responsive genes shown before, genes of this set also appear not to require de novo protein biosynthesis for their response to SA. In contrast to the previous gene set, however, the observed CHX-induced up-regulation may point to a de-repression 30 mechanism.

#### Table 32

	12505_s_at	gb AAC63643.1  (AC005309) putative CONSTANS-like B-box
	•	zinc finger protein
35	13656_at	gb AAD22649.1 AC007138_13 (AC007138) predicted protein of
	•	unknown function
	14116 at	gb AAC26243.1  (AF077407) contains similarity to sugar
	_	transporters
	14170 at	gb AAF29406.1 AC022354_5 (AC022354) unknown protein
40	14223 at	emblCAA19683.11 (AL024486) putative protein

```
gb|AAD31062.1|AC007357 11 (AC007357) Strong similarity to
     14248 at
                   gb|X97864 cytochrome P450
     14608 at
                   gb|AAD31074.1|AC007357 23 (AC007357) Similar to
                   gb|AF038007 FIC1, member of the PF|00122 E1-E2 ATPase
 5
                   family.
     14614 at
                   gb|AAC16958.1| (AC004165) putative glucosyltransferase
     15646 s at
                   gb|AAC37474.1| (L42212) serine acetyltransferase
     16054 s at
                   emb|CAA74639.1| (Y14251) glutathione S-transferase
     16105_s_at
                   gb|AAC31756.1| (U68017) heat shock transcription factor 4
10
     16968 at
                   emb|CAA17559.1| (AL021961) glucosyltransferase -like protein
                   gb|AAB61480.1| (AC000348) T7N9.4
     18235 at
     18567 at
                   gb|AAC34217.1| (AC004411) putative alcohol dehydrogenase
     18591 at
                   emb|CAA52772.1| (X74756) ATAF2
     19845 g at
                   emb|CAB37510.1| (AL035540) monooxygenase 2 (MO2)
15
     20017 at
                   gb[AAC16079.1] (AC004521) unknown protein
            To further subcategorize these genes based on RPP4 triggered expression
     timing, the set of 17 SA/CHX super induced genes was K-means clustered over
     the RPP4 (Emoy2) data set (Table 33; SEQ ID NOs: 214, 621, 71, 439, 78, 80,
     264, 265, 613, 583, 594, 616, 355, 503, 168, 447, and 551). A group of five
20
     genes was identified that consistently respond quickly ("fast", within 12 hpi) and
     that have elevated expression ground states in the npr1 mutant. The fast
     response is SA dependent (is abolished or attenuated in pad4 and NahG). The
     elevated expression ground states in npr1 together with the observed CHX
     inducibility may point to an NPR1-dependent repression mechanism. Since
25
     NPRI has been shown previously to act closely at the level of transcriptional
```

NPR1 has been shown previously to act closely at the level of transcriptional regulation, a NPR1 dependent repressor may shut expression of these genes down. Upon CHX treatment this hypothetical repressor is not synthesized anymore and expression of these genes is elevated. This repressor may also participate in RPP4 triggered activation of these genes and allow Emoy2-

30 induced de-repression.

#### Table 33

	18591_at	emb CAA52772.1  (X74756) ATAF2
	14248_at	gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to
35		gb X97864 cytochrome P450
	14614_at	gb AAC16958.1  (AC004165) putative glucosyltransferase
	15646_s_at	gb AAC37474.1  (LA2212) serine acetyltransferase
	16105_s_at	gb AAC31756.1  (U68017) heat shock transcription factor 4

To identify binding sites of activating or repressing factors responsible for the co-regulation of this small cluster, the promoters of these genes were searched for conserved compact motifs with "AlignACE" (Table 34; SEQ ID NO: 766-772, SEQ ID NO: 773 is a consensus sequence). One strictly

5 conserved octameric motif was found, which, however, is absent in one of the promoters. Yeast one hybrid screens can identify factors binding to this motif.

# Table 34

Random20:

SOMc3:

SOMc1:

25

0.00/lkb

0.10/1kb

0.10/lkb

Motif 15			<i>y</i>
MAP Score: 1.23758			•
AATCGAA	T 0 40 0	(18591)	<b>SEQ ID NO:766</b>
AATCGAA	T 0 1741 1	(18591)	<b>SEQ ID NO:767</b>
AATCGAA	T 1 386 1	(PAD3, 14248)	<b>SEQ ID NO:768</b>
AATCGAA	T 2 334 1	(14614)	SEQ ID NO:769
AATCGAA	T 2 660 1	(14614)	<b>SEQ ID NO:770</b>
AATCGAAT 2 2105 1		(14614)	<b>SEQ ID NO:771</b>
AATCGAAT 3 1570 1 (15646)		(15646)	SEQ ID NO:772
******	*		
AATCGAAT		<b>SEQ ID NO:773</b>	
•	•	•	
expt.:	0.08/lkb	·	
obs.:	0.70/lkb		,
SOMc7:	0.06/1kb	,	•
	MAP Score AATCGAA AATCGAA AATCGAA AATCGAA AATCGAA AATCGAA AATCGAA AATCGAA expt.: obs.:	MAP Score: 1.23758 AATCGAAT 0 40 0 AATCGAAT 0 1741 1 AATCGAAT 1 386 1  AATCGAAT 2 334 1 AATCGAAT 2 660 1 AATCGAAT 2 2105 1 AATCGAAT 3 1570 1 ***********  AATCGAAT  expt.: 0.08/lkb obs.: 0.70/lkb	MAP Score: 1.23758  AATCGAAT 0 40 0 (18591)  AATCGAAT 1 386 1 (PAD3, 14248)  AATCGAAT 2 334 1 (14614)  AATCGAAT 2 660 1 (14614)  AATCGAAT 2 2105 1 (14614)  AATCGAAT 3 1570 1 (15646)  *************  AATCGAAT  expt.: 0.08/lkb obs.: 0.70/lkb

As described hereinbelow, sets of genes specifically controlled by the RPP7 and RPP8 pathways were defined. Similarly a set of RPP4 pathway

30 controlled genes was defined (see Figure 4). Figure 4 shows a "Venn diagram" including only the transcription factor genes of these RPP4, RPP7 and RPP8 controlled genes. Three transcription factor genes were found to be specifically upregulated by each of the three Peronospora defense pathways: AtERF1, HSF4, ATAF2. Furthermore, ERF transcription factors appear to play an important role in controlling defense responses directed against Peronospora in general. Table 35 gives four potential target genes of ERF transcription factors. These genes contain ERF binding motifs in their promoters.

## Table 35

40 Genes with potential ERF binding sites (GCCGCC or GCCGAC):

thioredoxin (13189) RPP7/RPP8 and (weakly) RPP4 controlled C2H2 zinc finger (15665) RPP7/RPP4 and (weakly) RPP8 controlled SigA binding protein (14148) RPP7/RPP4 controlled HSP70 (13284) RPP7/RPP8 controlled

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These genes are specifically controlled by at least two different *Peronospora* defense pathways and show an almost linear increase of transcript levels during incompatible Hiks1 interactions (K-means clusters 3+5).

. 6 potential ERF binding sites in K-means cluster 3+5: 0.5 motifs /

10 1kbExpected: 0.1 motif / 1kb; Enrichment in K-means cluster 3+5: 5-fold

**SEQ ID NO:789** 

MPTSATAVAPSTGSVQKKDQDWRAILSPEQFRVLREKGTENRGKGEYT KLFDDGIYSCAGCATPLYKSTTKFDSGCGWPSFFDAIPGAIKQTDMFGSN

15 AADGSIVTSGLDYILISINEKLKAYT

#### **SEQ ID NO:790**

- 1 tgcattcttt tgaggggttt aattttctgc atagctttgt ctaatctctt agagctcaat
- 61 aagagaagat ggatgttcca cggccagctt tcaaatgttt tgatgacgat ggccggctta
- 20 121 aacgttcagg gacggtttgg accgcgagtg cgcatatcat aaccgccgtg attggatctg
  - 181 gtgttctatc gcttgcgtgg gctataggtc aactcggttg gatcgcaggt cctacagtga
  - 241 tgttgttgtt ctcttttgtc acttactact cttccacgct tcttagcgac tgctacagaa
  - 301 ccggagatec tgtetetggg aagagaaact atacttacat ggacgetgte cgatcaatec
  - 361 taggtggett taggttcaag atttgtggge tgattcagta tttgaatetg tttggtatca
- 25 421 cggtcgggta cacaatcgca gcatctataa gtatgatggc gatcaagagg tccaactgtt
  - 481 tecaegagag eggagggaaa aaccegtgte acatgtegag caatecatae atgateatgt
    - 541 ttggtgtgac cgagatettg eteteteaga teaaagattt tgaccagatt tggtggetet
    - 601 ccattgtcgc tgctatcatg tccttcacat actctgcaat cggtttagct ctcggaatca
    - 661 ttcaagtcgc ggcaaatgga gttgtcaagg gaagtctcac cggaattagc atcggcgcag
- 30 721 tgactcagac ccaaaaaata tggagaacct ttcaagcact tggagacatt gcctttgctt
  - 781 atteatacte tgttgttett attgaaatte aggacactgt aagateteea eeageagaat
  - 841 caaaaacgat gaagategee acaagaatea geategetgt tacaacgaea ttttacatge
  - 901 tatgtggttg tatgggctat gcggccttcg gagataaagc accgggaaac ctcttaaccg
  - 961 gttttggttt ctacaatccg ttttggctcc ttgacgtggc taacgctgcc atagttatcc
- 35 1021 accttgtagg agettateaa gtetttgete ageceatett egeetttatt gagaaacaac
  - 1081 tggccgctag gtttcccgac agtgacttgg tgaccaagga atacgaaatc cgaatccctg
  - 1141 gttttaggtc accgtacaaa gtcaacgttt tcagagcagt ttaccgaagc gggtttgtgg
  - 1201 ttttgaccac tgtgatatcc atgcttatgc cgtttttcaa cgacgtcgta gggattttag
  - 1261 gtgcgttagg gttttggcct ttgacggttt actttccggt ggagatgtat ataagacaga
- 40 1321 ggaaggttga gagatggagt atgaagtggg tttgtctgca gatgttgagc tgtggttgtt
  - 1381 tgatgatcac gttggtcgcc ggagttggct ccatcgccgg agtaatgcta gaccttaagg
  - 1441 tttacaagcc gttcaagact acttactaaa caaaccatga tgatagatga agaagaagaa
  - 1501 ggtggtggag aaaaaaaaca aaagatataa attttaatga tgattttcat tggggaaatg
  - 1561 tanatastat anno citati antitantat antitutat ilia anti-tati antitutat ilia anti-tati della anti-

1561 tgaataatgt aaaagttett egtttegtat aatttttate ttgegtaatt tatataeat

45

### **SEQ ID NO:791**

---WO-02/22675 PCT/US01/28506

MVKNLKVDPLAKVTASTTSMVIKILSSLFITDDSYVLVSAKENKNQSEAE PSYYETLETYQGLPCPYGGYYGYYYPGLDGSVGEAKDNGYYGYGTEVQ YPVMQGENGSVIYLMPGFQSYDASQTYMPINPVGVSSQALHSPMYAAQ GYYQNQFGYADVSSPTYLWDPVGDRYVYGVASYTPPLKQNISSSSHNH

- 5 NNYYSKSKNSFTGHGMGDRPKTPRKASQNSYAPPPLLNQEKGRIAYPM DPVKKKSGALNRDETEKAKARTKENGTSMNDLANGQDHITNGECESCS LDAEGNERSNGVGSVIRRDQYNLPSFQTKYEEAIFFVIKSYSEDDIHKSIK YNVWSSTLNGNKKLDSAYQESQKKAADKSGKCPVFLFFSVNASGQFCG VAEMIGRVDYEKSMEFWQQDKWTGYFPVKWHIIKDVPNPQLRHIILEN
- 10 NENKPVTNSRDTQEVRLPQGNEVLNIFKNYAAKTSILDDFDFYENREKV MVQKKLRFPPVLKKKEEDLVADFKTMEMSNTVEEGNTELTGTVS

# **SEQ ID NO:792**

- 1 cgatcacgga tetggettgg tteatacaaa accgeegttg eegeggeaeg ggeetaegat
- 15 61 acceptigg tittactiacy tegic tetre gegagactea attracetiga agaggietti
  - 121 aaggatggaa acggcggtga aggcttagga ggagatatgt ctccgacgtt gatacggaag
  - 181 aaggeggetg aggtgggage tagagtegae geagagttge ggttagagaa taggatggtt
  - 241 gagaacttag acatgaataa gttgccggag gcatatggat tgtaatttat agtttggtag
  - 301 tttataggtt ggagattgcc cggagacaga gtcaaacaga ggttctctga ctcatatgag
- 20 361 gcataatata gttaatatag taatttttgt tttgagcata gtaattatgt cataacc

### **SEQ ID NO:793**

- 1 gggcaatgat tattcgttcg ccggaaccag aagtcaaaat tttggtagat agggatccca
- 61 taaaaacttc tttcgaggaa tgggctaaac ccggtcattt ctcaagaaca atagctaagg
- 25 121 gacctgatac taccacttgg atctggaacc tacatgctga tgctcacgat tttgatagtc
  - 181 ataccagtga tttggaggaa atctctcgaa aagtatttag tggccatttc ggccaactct
  - 241 ctatcatctt tctttggctg agtggcatgt atttccatgg tgctcgtttt tccaattatg
  - 301 aagcatggct gagtgateet acteacattg gacetagtge teaggtggtt tggceaatag
  - 361 tgggccaaga aatcctgaat ggagtagtgg gcggaggctt ccgaggaata caaataacct
- 30 421 caggettitt teagattigg egageateeg gaataactag tgaattacaa etttattgta
  - 481 ccgcaattgg cgcattggtc ttcgcagcct taatgctttt tgctggttgg ttccattatc
  - 541 acaaagcagc tecaaaattg gettggttee aagatgtaga atetatgttg aateaccatt
  - 601 tagcaggget actaggactt gggtcccttt cttgggcagg acatcaagta catgtatett
  - 661 atccgattaa ccaatttcta aacgctggag tagatcctaa agaaataccg cttcctcatg
- 35 721 aatttatett gaategggat ettttggete aactttatee aagttttget gaaggageaa
  - 781 ctcccttttt taccttaaat tggtcaaaat actcggaatt tcttactttt cgtggcggat
  - 841 tagatecagt gactgggggt ctatggttaa cegatatage acateateat ttacgtateg
  - 901 caattettit tetaategeg ggteatatgt ataggaceaa etggggtatt ggteatggta
  - 961 taaaagatat tttagaggct cataaaggtc catttacagg ccaaggccat aaaggcctat
- 40 1021 atgaaattct aacaacatca ttgccc

#### **SEO ID NO:794**

- MEGSSSSSSLISKSDAELEEMLDRMLTRLALCDDSKLESLVSNLLPLTISS LSSQSPVVRNKAMCVDFIFQVLEILSHVNKRVKHQHEIGLPLLALWKLY
- TDPAAAPMVRNFAIVYVEMAFERAPAKVIGECHASKISDDVSAKYRSLIT SQDKDLFLDFCLHMLLYQPSSQGGGSSPGLSVFQVNRIIGKQALKGDTLT RRKLPSNTFLTKNYHFLKINQFLKQLGILNVIGNMDLPGESVYPLYIAAS VDRVGMDLLVKIHSSQEPVAKRGEELLKKIASGTNLDDPKLINRLFLLFN GTTGTENVAPEHNVAPGNISLKMKLMSGFCRSIAAANSFPATLQCIFGC
- 50 MYDILFLLNLTFREKTEMAVRLFDALKLETQSLRSTIQEAIVSLAAAYKD

SPENILRDLEVLLLANSLAEQNEARFCALRWATSLYNSHHCPSLYICMLS AADPKLDIREIALEGLFLKEEGRSIVSNHDHKYPKFIEMLEYILKOOPKLL DSSEMRSQKLLFPSQVYLVMIKFLVKCFELEMEESNTQAVGTEFLDSAQ KMCSLLEHSLAFEGSAELHACASKALVSVGSYLPEMVELYFSRKIVWLR 5 SLLSHTDLSTRESVSRLLGMASCALSDAESCSLLSELISSISQPQKLRFEAQ HGGLCAVGFVSAHCLHRIPTVSKAVTQNAVKYLVEVVNLETAPLASVA MEALGHIGICGALPFLVNDSSPGTQVLEILQERLSKLLSGDDIKSVQKIAL SLGHICSNETSSSHLKIALDLLFSLSRSKAEEILFAAGEALSFLWGGVPVT ADMILKTNYTSLSTDSNFLMKEVKSLSDVKTDTEEDSRTTTRETITGKLF 10 DTLLYSSRKEERCAGTVWMLSLTMYCGQQPSIQLMLPKIQEAFSHLLGD **ONELTQELASQGMSIIYELGDASMKKSLVDALVNTLTGTSKRKRAIKLV** EESEVFQEGTIGESPSGGKISTYKELCNLANEMGQPDLIYKFMDLANHQA SLNSKRGAAFGFSKIAKQAGDALRPHLRLLIPRLIRYQYDPDKNVQDAM AHIWKALIQDPKKAVDEHLNHIFDDLLVQCGSRLWRSREASCLALADIIQ 15 GRKFDQVKEHLKKLWIAAFRAMDDIKETVRNAGDKLCRAVTSLTIRICD VTLTELADAKOAMDIVLPFLLSEGIMSKVNSVRKASIGVVMKLAKFYSK HEIINQFELLASFGELNVLLGNIYDPLSDFVLFVTVCRLHAANIGIETEKLE , NLRISISKGSPMWETLDLCINIVDIESLEOLIPRLTOLVRGGVGLNTRVGV ASFISLLVQKVGSEIKPFTGMLLRLLFPVAKEEKSSAAKRAFSSACGIVLK 20 YSSPSQAQSLIEETAALHSGDRSSQIACASLFKSFSSTAADIMSSHQSAIVP AIFISRFEDEKOISSLFEEVWEDITSGERVTLOLFLOEIVNHICESITSSSRFK LSFSLGKDALLDALGALSVACHEAITKEDPTTPTTILSLICSACKKKLKKY RESAFSCLEKVIIAFGDPKFFHAVFPMLYEMCNTASIKTNTQVQAASDAV KTESENGEDGHVPLEKIMECVKSCIQVATIDDILSAKADLIHVLIISLSPGF

## **SEQ ID NO:795**

KSEEAKSLLRKSRDALANLPSLN

25

30 MDKETEILSRLAANHLHLAQFEPLKATLLALRVRNPDLALTILQTIVSNA GRFDNVLWSRSCPSPSLLSFLSTIELLRFENPTSPWGFDSETLSLRADFLL MVQVLIDRVTERIKEDEESEDENSGLGNCLRVLQGVLELGVERLKFVVD TSSSEGSNKIEEDAVVSLRSIVLDYSDVFDALCCNIQRQLAGCESYGTCL VEEVQGEEQRKEMNEATCIGSPELDNINVFALIQRNVQLAQLDAMKTKL

LWTVKMSGISCVGKLCSRFPSLWTDSMDDLSPSDATKFVHELFHSLVPK LLECIHTVKIAQFHVAASQCLLELIELYSTISSLHPVEVDFKAEVVSLLELE

- 35 DEGDERGAADRIRYLHLDYGVEKENYHAVLKALLSRVMEKKDEYGDS
  WHMVRQNLLFMYKEALSSNCGDLVQMIQGIQDDMLLPHSQLHLSLDNE
  QIPLPLECFRRYLVDLKTERNIEDKSSPMSRAINSCLRDMYHYARISGSHV
  LECVMCAALSSVKKEKLQEANDVLTLFPRLRPLVASMGWDLLPGKTAT
  RRKLMRLLWTSDSQALRLEESSLYGNQTDELELASFAACVNSGKSWTP
- 40 KASFLMHGNVSSAHDDAEVDPFVENLVLERLSAQSPLRVLFDVVPGIKF QDAISLISMQPIASTAEAWKRIEDIELMHMRYALEAIVLALGAMEEAMK DETDASHRVVFYHLKDLTNHLEAIKNVPRKIMMVNIVISLLHIDDIRLSST QSASSACFSEKSNTPGLDPGDLGTEGEKEIVISFTKQLLDVLRRNLPSHPIE QECQLDGNYSTDGRQALEWRVSMAKRFIEDCEWRLSVMQHLLPLSERQ
- 45 WGLKEVLSILRAAPEKLLNLCMQRAKYDIGEEAVNRFALSAEDKATLEL AEWVDNAFKGTLVEDVMSRTAEGAAAVQDLDFHSLGSQLSPLAMVLLF AQSQVMLSEIYPGGAPKVGFTYWDQVHEVAIISVLRRILKRLQEFLEQDD PQILQASFSGDTIISSCTESHRQGQKDRALAMLHQMIEDAHRGKRQFLSG KLHNLARALADEKPEVDVLKGDGSDMAVEKDGVLGLGLKYTKQSPGS
- 50 ANRAVDGNPVSHETEDKGKKSFGPLSNKTSTYLSQFILYTAAIGDIVDGT

DTTHDFNFFSLVYEWPKDLLTRLVFDRSSTDAAAKVAEVMSADFVHEVI SACVPPVYPPRSGHGWACIPVIPTTPCSHSEGKVLSPSIEAKPNCYVRSSA TPGVPLYPLOLDVIRHLVKISPVRAVLACVFGGSILYNGSDSIISSSLNDEF PSSPDADRLFYEFSLDQSERYPTLNRWIQMQTNLHRVSEFVVTPKQKPD DTRIKPDERTGIKRLLEHDSDSESDTEETFSKNNIQPALTDGSARDGGSFE NGVCRTDPTVFLSFDWENEVPYEKAVNRLIDEGKLMDALALSDRFLRN GASDWLLQLLIKSREENPSTSGRSQGYGGQSNSWQYCLRLKDKQLAAT LALKCCIGDKLCRSTATYFRQMIAIIAGKRLSFFLLFEIMFGSWYARCVTL KNLNGKOVEAECKEDPEGLALRLAGKGAVSAALEVAESAGLSIDLRREL QGRQLVKLLTTDPLNGGGPAEASRFLSSLQDSADALPVVMGAMQLLPD 10 LRSKQLLILKEFPALRDNNVIMAYAAKAISVTIIPPPREPRITVSASRLROK SRAGPAVKASFTSSLSNFQREARRAFSWAPRNAENRTTSKDVYRKRKNS GLGASERAAWEAMTGIQEDQGSSYSADGQDRLPSVSIAEEWMLTGDKT KDEGVRASHKYESTPDIILFKALLSLCSDELVSARSAMDLCISOMKNVLS 15 SKQLSEGASVETIGRAYHATEAFVQGLSYAKSLLRKLLGTTESTNNNGE RSRDVDDISSDAGSSSVGSQSTDEPSDVLSLTEIWLGRAELLQSLLGSGIS TSLDDIADOLSSECLRDRLISDERYSMAVYMCKKCKIDVFPVWKAWGL ALLRMERYAQARVKFKQAFQLKGEDIPDVIQEIINTIEGGPPVDVSIVRS MYDHLAKSAPTILDDSLSADSYLNVLHMPSTFPRSERSRRSLESEKNSSV

20 PGSDFEDGPRSNLDTTRYSECTNYLQEHARQNLLGFMFRHGHFKDACM LFFPQSGLPPPLQTSSVGAVSTSSSPQRTDPLATEYGTIESLCEFCVGYGAI SSLEEVITERLESAKNQDQAINQYIAGALTRICAFFEINRHFNYLYKFLVL KKDYVTSGYCCIQLFMNSTTQEDAVRHLEHAKKYWSLTILGVQAHFEE ALTARHRGSDSKKLVTKGVRGKSAAEKLSEETLVKLSSRVKMQIDVVK 25 SFSDSEGAPWKHSLFGNPNDSETSRRCEIVETLVEKNFDLAYSVIYEFK

SFSDSEGAPWKHSLFGNPNDSETSRRRCEIVETLVEKNFDLAYSVIYEFK
LSAVDIYAGVATSLADRKKGSQLTELFKNIKGTIQDDDWDQVLNIADTG
KARSVWLIFCEMLQVLGAAINIYANKHKERPDRLIDMLTSSHRKVLACV
VCGRLKSAFQIASKSGSVADVQYVAHQALHANSHTVLDMCKQWLAKY
M

30

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All publications, patents and patent applications are incorporated herein by reference. While in the foregoing specification this invention has been described in relation to certain preferred embodiments thereof, and many details have been set forth for purposes of illustration, it will be apparent to those skilled in the art that the invention is susceptible to additional embodiments and that certain of the

details described herein may be varied considerably without departing from the basic principles of the invention.

#### WHAT IS CLAIMED IS:

- 1. An isolated polynucleotide comprising a plant or fungal nucleotide sequence encoding a polypeptide which is substantially similar to an *Arabidopsis* polypeptide encoded by a nucleic acid sequence comprising an open reading frame selected from the group consisting of SEQ ID NOs: 1-684 and 789-795 or the complement of the open reading frame, wherein the nucleotide sequence is not one of SEQ ID NOs: 1-684 and 789-795.
- 2. An isolated polynucleotide comprising a plant or fungal nucleotide sequence which is substantially similar to an *Arabidopsis* nucleic acid sequence comprising an open reading frame selected from the group consisting of SEQ ID NOs: 1-684 and 789-795, wherein the nucleotide sequence is not one of SEQ ID NOs: 1-684 and 789-795.
- 3. An isolated polynucleotide comprising a plant or fungal nucleotide sequence which hybridizes under high stringency conditions to the complement of a nucleic acid sequence comprising an open reading frame selected from the group consisting of SEQ ID NOs: 1-684 and 789-795, wherein the nucleotide sequence is not one of SEQ ID NOs: 1-684 and 789-795.
- 4. The isolated polynucleotide of claim 1, 2, or 3 which is DNA.
- 5. The isolated polynucleotide of claim 1, 2, or 3 which is RNA.
- 6. The isolated polynucleotide of claim 1 wherein the nucleotide sequence encodes a polypeptide having at least 90% amino acid sequence identity to the *Arabidopsis* polypeptide encoded by the open reading frame.
- 7. The isolated polynucleotide of claim 1 wherein the nucleotide sequence is a maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat sequence.

8. The isolated polynucleotide of claim 1, 2, or 3 wherein the nucleotide sequence is from a dicot.

- 9. The isolated polynucleotide of claim 1, 2, or 3 wherein the nucleotide sequence is from a monocot.
- 10. The isolated polynucleotide of claim 1, 2, or 3 wherein the nucleotide sequence is from a cereal plant.
- 11. An isolated polypeptide encoded by the polynucleotide of claim 1, 2, or 3.
- 12. An expression cassette comprising the polynucleotide of claim 1, 2, or 3 operably linked to suitable regulatory sequences.
- 13. The expression cassette of claim 12 wherein the polynucleotide is linked to a promoter for expression in a plant.
- 14. A recombinant vector comprising the polynucleotide of claim 1, 2, or 3.
- 15. A host cell comprising the expression cassette of claim 12.
- 16. A host cell comprising the polynucleotide of claim 1, 2, or 3.
- 17. The host cell of claim 15 or 16 which is selected from the group consisting of yeast, bacteria and plant.
- 18. A transformed plant, the genome of which is augmented with the polynucleotide of claim 1, 2, or 3 or a polynucleotide which comprises one of SEQ ID NOs: 1-684 and 789-795 which is expressed in an amount which confers resistance or tolerance to the plant to pathogen infection.

- 19. A transformed plant, the genome of which is genetically altered so as to inhibit the expression of a gene corresponding to the polynucleotide of claim 1, 2, or 3 or a gene corresponding to one of SEQ ID NOs: 1-684 and 789-795.
- 20. The plant of claim 19 which is altered by T-DNA insertion, transposon insertion, or targeted DNA insertion.
- 21. The plant of claim 19 in which expression is inhibited by transcription or post-transcriptional mechanisms.
- 22. The plant of claim 18 or 19 which is a monocot.
- 23. The plant of claim 18 or 19 which is a dicot.
- 24. The plant of claim 18 or 19 which is a maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat plant.
- 25. A method of expressing a polynucleotide in a cell, comprising: introducing the polynucleotide of claim 1, 2, or 3 into a cell so as to express the open reading frame.
- 26. The method of claim 25 wherein the cell is a plant cell.
- 27. The method of claim 25 wherein the cell is a monocot cell.
- 28. The method of claim 25 wherein the cell is a dicot cell.
- 29. A composition comprising the polynucleotide of claim 1, 2, or 3 or comprising the polypeptide of claim 11.

30. A method to identify an open reading frame in the genome of a plant cell, the expression of which is altered by compression of the cell, comprising:

- a) contacting a plurality of isolated nucleic acid samples on a solid substrate with a probe comprising plant nucleic acid corresponding to RNA isolated from plant cells infected with an oomycete, so as to form a complex, wherein each sample comprises a plurality of oligonucleotides corresponding to at least a portion of one plant gene; and
- b) comparing complex formation in a) to complex formation between a second plurality of isolated nucleic acid samples on a solid substrate with a second probe comprising plant nucleic acid corresponding to RNA from uninfected plant cells or from mutant plant cells which have a response to pathogen infection that is different that the plant cells in a), so as to identify which samples corresponding to genes having an open reading frame, the expression of which is altered in response to oomycete infection, wherein the genes comprise orthologs of *Arabidopsis* genes comprising one of SEQ ID NOs: 1-684 and 789-795.
- 31. The method of claim 30 further comprising identifying an open reading frame in the gene, the expression of which is induced in response to infection.
- 32. The method of claim 30 further comprising identifying an open reading frame in the gene, the expression of which is decreased in response to infection.
- 33. The method of claim 30 wherein the probe comprises nucleic acid from a dicot.
- 34. The method of claim 30 wherein the probe comprises nucleic acid from a monocot.

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35. An isolated polynucleotide comprising an open reading frame of a gene identified by the method of claim 30.

- 36. A method to confer resistance or tolerance to a plant to a pathogen, comprising:
  - a) contacting plant cells with an expression cassette comprising a polynucleotide encoding a polypeptide that is substantially similar to a polypeptide encoded by a nucleic acid sequence comprising an open reading frame comprising one of SEQ ID NOs: 1-684 and 789-795 so as to yield transformed plant cells; and
  - b) regenerating the transformed plant cells to provide a differentiated transformed plant, wherein the differentiated transformed plant expresses the polynucleotide in the cells of the plant in an amount effective to confer resistance or tolerance to the plant to the pathogen relative to a corresponding plant which does not comprise the expression cassette.
- 37. The method of claim 36 wherein the polynucleotide hybridizes under moderate hybridization conditions to the complement of one of SEQ ID NOs: 1-684 and 789-795.
- 38. The method of claim 36 wherein the polynucleotide hybridizes under stringent hybridization conditions to the complement of one of SEQ ID NOs: 1-684 and 789-795.
- 39. A transformed plant prepared by the method of claim 36.
- 40. A seed of the plant of claim 39.
- 41. A progeny plant of the plant of claim 39.
- 42. A method for identifying a plant cell infected with a pathogen, comprising:

a) contacting isolated nucleic acid obtained from a plant cell suspected of being infected with a pathogen with at least one oligonucleotide under conditions effective to specifically amplify a nucleotide sequence substantially similar to one or more of SEQ ID NOs:1-184, 301-494 or 500-803, or a portion thereof so as to yield an amplified product; and

- b) detecting or determining the presence or amount of the amplified product, wherein the presence or amount of the product is indicative of pathogen infection.
- 43. An isolated polynucleotide comprising a plant nucleotide sequence that alters transcription of an operatively linked nucleic acid segment in a plant cell after pathogen infection, which plant nucleotide sequence is from a gene encoding a polypeptide which is substantially similar to an *Arabidopsis* polypeptide encoded by a gene comprising at least one of SEQ ID NOs: 685-788.
- 44. An isolated polynucleotide comprising a plant nucleotide sequence that directs transcription of an operatively linked nucleic acid segment in a plant cell, which nucleotide sequence hybridizes under high stringency conditions to the complement of at least one of SEQ ID NOs: 685-788 or which nucleotide sequence hybridizes under very high stringency conditions to the complement of at least one of SEQ ID NOs: 685-788.
- 45. An isolated polynucleotide comprising a plant nucleotide sequence that directs transcription of an operatively linked nucleic acid segment in a plant cell, wherein the plant nucleotide sequence comprises SEQ ID NO: 710, SEQ ID NO: 711, SEQ ID NO: 714, SEQ ID NO: 715, SEQ ID NO: 764 or SEQ ID NO: 773.
- 46. The isolated polynucleotide of claim 43, 44, 45 or 46 wherein the nucleotide sequence which directs transcription is 25 to 2000 nucleotides in length.

47. A recombinant vector comprising the polynucleotide of claim 43, 44, 45 or 46.

- 48. The vector of claim 47 which is a plasmid.
- 49. An expression cassette comprising the polynucleotide of claim 43, 44, 45 or 46 operatively linked to an open reading frame.
- 50. The expression cassette of claim 49 operably linked to other suitable regulatory sequences.
- \_\_51. A host cell comprising the expression cassette of claim 49.
  - 52. A transformed plant, the genome of which is augmented with the expression cassette of claim 49.
  - 53. A plant cell containing the expression cassette of claim 49.
  - 54. A transformed plant comprising transformed plant cells, the transformed plant cells containing the expression cassette of claim 49.
  - 55. The transformed plant of claim 54 wherein the plant is a dicot.
  - 56. The cell of claim 53 which is a dicot cell.
  - 57. The transformed plant of claim 54 wherein the plant is a monocot.
  - 58. The cell of claim 53 which is a monocot cell.
  - 59. The transformed plant of claim 54 which is a cereal plant.
  - 60. A method of augmenting a plant genome, comprising:
    - a) contacting plant cells with the expression cassette of claim 49 so
       as to yield transformed plant cells; and

b) regenerating the transformed plant cells to provide a differentiated transformed plant, wherein the differentiated transformed plant expresses the open reading frame in the cells of the plant.

- 61. A transformed plant prepared by the method of claim 60.
- 62. A seed of the plant of claim 61.
- 63. A progeny plant of the plant of claim 61.
- 64. A method of using a plant promoter, comprising: introducing the expression cassette of claim 49 to a plant cell and detecting the expression of the product of the open reading frame.
- 65. A recombinant vector comprising the expression cassette of claim 49.
- 66. A plant cell comprising the vector of claim 65.
- 67. A transformed plant, the cells of which comprise the vector of claim 65.
- 68. The plant of claim 52, 54, 61 or 67 which is a maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat plant.
- 69. The expression cassette of claim 49 wherein the open reading frame is in an antisense orientation.
- 70. The expression cassette of claim 49 wherein the open reading frame is in a sense orientation.
- 71. A transformed plant, the genome of which is augmented with a polynucleotide which is substantially similar to any one of SEQ ID NOs:

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1-684 and 789-795 and which is expressed in an amount which inhibits or prevents pathogen infection.

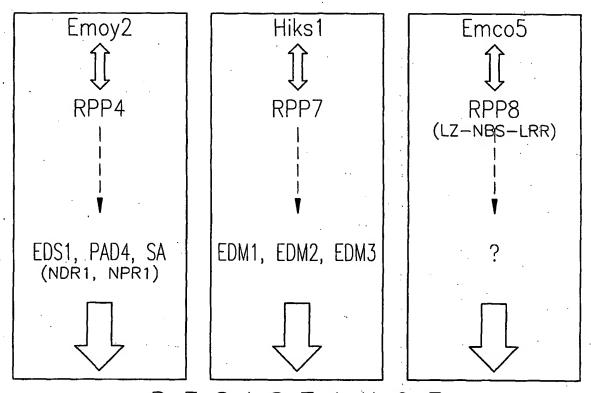
- 72. A transformed plant, the genome of which is genetically altered so as to inhibit the expression of a gene comprising a polynucleotide substantially similar to any one of SEQ ID NOs: 1-684 and 789-795.
- 73. The plant of claim 71 or 72 wherein the polynucleotide is substantially similar to SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 16, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 32, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 47, SEQ ID NO: 48, SEQ ID NO: 792, SEQ ID NO: 57, SEQ ID NO: 68, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 102, SEQ ID NO: 111, SEQ ID NO: 116, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 682, SEQ ID NO: 129, SEQ ID NO: 137, SEQ ID NO: 140, SEQ ID NO: 141, SEQ ID NO: 152, SEQ ID NO: 154, SEQ ID NO: 157, SEQ ID NO: 161, SEQ ID NO: 165, SEQ ID NO: 170, SEQ ID NO: 174, SEQ ID NO: 179, SEQ ID NO: 180, SEQ ID NO: 181, SEQ ID NO: 184, SEQ ID NO: 189, SEQ ID NO: 190, SEQ ID NO: 191, SEQ ID NO: 197, SEQ ID NO: 208 or SEQ ID NO: 211.
- 74. The method of claim 36 wherein the polynucleotide is substantially similar to SEQ ID NO: 308, SEQ ID NO: 300, SEQ ID NO: 272, SEQ ID NO: 362, SEQ ID NO: 265, SEQ ID NO: 241, SEQ ID NO: 261, SEQ ID NO: 380, SEQ ID NO: 228, SEQ ID NO: 658, SEQ ID NO: 243, SEQ ID NO: 254, SEQ ID NO: 216, SEQ ID NO: 225, or one of SEQ ID NO: 400-684.
- 75. The expression cassette of claim 12 wherein the polynucleotide is in antisense orientation.
- 76. The expression cassette of claim 12 wherein the polynucleotide is in sense orientation.

- 77. A method for identifying a plant cell infected with a pathogen, comprising:
  - a) contacting a sample comprising polypeptides obtained from a plant cell suspected of being infected with a pathogen with an agent that specifically binds to a polypeptide that is substantially similar to an Arabidopsis polypeptide encoded by an open reading comprising sequences selected from the group consisting of SEQ ID NOs: 1-684 and 789-795 so as to form a complex; and
  - b) detecting or determining the presence or amount of the complex, wherein the presence or amount of the complex is indicative of pathogen infection.
- 78. A method for identifying a plant cell infected with a pathogen, comprising:
  - a) contacting isolated nucleic acid obtained from a plant cell suspected of being infected with a pathogen with a probe comprising at least a portion of a polynucleotide that is substantially similar to any one of SEQ ID NOs: 1-684 and 789-795 under conditions effective to form a specific complex between the probe and the nucleic acid; and
  - b) detecting and determining the presence or amount of complex formation wherein the presence or amount of complex formation is indicative of pathogen infection.
- 79. A computer-readable medium having stored thereon a data structure comprising:
  - a) a nucleic acid molecule that has at least 70% nucleic acid sequence identity to a nucleotide molecule selected from the group consisting of SEQ ID NOs:1-791 or the complement thereof; and
  - a module receiving the nucleic acid molecule which compares the nucleic acid sequence of the molecule to at least one other nucleic acid sequence.

80. The computer readable medium of claim 79 wherein the medium is selected from the group consisting of magnetic tape, optical disk, CD-ROM, random access memory, volatile memory, non-volatile memory and bubble memory.

- 81. A computer-readable medium having stored thereon computer executable instructions for performing a method comprising:
  - a) receiving a nucleic acid molecule having at least 70% nucleic acid sequence identity to a nucleotide sequence selected from the group consisting of SEQ ID NOs:1-791 or the complement thereof; and
  - b) comparing the nucleic acid sequence of the molecule to at least one other nucleic acid sequence.
- 82. The computer readable medium of claim 81 wherein the medium is selected from the group consisting of magnetic tape, optical disk, CD-ROM, random access memory, volatile memory, non-volatile memory and bubble memory.
- 83. The method of claim 42, 60, 77 or 78 wherein the cells are dicot cells.
- 84. The method of claim 42, 60, 77 or 78 wherein the cells are monocot cells.
- 85. The method of claim 42, 60, 77 or 78 wherein the cells are maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat cells.

### RPP-DEPENDENT DEFENSE PATHWAYS



RESISTANCE

FIG. 1

motifs:

Motif 1/ MAP Score: 6.87857

CAACAATGAC TT TT

Motif 2 MAP Score: 2.22382

GATGGGNCNAA

TT A

C

Known stress-responsive cis-elelemts:

ttgacc, ttgact, ggtcaa and agtcaa W box

ttgaca and tgtcaa W box-like

tgacg and cgtca TGA-bZIP binding site

acgt bZIP binding site core

cacgtg G Box

gccgcc and ggcggc GCC box

| Subcluster A 17014 ribonuclease RNS1

SEQ ID NO:

FIG. 2A

14609 putative cytochrome P450

SEQ ID NO: 775

tttgctcagccttttgtggtctctaccgcttaggaccacttactgcaaatctagaaaat taaaatata<u>tgac</u>tagctaaaatgtgaggtatattgagaagtgagttcttaaaactact tggacttgtctacttagaatttgagacattataagtacaattgatgttaatctaaaagt gtagaacattgttaatttettgaacagetgttage<u>caacaatgae</u>ggaegataegatea ttgatcgaagaggcgattttgtttcacatcgatcagtggatctctttggcgaagtaca<u>tg</u> accaaccaccagatgatcagaagcattcaacacctctctatttatcccaatttgtaagt acatatatgtatgatagtatgtatgtatatattatgagtacatatacattagaaa atactattaactttcaagtcgttatagtaatggacaatattattcgtaattatcaaaag cgctttttcattttctatctgaacctaatgcctagcgagttgaaaacatagctacctaa taggctactactacacaatcttgaagcacaaataatacaacgaaACGTcccttgggtat gagattatttagaagtttcataagattctagtccttattcaattgtgcccatcaagcac aaagtacggaagcgataggaaggACGTCAcagtcctgttttgataatctaatgatcgta gaaagcGTTGGGtgcttttaagttttttctttcgcctaaatattttaaggtcttttgcc aggaatagaaatatgataaagtctattgagtttgtaagacctatgtatattgggtccgg atgcaatgcttttttactcatgcgcattcaatagaagaatttgttatcagtgaacgagt acaaTGTCAAtatatttaacaaattatttaaaaccttttaaaatatgtgcatagtttttt ttcgtactcggatagaaattaataataacaaaaattactgatttgataaaatgaatttc gctttgtaaataaattatatatattcaaaatcctcatcgtcgaaaacaattttaaaaat aaaacaaccaaagatattttccacaaattaaattaattttagttttgaattcagatata ttatttattaatttggatatacttattaccgaaataaaggattgtttatttcttacaac tcgtcttattagtgttgcatagcataacaaggtgaaagagaaaacatq

16649 putative S-adenosyl-L-methionine:trans-caffeoyl-Coenzyme A 3-O-methyltransferase (function:disease SEQ ID NO: resistance) (promoter up to next ORF) 776

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17653 (similarity to DNA damage inducible) (promoter up to next ORF)

SEQ ID NO:

acaataatttccataattcaaatgctttcagaaaggatttctcaatacaggttaaaaat atcaatctaatctttgcaaactcattaaactaatttagttactttccttaactttaaga aacggtctgaaagaggacaaaaagattttcacaagtgtacaccaataaaagaaaatcgt ttttttcccttataaaatgacaaaaattaccaagaaattaaatggaacataacttacga cgaaatttacctgacgattttttttccttactgaaaattaccaagaaatcgcgtggaag aaagtcaaccttttgggaaaattgtcgtctttgacatcagaagagattcaaactaaaat gaatgagtacgttacttgcgttatggttgggttttcctttttaattttagaaacttttc ttacttatattttgaatcaacagttgataacacaagtataaattattattttcgccatc acactggagtcaagctcgagagagacttgtttcctaatttaattcataaacttgtttcg acaatagtcttataatttattcacacttaaaattaacaatacaagtcttttcaa agtcttttaattatataaacaatagaagtctttttcattgtctctttttgactaattaa taaagaccatgaaaagacttgtattgtaagttttgagtgtaaattgtaataaattacta tggttttgttttatatttctgccatctaaataattacattatatgataaaataat taccctagcgacaaatgacagctagaaatgtaataagaacaccaactaaataattatgc aggttaataatggagcaagtgatgttctttctatatactgtacattttcttttaaaaa attatgtaatttcgcagaagagaaaagaaaaacacgtggtgattagagagtagtataaa gatagttggttgggttctgttttttctgtatctcgaggcgccaaaaacaacaacaaa aaaacttcagagcggtgatcagattcaccgatttttctcaaaatq

#### 17008 putative tyrosine aminotransferase

SEQ ID NO: 778

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## 15042 cinnamyl alcohol dehydrogenase-like protein (promoter up to BAC end)

SEQ ID NO: 779

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12778 Lipase/Acylhydrolase with GDSL-motif family SEQ ID NO: (promoter up to next ORF)

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#### || Subcluster B 20245 glutathione-conjugate transporter AtMRP4

SEQ ID NO: 781

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# 17051 CTF2B involved in hydroxylation and oxidation of an aromatic ring

SEQ ID NO: 782

<u>cat</u>agccgcgttccaagaggaGGTCAAgtggattcacttttctataatcaatattaaat tgtagtcacaaaattcaagaaacttatatagaccttattttatttttgaacatcaatag  $\verb|ttgtcgttgaaaagtatcttggggatatttagatttagcatatgA| \verb|GTCAATGTTG| ggtc|$  $\verb"cttaagagttgaggctacaaatttataactcagaaaaagtgtatatgccctttaattt$  ${\tt tggcctattggtcttttatgg} \textbf{CGTCA} {\tt actccaatttcagtttgaaaagaaaTGTCAAta}$ actaacaatgttgtcttTTGACTttgaaatcttagaagtttcgacttacccatttctac aagtgttaaatagcactgttattcaaaatcttatatttgttaattgtgcttaatattt ttaaaatttaaatacttcattaaaatccaatgttattcaaaactaaattacttttgaaa tatgtgtatatgaagtcatttgaaatgaaatctcataagtttcataagtaaattatta gaataaaaacccaaggtaattgtatagattttgaaatccacacaataatatcgaattgt  ${\tt agttagttgattgaatataaaagtaggaaagatgtgggattcatgtggtggtttggccc}$ atctccatgatctctagtgaatttcagctTGACGcaactttcgttaagatcttacttca ttgtgtgttggtgttgagcctcactggattggtgtgtcggctttttagttcactcagag ataattcaaatggatctccttcattccatagttgcatTTGACAtttttatataaaactt  $\verb|gctgattatctaag| teactag| t$  $\verb|atgtctaatagattaaaaattaaaagatagaaatgtctaatggattgaaatatataaag\underline{\textbf{t}}$ gactcagccaaaaacatgttaactgaaaatcttgtataaaatggagattctctgataac

### 19640 putative glutathione S-transferase

SEQ ID NO:

aatgtagttttttctggttttgtggagttaatcactaccttttagcttggatttatagc
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ggcagtgtgatgtaaatgcgaagagtctagaaggaaacagatgTTGACTttacaacata
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ccgcaacaaatagacccacatgtccgttgtactttcctctttttacacaacttcttACG
TgtcgaaatacatgttcacatttaatcatagtggttttttataagaaggatggtTTG

ACTaaattatagcttttatctacttttctttcatatgaaaactccaggtttatgaacca atttcttatgatttgtaagaaattgaatactactacatttaaaatgtttataataatta aTTGACAttagaatttagtaaaaacactaattgttcatacaaatgtttcatttattgtt catacaaactattataacaagattacacaaacatatagttaacatctactgatcgaaga gt<u>atcattgttg</u>tctcttaggctgatttgattctttctacaactttcttcat<u>atq</u>

## 14248 PAD3\_at member of the PF | 00067 Cytochrome P450 family

SEQ ID NO:

gttctggtcgaaggatttgtcccggaatgccaatggcgattgctagtgtggaactagca ttgatgaatttgctttattattttgattggagtatgcctgatgggactaaaggtgaaga cattgatatggaagaagctggtaatatctctatTGTCAAgaaaatacctcttcaacttg tgcctgttcaACGTtattgatgagcaataatataagctcgataacatggaaagtgtatg ataataaatcaagaaagaataacttttgttcatttacttaaaaactagatcataatcaa cacggattaagatcttttcattcaacaaaattctaaattttgttttttaagtaaaaa atgttatagcataaatgttcagatttttatgtcgtattatattqaattaaatataqqac aataagatctagaacaaaaacgatacattcagattgattattgaaatcaaataatcgaa tttataattgaatttgatggaatttataatgtattttttgtcatggaaataataaagtt tggaagatgctggaatgatctctttttacaagaaagctcctctcattcttgttccggtt aaacaccattaatgacatcggcacacaaaaattcaaacaccgttgaagattgtttgcta tactttatttttatcaataataagttgcggtacttggttaactgtatACGTacttctat ttcttatagttgtgcgataattaccacaagtccaaaaaagtaaaatatgaatttgataa agcagtgtaaacctgagattttcaagatttgggcctaaacccattagagaatgctACGT agagaaccattgggcccataaacttattttattcccgcaaagcctgagtggattcgaac cactaccacaaaagtaagagtttgggttatttgattttaatcattcactttgaattat tatagacaccggacagtgacttatgataaagagatttttaacctttaaaactaaaacac taaaatacataatccaaagaagacggaacaaaaaacaaaaacagagccaggaataatq

## 17500 athcallga\_s\_at calmodulin-like protein (promoter up to next ORF)

SEQ ID NO: 785

18928 putative endochitinase (promoter up to SEQ ID NO: next repeat region) 786

<u>cat</u>tttgtgtatggagggtgttgtgtgaagaaatgaagaaggtgtgttgatgtatttat agtgtaaatttggtctattcaaattgaaatattgatcagtgttagacatctttcacgat tattgattggctttttctgagtcaaagtcaatgccactttgtaccacttaacaaaagtc aatgacacagttttttctcttgtcgttatgagaaattccatgtcggtcatagatcagat caaaaacgaatatgtaatcaaaattttaaagagccaagttacttggatgaaaatcattc tcataaaaacttgattgagataatcaagtttagattagttcaaacgtttgagatttttc tattgaatagttccaagtttttgttaattagctcaaacgatattgttagctaatatagc attttttttggtcaactaatatagcatttagaacgttcttttaaagtcttactttgatt tttaacaaaaattgtatggaattgtgagaaaaatgactagtgaagccaaaatgtttcta tttataaatcaaaatcaaccaaaattagaaaaatccaactcaattattgcaatttccaa atagtcatgcgtcaaaattaaaccagcatatcaataaaaagcggtttagtcttaccaac cctggtctagagactctagaaccatcggagctcaagtgaaaaacgacgccgttaagctg catctttcaagatagataaaaacttttgtcgatcgcttcatctctcacagagtttcggc agcgaaactagggtttcgaaagtctctgtctcatcaaagcttcgaaaaggtagatttat ggtttcttactctctctatgatattccaatttcttgcgagcttgctcagtgttc ttaccttatatttaccgaaccagtgcctttatattctatcacattcgaagttattggtt tctggttacttatattgcttattgttctatgactacgatataattatcccaaagatttg ttcttttctccgaatttggattagaaatggatatactcaaattctgaacatgcccagaa gaatcct

14614 putative glucosyltransferase (promoter SEQ ID NO: up to next repeat region) 787

13176 Contains Myb DNA-binding domain repeat signatures

SEQ ID NO: 788

ttccaaaacttaaacctttaaaccttaaatgaacttcaatctacaccatataaagagga  ${\tt agagatataccttcgccatCGTCAttggcccaaagaacaaaatgtacatactttacaag}$  ${\tt aggaatcaatattaaagtgtatataaccaa} \underline{{\tt tgac}} {\tt aaggctccaataatatcttctttgt}$  $\verb|cgttaattggagatcttctgaaca| \verb|tgac| agtgaaggtatacaattgggctagttccaata| \\$ gtcctgaaaaaacacattaccaagttcaacaactcaatcagatcgaaaagtaaacaaca tgaagaagaagaagaacattcacacacaaatgctaacctcgatttcgtttttctgagtt ccaggaatctcaagagcatctacatcgaaagAGTCAAcggcagggctagtgcgaatcaa acgctgctccacattgtcatcctcctcatcggaatccatttccgggtgattatgctcat catatccatcatcgtcgtaatcttcattaacctcataatcatctttctcatcgaacacc catcgcgacaa**TGACGT**cgttgaatcgtctccgccaaactcctcatcaatctcagaccc atcgcctccactgctttcttcttc<u>cACGTg</u>aaacatcaatcaccgttggaaaacactga agatctcgagattgtgattcagattcgtatctctgatccaaggaaacaggattggaatt acctggcacgaatgagaaagaagcttacACGTgtccaatcatgattggattcgagactc acggtttaaggaaaaacaaaccagaccaaattaggcttaaccgctaaaaaaccgggttc tcgttttgaaagattgagagagacgatctacaaaggaggacaggacccggcacgaatga gaagaagcttacACGTgtccaatcaggattgaacgatttaatcaagcttaaccgtatgt aaaccggattttagctgggtccacaagtAGTCAAatatagattttttaatAGTCAAata attttcataggggcgaagttcaagatgagttactacactcatcaaagctcacaaaaaga gaagagaagagaggatcaatcaccattctcatg

26 Peronospora (Hiks1 and Emco5) induced RPP7, EDM1,2,3 and RPP8—Dependent genes

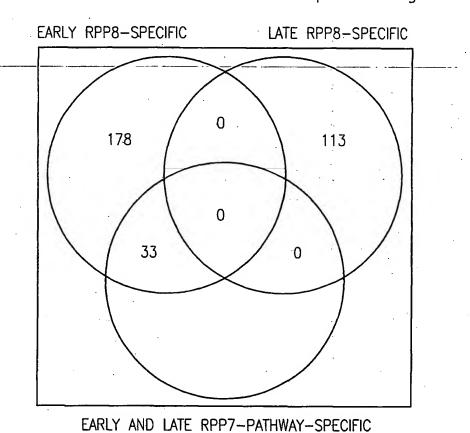
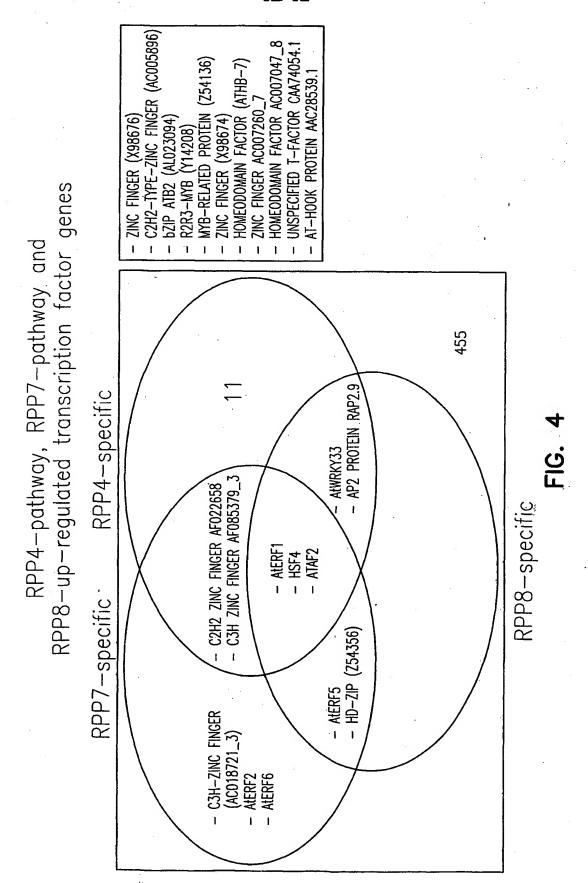


FIG. 3



**SUBSTITUTE SHEET (RULE 26)**